

Commentary

Immunologists getting nervous: neuropeptides, dendritic cells and T cell activation

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(Print ISSN 1465-9921; Online ISSN 1465-993X)**Abstract**

It is increasingly recognised that the immune and nervous systems are closely integrated to optimise defence systems within the lung. In this commentary, the contribution of various neuropeptides such as substance P, calcitonin gene-related peptide, vasoactive intestinal peptide and somatostatin to the regulation of T cell activation is discussed. These neuropeptides are released not only from nerve endings but also from inflammatory immune cells such as monocytes, dendritic cells, eosinophils and mast cells. On release they can exert both direct stimulatory and inhibitory effects on T cell activation and also indirect effects through their influence on the recruitment and activation of professional antigen-presenting dendritic cells. Neuropeptides should therefore be included in the conceptual framework of the immune regulation of T cell function by dendritic cells.

Keywords: calcitonin gene-related peptide, dendritic cells, substance P, T cells, vasoactive intestinal peptide

Introduction

Homeostasis within the body is regulated by three interwoven systems: the endocrine, nervous and immune systems [1]. It is increasingly clear that exchange of information between these systems is facilitated by the endocrine and/or paracrine release of hormones, neuro-mediators and cytokines by either of these systems and by the shared expression of reciprocal receptors for these mediators. As an example, T lymphocytes express neuropeptide receptors for substance P (SP), calcitonin gene-related peptide (CGRP), somatostatin and vasoactive intestinal peptide (VIP). These neuropeptides are released from the unmyelinated nerve endings within the central lymphoid organs and peripheral tissues. At the same time, neural cells express receptors for cytokines, which are released from the immune system in a paracrine fashion and affect neural growth and differentiation. To complicate things further, immune cells themselves can produce

neuropeptides, which influence nervous or immune cells in a paracrine or autocrine fashion.

In this commentary the pivotal role of neuropeptides in the process of T cell activation is discussed against the currently prevailing paradigm of T cell activation by professional antigen-presenting dendritic cells (DCs) [2]. In this paradigm, the first step in the adaptive immune response of the T cell is the recognition and uptake of antigen by immature DCs derived from bone marrow that reside in the periphery of the body and the marginal zone of the spleen, followed by processing of the antigen into an MHC-associated peptide that can be recognised by the T cell receptor (TCR).

DCs are professional antigen-presenting cells for three reasons. First, they express many pattern-recognition receptors for foreign antigen and have the necessary intra-

CGRP = calcitonin gene-related peptide; DC = dendritic cell; IFN- γ = interferon- γ ; IL = interleukin; NANC = nonadrenergic and noncholinergic; R = receptor; SP = substance P; TCR = T cell receptor; VIP = vasoactive intestinal peptide.

cellular enzymes to degrade the antigen into immunogenic peptides. Second, an encounter with 'dangerous' antigens induces the functional maturation of DCs and their migration into the T cell area of draining lymph nodes and spleen, carrying the antigenic cargo into the sites of T cell recirculation. Third, when mature DCs have reached the draining lymph node and spleen, they express co-stimulatory molecules such as CD80, CD86 and intercellular cell-adhesion molecule-1 (ICAM-1) (which are necessary for optimal T cell activation and the avoidance of T cell anergy), and produce cytokines such as interleukin (IL)-12 and IL-10 that critically determine the type of T helper response that is induced [3]. By performing these three essential functions, DCs are the only antigen-presenting cells that can induce a primary immune response after transfer into unimmunized mice, whereas B cells and macrophages fail to do so. Any discussion on the role of neuropeptides on T cell activation should therefore take into account not only the direct effects of these mediators on T cells but also their indirect effects through the modulation of DC function.

Anatomy of interaction of neuropeptides with the immune system

Direct interactions between neuropeptides and immune cells are facilitated by the well-known innervation of both primary (thymus and bone marrow) and secondary (spleen, lymph nodes, Peyer's patches, tonsils) lymphoid organs by capsaicin-sensitive nonadrenergic and noncholinergic (NANC) primary afferent nerve endings and by autonomic nerves containing VIP, somatostatin and neuropeptide Y [4–6]. Within these secondary immune organs, SP and CGRP containing afferent nerve endings of the NANC system terminate around high endothelial venules, the sites of specialised extravasation of recirculating T cells, and in the T cell area and lymphoid follicles, interacting with T cells, macrophages, mast cells and possibly DCs [6,7]. Outside the immune system, the direct interaction of nerve endings containing SP and CGRP with DCs has been described in the skin and in the airway epithelium [8,9]. Within these tissues, the long surface extensions of DCs run parallel to the extensive network of unmyelinated nerve endings, making interaction very likely [10].

Although immunohistochemical staining of thymus, spleen and lymph nodes has demonstrated that neuropeptides such as SP are confined mainly to unmyelinated nerve endings [11], non-neuronal cells of the immune system can be a source of tachykinins [5]. Human T lymphocytes contain preprotachykinin-A mRNA, encoding SP, and produce endogenous SP [12]. Human and rodent monocytes and macrophages produce SP under baseline conditions [13–15]. More importantly, murine DCs derived from bone marrow were shown to contain mRNA for the preprotachykinin A gene, and transcription was confirmed by the demonstration of SP by ELISA and immunohisto-

chemistry [16]. On activation with lipopolysaccharide *in vitro* there was a marked increase in SP expression by mononuclear phagocytes and DCs [13,14,16]. The expression of neuropeptides by these various immune cells could be an explanation of why not all immunoreactivity for neuropeptides is confined to nerve endings within secondary immune organs.

During the effector immune response, the process of lymphocyte migration also allows T cells to migrate into inflammatory lesions within non-lymphoid organs such as the skin, gut, joint and lung. In these sites, lymphocyte extravasation is facilitated by neurogenic inflammation and plasma extravasation that is dependent on the release of SP from capsaicin-sensitive primary afferent nerve endings via an axon reflex. In a mouse model of delayed-type hypersensitivity inflammation of the lung parenchyma, it was shown that SP and VIP were released extensively (nanomolar concentration range) into the lung parenchyma after challenge with sheep erythrocytes in sensitised mice, and closely followed the kinetics of increase in lymphocytes, granulocytes and macrophages in bronchoalveolar lavage fluid as well as the production of cytokines. During the induction of inflammation there was an increase in SP immunoreactive nerve endings within the peribronchial and perivascular leukocytic infiltrates [17]. Not only are inflammatory areas richly supplied by NANC neurons, they also contain many inflammatory immune cells, known to produce neuropeptides (namely macrophages, DCs and lymphocytes). Eosinophils, extracted from *Schistosoma mansoni*-induced liver granulomas, have been shown to produce SP that can influence interferon- γ (IFN- γ) production by intralésional T lymphocytes [18,19]. The same granulomatous lesions also contain immunoreactive somatostatin and VIP [20]. Sites of inflammation within the lung are therefore potentially important areas of interaction between effector immune cells and locally released neuropeptides.

SP as an immunostimulatory neuropeptide

The NANC nervous system acts through neuropeptide mediators such as the tachykinins SP, neurokinin A and neurokinin B. There are at least three distinct tachykinin receptors: neurokinin-1 receptor (NK-1R), NK-2R and NK-3R, which bind preferentially to SP, neurokinin A and neurokinin B, respectively [5]. SP is the most widely studied member of the tachykinin family and modulates a number of important immunological functions, among which are direct effects on T cell activation. Physiological concentrations of exogenously added SP (10^{-11} to 10^{-13} M) augment antigen- and mitogen-induced production of IL-2 [21,22] and proliferation in T lymphocytes *in vitro* and *in vivo* [23–25]. After administration of SP to normal and neonatal capsaicin-treated rats, there was an increase in concanavalin A-induced proliferation of spleen and peripheral blood lymphocytes, which correlated with an enhanced production of IL-2 and expression of the IL-2R,

CD25, on CD4⁺ T cells. Moreover, SP markedly enhanced the percentage of circulating CD25⁺ CD4⁺ T cells in the peripheral blood [26].

Another well-known effect of SP is the stimulation of IFN- γ production by T cells, an effect that could be due to enhanced IL-12 production by antigen-presenting cell types [19,27,28]. With few exceptions, the immunomodulatory effects of SP on lymphocytes can be inhibited by pharmacological antagonists of NK-1R such as the non-peptide antagonist SR140333 [22,26]. Moreover, biochemical and molecular evidence has been obtained that human [12,29,30] and murine lymphocytes [18,31] express NK-1R but not NK-2R or NK-3R. Human monocytes were also shown to contain NK-1R, particularly when obtained from lamina propria of mucosal tissues [32]. Lung and skin DCs also contain binding sites for SP, most probably NK-1R [9].

The demonstration that most immunocytes (monocytes, DCs and lymphocytes) producing SP also express its receptor led to the hypothesis that SP not only acts as a mediator of the crosstalk between the nervous and immune systems but is also biologically involved in the direct interaction between immune cells in a paracrine and/or autocrine fashion, independently of sensory nerves or neurogenic inflammation [12,14,16]. Because DCs are highly involved in the induction and regulation of many immune responses, we have examined the endogenous expression of SP by DCs and studied its role in the activation of T lymphocytes. On co-culture of DCs and allogeneic or syngeneic ovalbumin-specific T cells the addition of a specific NK-1R antagonist, SR140333, led to a decrease in T lymphocyte proliferation induced by DCs, an effect that was enhanced when blocking the co-stimulatory CD80/86-CD28 pathway. These findings were confirmed by the use of responder T cells derived from NK-1R knockout animals, ruling out any toxic effects of SR140333 on the observed effects. Moreover, when purified naive *NK-1R*^{-/-} T cells were stimulated with stimulatory anti-TCR and anti-CD28 antibodies in the absence of DCs, there was a decrease in T cell proliferation, revealing the autocrine release of stimulatory SP by T cells themselves [16]. Indeed, T cells have been shown to transcribe the mRNA for preprotachykinin A and release SP on activation with capsaicin [12].

From a number of experiments, direct autocrine and/or paracrine effects of endogenously released SP on the immunostimulatory capacity of DCs seem less likely, although it has been shown that SP induces activation of the transcription factor nuclear factor- κ B in murine DCs [9,16,33]. This transcription factor was previously shown to be pivotal in the upregulation of stimulatory activity in DCs by upregulating the expression of MHC class II, the co-stimulatory molecules CD86 and CD80, and levels of IL-12 production [34]. One way in which SP might

enhance T cell responses is by recruiting DCs into sites of damage, when it is released very rapidly from nerve endings. SP is a chemoattractant for lung-derived DCs *in vitro* and *in vivo*, and in this way it might stimulate the primary immune response by enhancing immune recognition of dangerous antigens. Moreover, SP is implied in the recruitment of DCs into sites of inflammation during secondary T cell responses in the lung and skin, and its depletion leads to severely reduced delayed hypersensitivity reactions [9]. It is currently unclear how DCs regulate the release and activity of SP during interaction with T cells, but one interesting study demonstrated the presence of aminopeptidase N on the surface of bronchial mucosal L25⁺ DCs in patients with asthma. This enzyme has the potential to break down SP [35].

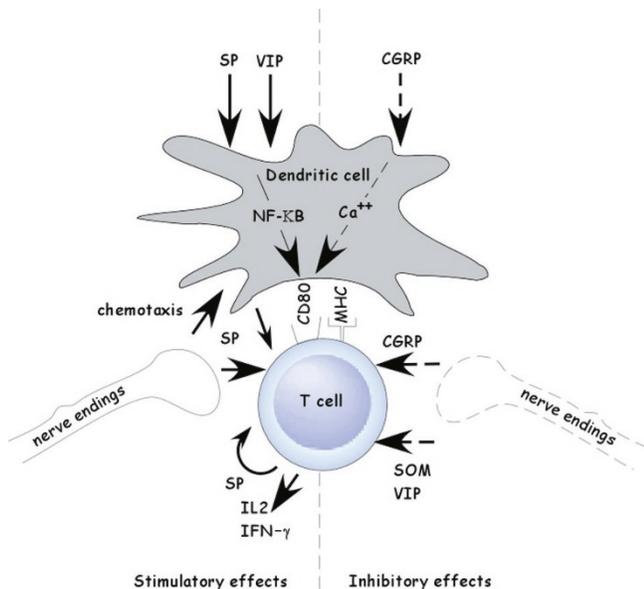
CGRP, somatostatin and VIP as generally suppressive neuropeptides

CGRP is released simultaneously with SP from capsaicin-sensitive nerve endings. In contrast with SP, CGRP directly suppresses IL-2 production and proliferation in murine T cells [36]. In addition, CGRP-containing nerve endings are found in close proximity to skin Langerhans cells, and CGRP has several suppressive effects on DC activation [8,37]. Pretreatment of murine skin DCs with CGRP led to a decrease in alloresponses in the mixed lymphocyte reaction, as well as a decrease in ovalbumin-specific T cell responses of syngeneic T cells [8]. The mechanism by which CGRP mediates its effects on DCs is slowly being discovered. Signalling via the type I CGRP receptor expressed on human monocyte-derived DCs and long-lived murine DC cell lines leads to an increase in intracellular free Ca²⁺ and to a decreased expression of MHC class II, the co-stimulatory molecule CD86, and to a decreased production of IL-12, an effect that could be due to an enhanced production of IL-10 by these DCs [37,38].

Somatostatin is a widespread neuropeptide with generally inhibitory function on hormone release in the anterior pituitary and the gastrointestinal system (for extensive review and references see [39]). In the peripheral nervous system it is found in sympathetic and sensory neurons innervating the lymphoid organs, and receptors for somatostatin are located predominantly in lymphoid follicle germinal centres [6]. Additional non-neuronal sources of somatostatin (such as granuloma cells within *Schistosoma*-induced liver granulomata, lymphocytes, macrophages and thymic DCs) have been described. The presence of binding sites for somatostatin and the expression of mRNA for somatostatin receptors (sstr₁₋₅) on lymphocytes and monocytes is established, although the expression of a particular somatostatin receptor subtype on T lymphocytes seems to vary with species and with the origins of the T cells.

Somatostatin is generally inhibitory for T cell proliferation, especially in the presence of suboptimal stimulatory

Figure 1



Summary of known effects of neuropeptides on the interaction between dendritic cells and T cells. Neuropeptides can be released from nerve endings innervating the primary or secondary lymphoid structures or from nerve endings within inflammatory lesions. Alternatively, dendritic cells and T cells can produce neuropeptides that influence immune activation and/or suppression in an autocrine and paracrine fashion. Solid arrows indicate stimulatory effects; broken arrows indicate inhibitory effects. CGRP, calcitonin gene-related peptide; IFN- γ , interferon- γ ; IL, interleukin; NF- κ B, nuclear factor- κ B; SOM, somatostatin; SP, substance P; VIP, vasoactive intestinal peptide.

conditions. Indeed, the administration of antisense oligodeoxynucleotides designed to block the translation of somatostatin leads to an enhanced spontaneous proliferation of rat splenocytes *in vitro* [40]. In addition, somatostatin suppresses the production of IFN- γ from murine and human T lymphocytes, a finding that has deserved the most attention within the granulomatous inflammation induced by *Schistosoma*, where it generally antagonises the stimulatory effects of SP and vice versa [20,27]. Similarly, granulomata of sarcoidosis patients have been known to bind the somatostatin receptor ligand octreotide *in vivo* [39]. The precise contribution of somatostatin to the immunostimulatory function of DCs remains to be determined, although certain DC subsets contain immunoreactive somatostatin.

VIP and the structurally related pituitary adenylate cyclase-activating polypeptide (PACAP) are present within immune microenvironments and inflammatory pulmonary lesions and modulate a number of T lymphocyte functions ([17]; for review and references see [41]). Macrophages and lymphocytes themselves produce VIP. Receptors for VIP are located predominantly at the CD3⁺

T cell area in lymph nodes and spleen, and include the VPAC₁ receptor (also known as PACAP type II/VIP₁) and VPAC₂ receptor (PACAP type III/VIP₂) [6]. Although VIP has been known to be a suppressive neuropeptide for T cell proliferation and production of IL-2, IL-4 and IL-10, as well as an anti-inflammatory mediator, there are a number of recent studies suggesting that VIP might have a dual role, also enhancing certain lymphocyte functions by interacting with different VIP receptors [42,43]. VIP and PACAP inhibit the activation-induced cell death of activated T cells by inhibiting the expression of Fas ligand, possibly leading to a prolongation of immune responses [44]. The stimulatory effects of VIP on T cell proliferation occurred specifically when stimulated by antigen-pulsed antigen-presenting cells, suggesting an indirect effect of VIP. By signalling through the VPAC₁ receptor, VIP was shown to induce the maturation of immature DCs leading to an enhanced production of IL-12 and an enhanced expression of the DC-maturation marker CD83, especially in the presence of suboptimal amounts of tumour necrosis factor- α [45].

Conclusion and suggestions for the future

Many recent studies have illustrated the importance of immune regulation by neuropeptides through direct effects on T cells and indirect effects on antigen-presenting DCs (see Fig. 1). The accumulated data also suggest that neuropeptides are biologically involved in the direct interaction between immune cells in a paracrine and/or autocrine fashion, independently of sensory nerves. These studies have largely used *in vitro* systems in which concentrations of neuropeptides could be outside the physiological range and in which no account was taken of the normal anatomical distribution of lymphocytes and neural innervation of lymphoid organs and peripheral tissues. Most models that have emerged from these studies have not integrated the effects of neuropeptides with those of cytokines or mediators released from inflammatory cells. For example, it is at present unclear whether and how neuropeptides can modulate such important functions as T helper cell differentiation, tolerance induction and lymphocyte migration. Clearly, studies *in vivo* on the role of neuropeptides will be facilitated by the use of pharmacological antagonists of neuropeptide receptors and of knockout mouse strains lacking particular neuropeptides or their receptors [16,17]. Ultimate proof of the contribution of neuropeptides to human T cell-mediated diseases awaits the results of clinical interventions with the newer and highly selective antagonists of the various neuropeptide receptors.

Note added in proof

Using nested PCR and monoclonal antibodies, it was very recently shown that human and murine DCs express NK-1R (Marriott I, Bost KL: *J Neuroimmunol* 2001, **114**: 131–141).

References

- Goetzl EJ, Sreedharan SP: **Mediators of communication and adaptation in the neuroendocrine and immune systems.** *FASEB J* 1992, **6**:2646-2652.
- Banchereau J, Briere F, Caux C, Davoust J, Lebecque S, Liu Y-J, Pulendran B, Palucka KA: **Immunobiology of dendritic cells.** *Annu Rev Immunol* 2000, **18**:767-811.
- Moser M, Murphy KM: **Dendritic cell regulation of Th1-Th2 development.** *Nat Immunol* 2000, **1**:199-205.
- Lorton D, Bellinger DL, Felten SY, Felten DL: **Substance P innervation of spleen in rats: nerve fibres associated with lymphocytes and macrophages in specific compartments in the spleen.** *Brain Behav Immun* 1991, **5**:29-40.
- Maggi CA: **The effects of tachykinins on inflammatory and immune cells.** *Regul Peptides* 1997, **70**:75-90.
- Reubi JC, Horisberger U, Kappeler A, Laissue JA: **Localization of receptors for vasoactive intestinal peptide, somatostatin and substance P in distinct compartments of human lymphoid organs.** *Blood* 1998, **92**:191-197.
- Weihe E, Hohl R, Michael S: **Molecular anatomy of the neuro-immune connection.** *Int J Neurosci* 1991, **59**:1-23.
- Hosoi J, Murphy GF, Egan CL, Lerner EA, Grabbe S, Asahina A, Granstein RD: **Regulation of Langerhans cell function by nerves containing calcitonin gene-related peptide.** *Nature* 1993, **363**:159-163.
- Kradin R, MacLean J, Duckett S, Schneeberger EE, Waerber C, Pinto C: **Pulmonary response to inhaled antigen: neuro-immune interactions promote the recruitment of dendritic cells to the lung and the cellular immune response to inhaled antigen.** *Am J Pathol* 1997, **150**:1735-1743.
- Evans MJ, Van Winkle LS, Fnucci MV, Hyde D, Plopper CG, Davis CA: **Lateral intercellular space of tracheal epithelium of house dust mite sensitized rhesus monkeys.** *Am J Respir Crit Care Med* 2000, **161**:778.
- Bellinger DL, Lorton D, Romano TD: **Neuropeptide innervation of lymphoid organs.** *Ann NY Acad Sci* 1990, **594**:17-33.
- Lai J, Douglas SD, Ho W: **Human lymphocytes express substance P and its receptor.** *J Neuroimmunol* 1998, **86**:80-86.
- Bost KL, Breeding SAL, Pascual DW: **Modulation of the mRNAs encoding substance P and its receptor in rat macrophages by LPS.** *Regul Immunol* 1992, **4**:105-112.
- Ho W, Lai J, Zhu X, Uvaydova M, Douglas SD: **Human monocytes and macrophages express substance P, neurokinin-1 receptor.** *J Immunol* 1997, **159**:5654-5660.
- Germonpre PR, Bullock GR, Lambrecht BN, Van de Velde V, Luyten WHML, Joos GF, Pauwels RA: **Presence of substance P and neurokinin 1 receptors in human sputum macrophages and U-937 cells.** *Eur Resp J* 1999, **14**:776-782.
- Lambrech BN, Germonpre PR, Everaert EG, Carro-Muino I, De Veerman M, De Felipe C, Hunt SP, Thielemans K, Joos GF, Pauwels RA: **Endogenously produced substance P contributes to lymphocyte proliferation induced by dendritic cells and direct TCR ligation.** *Eur J Immunol* 1999, **29**:3815-3825.
- Kaltreider HB, Ichikawa S, Byrd PK, Ingram DA, Kishiyama JL, Sreedharan SP, Warnock ML, Beck JM, Goetzl EJ: **Upregulation of neuropeptides and neuropeptide receptors in a murine model of immune inflammation in lung parenchyma.** *Am J Respir Cell Mol Biol* 1997, **16**:133-144.
- Cook GA, Elliott D, Metwali A, Blum AM, Sandor M, Lynch R, Weinstock JV: **Molecular evidence that granuloma T lymphocytes in murine Schistosomiasis mansoni express an authentic substance P (NK-1) receptor.** *J Immunol* 1994, **152**:1830-1835.
- Blum AM, Metwali A, Cook G, Mathew RC, Elliott D, Weinstock JV: **Substance P modulates antigen-induced, IFN- γ production in murine Schistosomiasis mansoni.** *J Immunol* 1993, **151**:225-233.
- Blum AM, Metwali A, Mathew RC, Cook G, Elliott D, Weinstock J: **Granuloma T lymphocytes in murine schistosomiasis mansoni have somatostatin receptors and respond to somatostatin with decreased IFN- γ secretion.** *J Immunol* 1992, **149**:3621.
- Rameshwar P, Gascon P, Ganea D: **Stimulation of IL-2 production in murine lymphocytes by substance-P and related tachykinins.** *J Immunol* 1993, **151**:2484-2496.
- Calvo C, Chavanel G, Senik A: **Substance P enhances IL-2 expression in activated human T cells.** *J Immunol* 1992, **148**:3498-3504.
- Payan DG, Brewster DR, Goetzl EJ: **Specific stimulation of human T lymphocytes by substance P.** *J Immunol* 1983, **131**:1613-1615.
- Nio DA, Moylan RN, Roche JK: **Modulation of T lymphocyte function by neuropeptides. Evidence for their role as local immunoregulatory elements.** *J Immunol* 1993, **150**:5281-5288.
- Scicchitano R, Bienenstock J, Stanis AM: **In vivo immunoregulation by the neuropeptide substance P.** *Immunology* 1988, **63**:733-735.
- Santoni G, Perfumi MC, Spreghini E, Romagnoli S, Piccoli M: **Neurokinin type-1 receptor antagonist inhibits enhancement of T cell functions by substance P in normal and neuromanipulated capsaicin-treated rats.** *J Neuroimmunol* 1999, **93**:15-25.
- Blum AM, Metwali A, Mathew RC, Elliott D, Weinstock JV: **Substance P and somatostatin can modulate the amount of IgG2a secreted in response to schistosome egg antigens in murine Schistosomiasis mansoni.** *J Immunol* 1993, **151**:6994-7004.
- Kincy-Cain T, Bost KL: **Substance P-induced IL-12 production by murine macrophages.** *J Immunol* 1997, **158**:2334.
- Goode T, O'Connell J, Sternini C, Anton P, Wong H, O'Sullivan GC, Collins JK, Shanahan F: **Substance P (neurokinin-1) receptor is a marker of human mucosal but not peripheral mononuclear cells: molecular quantitation and localization.** *J Immunol* 1998, **161**:2232-2240.
- Payan DG, Brewster DR, Missirian-Bastian A, Goetzl EJ: **Substance P recognition by a subset of human T lymphocytes.** *J Clin Invest* 1989, **74**:1532-1539.
- Stanisz AM, Scicchitano R, Dazin P, Bienenstock J, Payan DG: **Distribution of substance P receptors on murine spleen and Peyer's patch T and B cells.** *J Immunol* 1987, **139**:749-754.
- Goode T, O'Connell J, Ho W, O'Sullivan GC, Collins JK, Douglas SD, Shanahan F: **Differential expression of neurokinin-1 receptor by human mucosal and peripheral lymphoid cells.** *Clin Diagn Lab Immunol* 2000, **7**:371-376.
- Marriott I, Mason MJ, Elhofy A, Bost KL: **Substance P activates NF- κ B independent of elevations in intracellular calcium in murine macrophages and dendritic cells.** *J Neuroimmunol* 2000, **102**:163-171.
- Grohmann U, Belladonna ML, Bianchi R, Orabona C, Ayroldi E, Fioretti MC, Puccetti P: **IL-12 acts directly on DC to promote nuclear localization of NF- κ B and primes DC for IL-12 production.** *Immunity* 1998, **9**:315-323.
- Van der Velden VH, Wierenga-Wolf AF, Adriaansen-Soeting PW, Overbeek SE, Moller GM, Hoogsteden HC, Versnel MA: **Expression of aminopeptidase N and dipeptidyl peptidase IV in the healthy and asthmatic bronchus.** *Clin Exp Allergy* 1998, **28**:110-120.
- Wang F, Miller I, Bottomly K, Vignery A: **Calcitonin gene-related peptide inhibits interleukin 2 production by murine T lymphocytes.** *J Biol Chem* 1992, **267**:21052-21057.
- Carucci JA, Ignatius R, Wei Y, Cypess AM, Schaer DA, Pope M, Steinman RM, Mojsov S: **Calcitonin gene-related peptide decreases expression of HLA-DR and CD86 by human dendritic cells and dampens dendritic cell-driven T cell-proliferative responses via the type I calcitonin gene-related peptide receptor.** *J Immunol* 2000, **164**:3494-3499.
- Torii H, Hosoi J, Beissert S, Xu S, Fox FE, Asahina A, Takashima A, Rook AH, Granstein RD: **Regulation of cytokine expression in macrophages and the Langerhans cell-like line XS52 by calcitonin gene-related peptide.** *J Leukocyt Biol* 1997, **61**:216-223.
- ten Bokum AMC, Hofland LJ and van Hagen PM: **Somatostatin and somatostatin receptors in the immune system: a review.** *Eur Cytokine Netw* 2000, **11**:161-176.
- Aguila MC, Rodriguez AM, Aguila-Mansilla HN, Lee WT: **Somatostatin antisense oligodeoxynucleotide-mediated stimulation of lymphocyte proliferation in culture.** *Endocrinology* 1996, **137**:1585-1590.
- Pozo D, Delgado M, Martinez C, Leceta J, Gomariz RP, Guerrero JM, Calvo JR: **Immunobiology of VIP.** *Immunol Today* 2000, **21**:7.
- Ganea D, Sun L: **VIP downregulates the production of IL-2 but not of IFN- γ from stimulated murine T lymphocytes.** *J Neuroimmunol* 1993, **47**:147-155.
- Taylor AW, Streilein JW, Cousins SW: **Immunoreactive VIP contributes to the immunosuppressive activity of normal aqueous humor.** *J Immunol* 1994, **153**:1080-1086.

44. Delgado M, Ganea D: **Vasoactive intestinal peptide and pituitary adenylate cyclase-activating polypeptide inhibit expression of Fas ligand in activated T lymphocytes by regulating c-Myc, NF- κ B, NF-AT, and early growth factors2/3.** *J Immunol* 2001, **166**:1028–1040.
45. Delneste Y, Herbault N, Galea B, Magistrelli G, Bazin I, Bonnefoy JY, Jeannin P: **Vasoactive intestinal polypeptide synergizes with TNF- α in inducing human dendritic cell maturation.** *J Immunol* 1999, **163**:3071–3075.