

Research

Open Access

## Hypoxia and dehydroepiandrosterone in old age: a mouse survival study

Edouard H Debonneuil<sup>\*1</sup>, Janine Quillard<sup>2</sup> and Etienne-Emile Baulieu<sup>1</sup>

Address: <sup>1</sup>Institut National de la Santé et de la Recherche Médicale, Unité Mixte de Recherche 788, Pincus Building, 80 rue du Général Leclerc, 94276 Le Kremlin-Bicêtre Cedex, France and <sup>2</sup>Service d'Anatomo-Pathologie, Hôpital de Bicêtre, Assistance Publique-Hôpitaux de Paris, Le Kremlin-Bicêtre, France

Email: Edouard H Debonneuil<sup>\*</sup> - edebonneuil@yahoo.fr; Janine Quillard - jeanine.quillard@bct.aphp.fr; Etienne-Emile Baulieu - baulieu@kb.inserm.fr

<sup>\*</sup> Corresponding author

Published: 18 December 2006

Received: 12 May 2006

Respiratory Research 2006, 7:144 doi:10.1186/1465-9921-7-144

Accepted: 18 December 2006

This article is available from: <http://respiratory-research.com/content/7/1/144>

© 2006 Debonneuil et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

**Background:** Survival remains an issue in pulmonary hypertension, a chronic disorder that often affects aged human adults. In young adult mice and rats, chronic 50% hypoxia (11% FIO<sub>2</sub> or 0.5 atm) induces pulmonary hypertension without threatening life. In this framework, oral dehydroepiandrosterone was recently shown to prevent and reverse pulmonary hypertension in rats within a few weeks. To evaluate dehydroepiandrosterone therapy more globally, in the long term and in old age, we investigated whether hypoxia decreases lifespan and whether dehydroepiandrosterone improves survival under hypoxia.

**Methods:** 240 C57BL/6 mice were treated, from the age of 21 months until death, by normobaric hypoxia (11% FIO<sub>2</sub>) or normoxia, both with and without dehydroepiandrosterone sulfate (25 mg/kg in drinking water) (4 groups, N = 60). Survival, pulmonary artery and heart remodeling, weight and blood patterns were assessed.

**Results:** In normoxia, control mice reached the median age of 27 months (median survival: 184 days). Hypoxia not only induced cardiopulmonary remodeling and polycythemia in old animals but also induced severe weight loss, trembling behavior and high mortality ( $p < 0.001$ , median survival: 38 days). Under hypoxia however, dehydroepiandrosterone not only significantly reduced cardiopulmonary remodeling but also remarkably extended survival ( $p < 0.01$ , median survival: 126 days). Weight loss and trembling behavior at least partially remained, and polycythemia completely, the latter possibly favorably participating in blood oxygenation. Interestingly, at the dose used, dehydroepiandrosterone sulfate was detrimental to long-term survival in normoxia ( $p < 0.05$ , median survival: 147 days).

**Conclusion:** Dehydroepiandrosterone globally reduced what may be called an age-related frailty induced by hypoxic pulmonary hypertension. This interestingly recalls an inverse correlation found in the prospective PAQUID epidemiological study, between dehydroepiandrosterone blood levels and mortality in aged human smokers and former smokers.

## Background

In human beings, pulmonary hypertension (PH) is a chronic and life threatening disorder in which a progressive increase of pulmonary vascular resistance leads to right ventricular failure. When detected, PH is often an already irreversible chronic pathology and leads to death after several years of severe illness and treatment [1-5]. Among various etiologies, PH often develops in aged smokers with hypoxemia associated with chronic obstructive pulmonary disease (COPD) [6-10]: in these cases survival can be extended by long-term oxygenotherapy [9-13].

Therapies under development may be studied in rats and mice by their effects on pulmonary arterial pressure or cardiopulmonary remodeling. Survival has been studied in rats with the use of monocrotaline injection to model PH [14-17], but the multiple disorders caused [18-20] and the brief period over which deaths are recorded [14-17] bias long-term PH survival analysis. In fact, PH may not be deadly in itself: young adult mice and rats survive and develop stable PH within 3 weeks of 50% hypoxia (11% FIO<sub>2</sub> or 0.5 atm) ([21-24], plus recurrent personal observation), and it was recently shown in rats that if hypoxia (0.5 atm) is maintained death does not occur until the rats are aged [25]. Since heart failure does occur in human PH, this brings into question today's development of PH therapies and their specific long-term global effects in laboratory animals.

Therefore we decided to use hypoxia, up to death in mice, starting at an age when they naturally start dying, in order to evaluate long-term positive or negative survival effects of hypoxic PH and a potential therapy. We considered dehydroepiandrosterone (DHEA), that has recently been shown to prevent and treat chronic hypoxic PH in rats when administered orally in its free (30 mg/kg every other day, 0.5 atm, [23]) or sulfate form (DHEAS; 9 mg/kg/day in drinking water, 11% FIO<sub>2</sub>, [24]; after oral ingestion most if not all the sulfate is converted into the free form).

Hypoxic pulmonary vasoconstriction helps oxygenating the blood but increases pulmonary arterial pressure. By relaxing contracted pulmonary arteries [23,26,27], DHEA inhibits both phenomena. Like any vasodilator it may therefore treat PH without being beneficial to the patient. Survival of aged mice will be our indicator of potential benefits. The old age is moreover of interest both because in humans PH complicating COPD often concerns aged persons [2-13] and because aged persons have lower blood DHEA(S) levels [28].

## Methods

### Conditions

Mice were obtained at the age of 17 months (240 C57BL/6 males from Elevage Janvier, Le Genest-St-Isle, France) and randomly distributed into 4 groups (N = 60) in cages containing 7 to 9 mice each with ad libitum standard diet (M20, Special Diet Services Ltd., Witham, Essex, UK) and water. At the age of 21 months – which we will refer to as t = 0 – each group received a different environmental condition, defined by a combination of hypoxia or normoxia and DHEA or not. Cages were changed weekly and food and drink renewed every other week. All procedures concerning animal care and use were carried out in accordance with the European Community Council Directive (86/609/EEC). All animal procedures were approved by the animal care and use committee at the institute. All treatments and measures were performed by investigators blinded to the treatment.

We chose normobaric hypoxia (11% FIO<sub>2</sub>) to avoid potential harmful consequences of rapid pressure variations. Hypoxic mice were housed in a home-made chamber homogeneously supplied by a flow of a filtered mixture of air and nitrogen (provided by a nitrogen generator from Air Liquide, Paris, France) at ambient pressure and 11 ± 1% oxygen (controlled by a ProOx controller from Biospherix, New York City, NY, USA). Control normoxic mice were housed in a similar chamber supplied by a flow of filtered air. Gas flowed sufficiently fast (15 l/min) into the chambers to ensure low carbonic gas levels (less than 0.05%). Hypoxia was interrupted weekly for roughly one hour for animal care.

DHEAS (Steraloids, Newport, RI, USA) was incorporated at 0.25 mg/ml (0.1 mg/ml gave partial results in rats, [24]) into the drinking water, except during the first two weeks where 0.1 mg/ml was used to allow taste habituation [29,30].

### Measurements

Survival was checked every one to three days until t = 180 days (when most animals had died in all groups). From time to time mice were weighed and their food and drink consumption was approximated by giving 350 g food and 500 ml drink per cage and measuring how much remained one week later.

Cardiopulmonary remodeling was measured in mice that died before t = 90 days (kept at -20°C when found – usually up to one day after death – up to analysis). Right ventricular hypertrophy was assessed by the right ventricle to left ventricle plus septum weight ratio (RV/LV+S) [23]. Lungs were formalin-fixed for histological study and pulmonary artery remodeling was expressed as percentage vessel wall thickness (100 × (external diameter-internal

diameter)/external diameter, measured on a computer screen) in small and medium-sized pulmonary arteries (80–150  $\mu\text{m}$ ), averaged over 10 pulmonary arteries per mouse [23].

Blood sampling was performed on one initially randomly chosen cage per group. Additional cages were randomly chosen if needed to have at least 5 mice tested per group. The mice to be tested were placed in clean cages with their usual drink but no food overnight, and were excluded from survival analysis. Blood sampling (300  $\mu\text{l}$ ) was performed retro-orbitally under inhaled isoflurane anesthesia, in the morning. Blood was mixed with 10% ethylenediaminetetraacetic acid at 0.5 M. A blood analyzer (ABC-Animal Blood Counter, Scil, Viernheim, Germany) provided hematocrit, hemoglobin content, and the count, volume and hemoglobin concentration of red blood cells.

### Statistics

Values are expressed as mean  $\pm$  SEM. Statistics were performed with JMP 6.0 (SAS Institute, Cary, NC, USA). Comparisons between two and several groups were done by Student and one-way ANOVA tests, respectively. Survival curve characteristics and comparisons were based on the proportional hazards Cox model. The method for choosing the number of animals is provided in an online additional file [see Additional file 1].

## Results

### Survival

Survival is clearly the main global health indicator. Note that mortality may affect the significance of results by death selection.

#### *Before treatment: low mortality*

It is rather unusual to start lifespan experiments with animals that are already aged. We wanted to start treatment (hypoxia or normoxia and DHEA or not) when the rate of 'natural death' becomes significant in C57BL/6 laboratory male mice. Starting with mice that are too old would imply that selection by death has commenced and that only resistant mice are being studied. If the mice are too young then in the short term no natural death will occur and any survival improvement due to a therapy may not be detected.

It appeared from the literature [31,32] that the appropriate starting age was 20 months. In fact our mice survived better than expected and we decided to start the treatments at the age of 21 months, with 5 deaths (plus 13 following arrival) instead of 20 or 25 as expected by extrapolating the literature. The results were then considered in terms of two 3-month time periods.

#### *First 3 month period of treatment: dehydroepiandrosterone reduces a drastic age-specific hypoxic mortality*

Survival curves are shown in Figure 1 ( $t = 0$  to 91 days; 21 to 24 month old mice) and relative risks of death for that period are shown in Figure 2a. Control mice – normoxia without DHEA – had a higher death rate than before the age of 21 months but there was still 89% survival at 24 months (compared to an expected ~70% from the literature). DHEA did not affect survival under normoxia (82% survival, relative risk of death: 1.24,  $p = 0.40$ ). However, for hypoxic mice – without DHEA – the death rate increased drastically between  $t = 20$  and  $t = 40$  days, leading to only 48% survival, and then they died at a lower rate, leading to 39% survival at 24 months (relative risk of death: 2.73,  $p < 0.001$  compared to control). Under hypoxia, DHEA led to 61% survival at 24 months with a roughly constant death rate: this treatment improved survival of hypoxic mice (relative risk of death: 0.68,  $p = 0.0065$ ) while the normoxic survival level was not reached (relative risk of death: 1.62,  $p < 0.013$ ).

#### *Second 3 months of treatment: various age-related deaths*

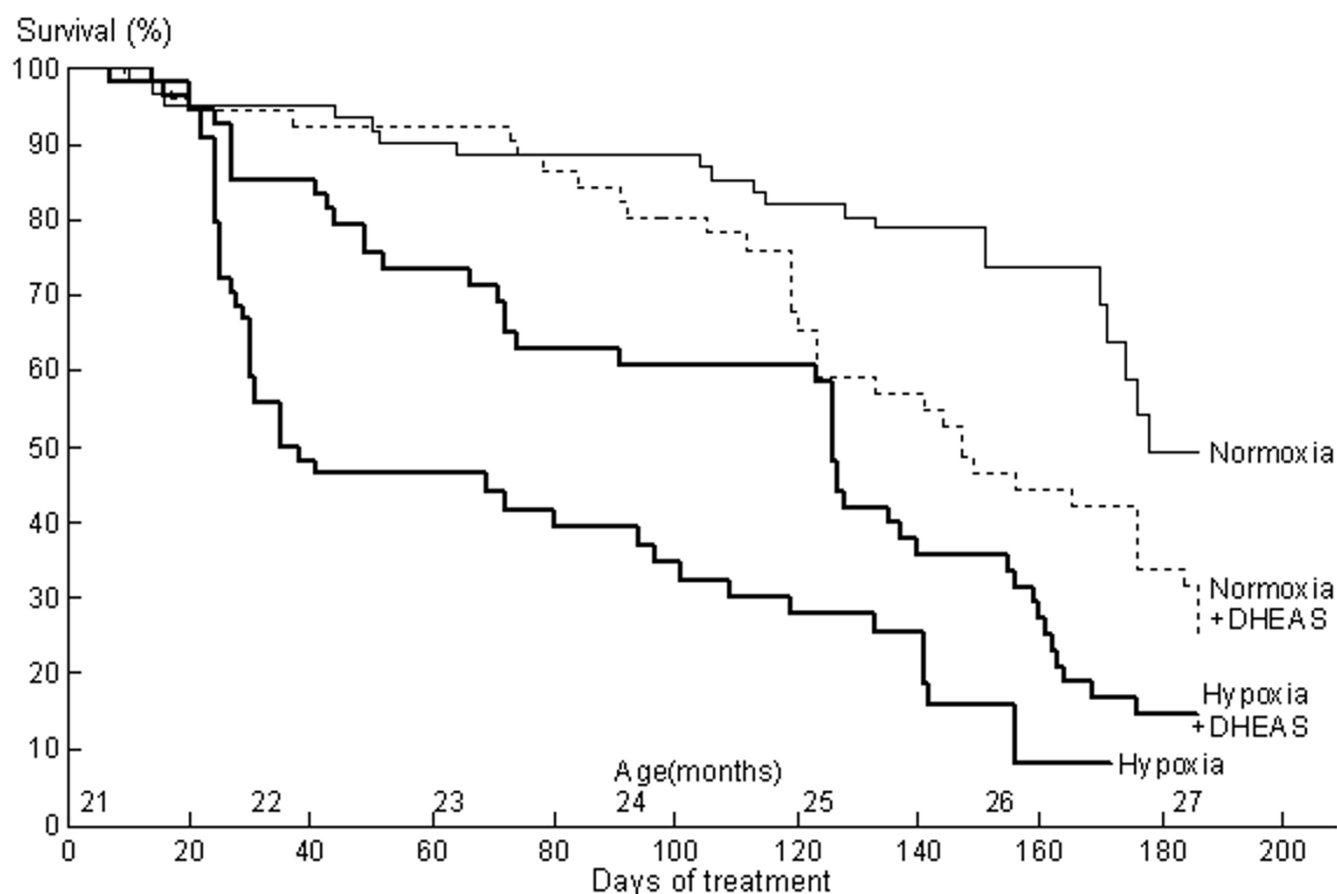
Over the next 3 months of treatment (24 to 27 month old mice,  $t = 92$  to 183 days, figures 1 and 2c), mortality largely increased in all groups. Under normoxia, the control group reached 75% and 50% survival at 26 and 27 months (24 and 26 months would have been expected from the literature), and fewer died than in the 3 other groups (relative risk of death: 0.66, 0.43, and 0.57;  $p = 0.014$ ,  $p < 0.001$  and  $p = 0.0014$ ; compared to DHEA, hypoxia, and hypoxia+DHEA, respectively). The only statistical difference among the 3 groups was that normoxic mice with DHEA had a lower death rate than hypoxic mice without DHEA ( $p = 0.05$ ).

#### *In summary*

Over the 6 months of treatment (21 to 27 month-old mice,  $t = 0$  to 186 days, figure 1 and 2b) hypoxia induced a much higher mortality (median survival: 38 days, relative risk of death: 2.53,  $p < 0.001$ ) than for control animals (mean survival: 184 days). DHEA globally improved survival under hypoxia (median survival: 126 days, relative risk of death: 0.72,  $p = 0.0025$ ) but reduced it under normoxia (median survival: 126 days, relative risk of death: 1.39,  $p = 0.0025$ ), compared with the corresponding untreated group.

#### *Cardiopulmonary remodeling*

After death, PH can be diagnosed by the consequential increase in pulmonary artery wall thickness and enlarged right ventricle. We assessed cardiopulmonary remodeling in mice that died before  $t = 91$  days (analysis of later deaths would lead to complex interpretations because of previous death selection and multiple age-related pathologies). Pulmonary artery remodeling (percentage vessel



**Figure 1**

**Survival.** Survival of 21-month-old male C57BL/6 mice under hypoxia or normoxia (thick or thin lines), with or without dehydroepiandrosterone (dashed or solid lines). Hypoxia induced a high mortality. Dehydroepiandrosterone sulfate (DHEAS) prevented it, despite detrimental effects perceived in normoxia, at the oral sulfate dose used.

wall thickness) is shown in figure 3A (typical micrographs in figure 4) and heart remodeling (RV/LV+S percentage) in figure 3B.

Compared to the control group, hypoxic mice had higher pulmonary artery (38 vs 23;  $p = 0.01$ ) and heart (0.325 versus 0.287;  $p = 0.05$ ) remodeling. DHEA had no effect on the normoxic cardiopulmonary system but under hypoxia DHEA significantly reduced pulmonary artery and heart remodeling (29 vs 38;  $p < 0.05$  and 0.286 versus 0.325;  $p < 0.05$ ).

#### Food and drink consumption

Overall, the mean daily consumption was of  $3.0 \pm 1$  g and  $3.25 \pm 0.28$  ml per mouse, with no particular distinction over groups and time. The consumption may have been lower because the determination did not take into

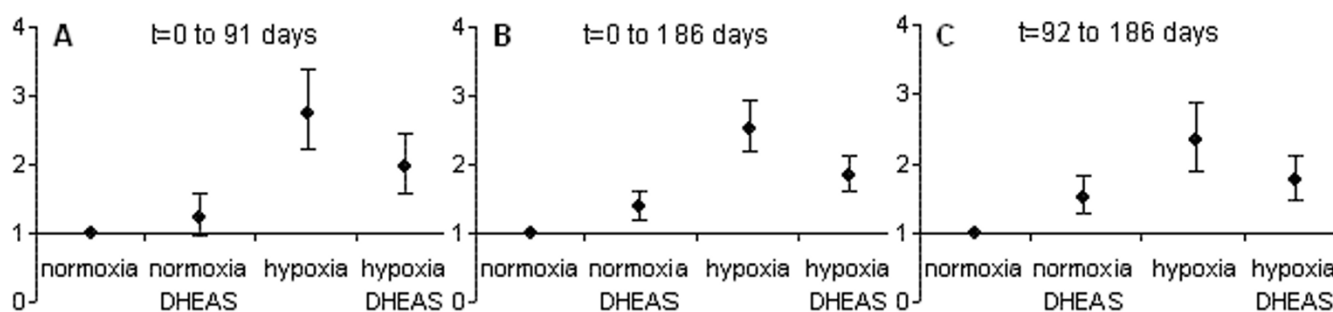
account food and drink remaining at the bottom of the cage, which depend on the number of mice per cage, on their activity and on cage manipulation during the week. For DHEA-treated mice weighing ~30 g, we estimate that the DHEAS consumption was on the order of 25 mg/kg/day.

#### Body weight

Sick mice generally lose weight and as such body weight (figure 5) may be used as an overall evaluation of the state of health.

#### Before treatment

When the mice arrived, we observed that they were thin (the mice had a similar diet before arrival, so the weight loss is probably due to the stress of transportation). The mice regained normal appearance within a month. When



**Figure 2**

**Relative risk of death.** Relative risk of death taken from Figure 1, with normoxia as a reference and at time intervals: (A)  $t = 0$  to 91 days (B)  $t = 0$  to 186 days (C)  $t = 92$  to 186 days. Despite temporary mortality patterns hypoxia and dehydroepiandrosterone (DHEAS) appear to globally have similar effects on survival at the three intervals.

measured two months before treatment, all groups had similar weights ( $27.9 \pm 0.12$  g) and food and drink consumption.

#### Normoxic animals

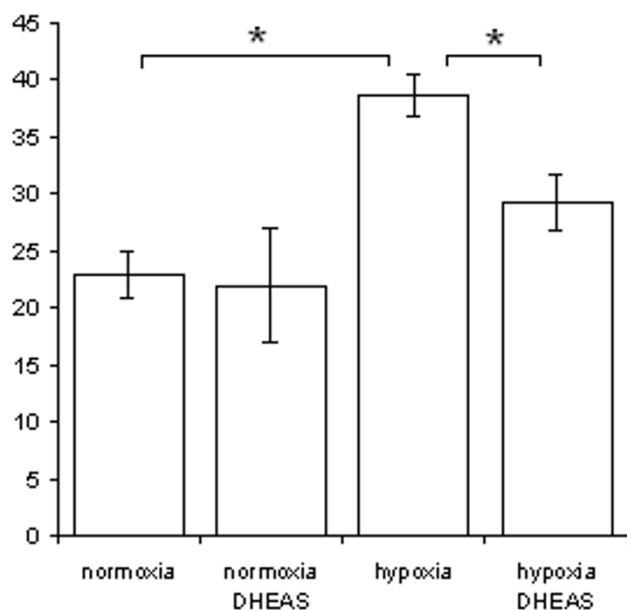
Weights of control mice gradually increased until the age of 25 months (by  $0.64$  g/month, reaching  $32.6 \pm 0.1$  g at  $t = 120$  days, figure 5). This is probably a long increase towards a higher equilibrium weight long after the transportation weight loss (similar long-term weight changes are observed after changing diets [32]). The weight then

slightly (but not significantly) decreased on average (figure 5), which may reflect negative selection of heavy animals by death. DHEA-treated normoxic mice also gained weight but to a lower extent (by  $0.42$  g/month up to  $t = 120$  days), weighing slightly but significantly less ( $p \sim 0.007$ ) than control mice at  $t = 30, 60$  and  $120$  days.

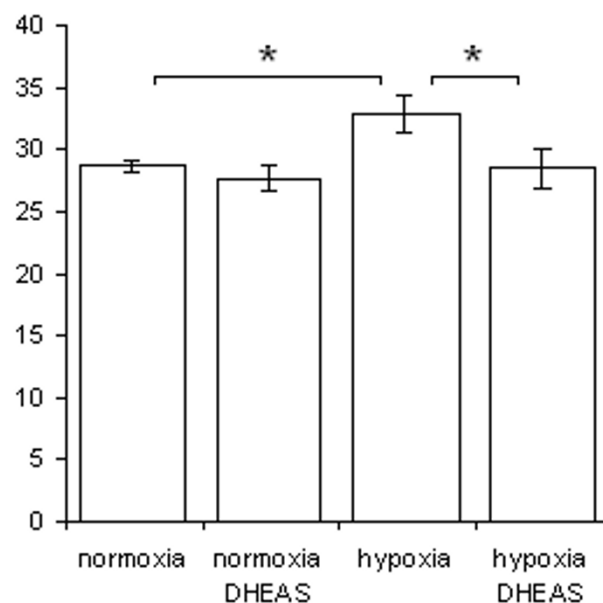
#### Hypoxic animals: temporary weight loss and trembling behavior

After two weeks of hypoxia, all aged mice, with and without DHEA, were particularly thin and for many, if not all of them, normal cage behavior was interrupted by periods

#### A Percentage vessel wall thickness

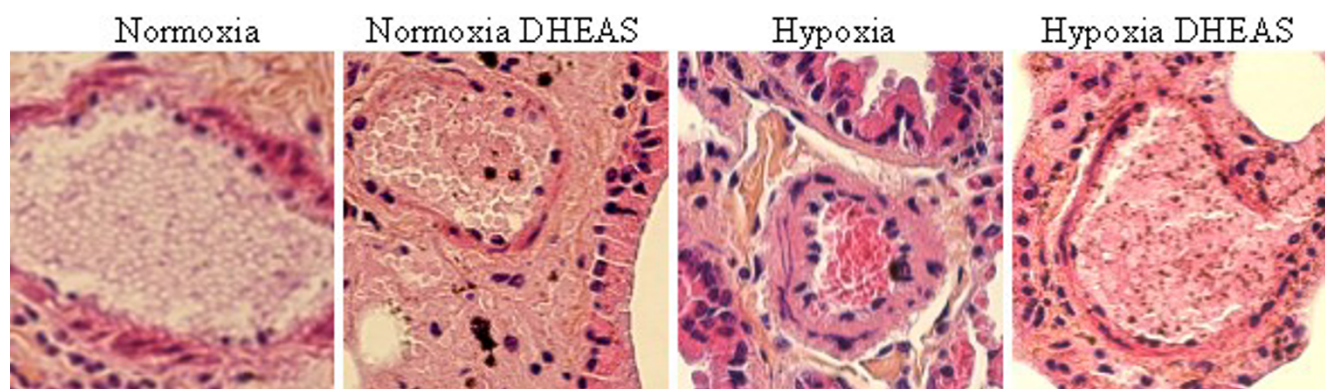


#### B Percent RV/(LV+S)



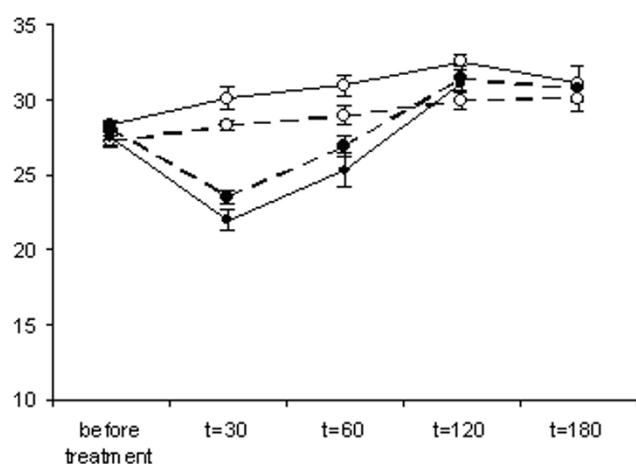
**Figure 3**

**Cardiopulmonary remodeling.** (A) Pulmonary artery remodeling (B) Heart remodeling in mice dead between  $t = 0$  and 91 days. Hypoxia induced cardiopulmonary remodeling and dehydroepiandrosterone (named DHEAS in the figure) prevented it (\*:  $p < 0.05$ ).

**Figure 4**

**Pulmonary artery sections.** Typical pictures of pulmonary arteries from mice under different conditions (image width: 150  $\mu$ m). Hypoxic mice without dehydroepiandrosterone (DHEAS) have a thicker vessel wall with respect to diameter.

of trembling while curling up. When measured after one month of treatment, the weight of hypoxic mice was indeed much lower than their normoxic counterparts ( $22 \pm 0.7$  g versus  $30.1 \pm 0.8$  g,  $p < 0.001$ ). After two or three months, the -remaining- mice regained normal size (and normal weight, figure 5) and trembling behavior became rare. The trembling behavior also occurred with DHEA. For weight, DHEA did not obviously reduce the hypoxic weight loss ( $23.6 \pm 0.5$  g versus  $22 \pm 0.7$  g,  $p = 0.11$  at  $t = 30$  days), but an already large selection by death in the hypoxic group without DHEA could mask the difference.

**Figure 5**

**Weight.** Weight of mice, two months before and after 30, 60, 120 and 180 days of treatment, under normoxia (empty circles) or hypoxia (filled circles), with (dotted line) or without (continuous line) dehydroepiandrosterone in their drinking water. Hypoxia induced a temporary weight loss, with and without dehydroepiandrosterone (\*:  $p < 0.05$ ) (it may be assumed that all the  $t = 0$  points should coincide).

### Hematocrit

The evolution of the hematocrit among groups is shown in figure 6 and other blood parameters in table 1. Hypoxia typically induces polycythemia which may compensate for the lack of oxygen [33] and is characterized by a high hematocrit.

One month before treatment, all groups had a similar hematocrit (figure 6). Under normoxia the hematocrit remained the same, at  $t = 5$  weeks ( $t = 33$  to 37 days) as well as at  $t = 5$  months ( $\sim 150$  days), with or without DHEA.

As expected, hypoxia increased the hematocrit. The hematocrit reached similar levels (45%) at  $t = 5$  weeks and  $t = 5$  months. The same trend was observed for red blood cell counts and blood hemoglobin content, while cellular hemoglobin content remained unchanged.

DHEAS did not affect the hematocrit nor red blood cell properties, neither in normoxia nor in hypoxia, at  $t = 5$  weeks and  $t = 5$  months.

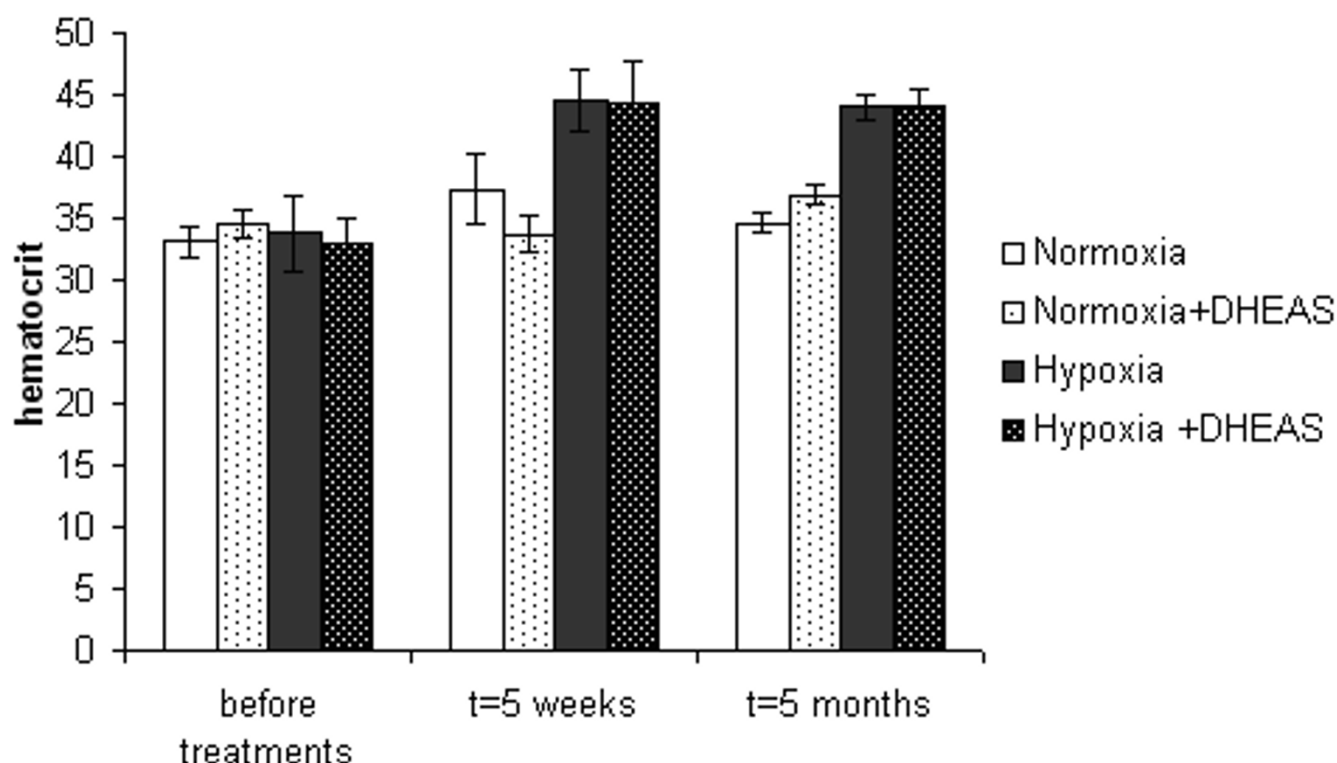
### Discussion

1. Hypoxia induced PH in old mice and DHEA prevented it. 2. Hypoxia drastically induced mortality and weight loss in old age. 3. In its sulfate form and at the used oral dose DHEA was detrimental to long-term survival in normoxia. 4. DHEA however largely prevented hypoxic death during the whole experiment.

### DHEA prevents hypoxic PH in old mice

#### Chronic hypoxia provoked PH in old mice

This is not particularly surprising as it also does it in young adult mice [21] and rats [23,24].



**Figure 6**

**Hematocrit.** Hematocrit as a function of groups and time. Hypoxia increased the hematocrit, and dehydroepiandrosterone (DHEAS) did not affect the hematocrit, in hypoxia or in normoxia.

#### *DHEA prevented hypoxic PH in mice*

DHEA has already been shown to prevent and reverse PH in rats [23,24]. DHEA is thought to be the relaxation of pulmonary arteries by opening large-conductance calcium-activated potassium channels [23,26,34], but this mechanism is controversial [26,27]. Mice knocked out for these channels [35-37] exist and it would be of interest to study the relaxation of pulmonary arteries by DHEA in such mice.

#### *DHEA prevented hypoxic PH in old age*

No previous study reported effects of DHEA on PH in old age. Old age is a common factor for PH incidence and low endogenous blood DHEA(S) levels in humans [28]. Therefore old age may play a particular role in the treatment of hypoxic PH by DHEA, and it was not obvious that results obtained in young adults could be transposed to old adults (especially from rats to mice). Application to humans is discussed further along with survival.

#### **Hypoxic death in old animals: a model for PH survival?**

We used old animals at an age when they naturally die in order to measure overall positive or negative health effects by increased or decreased survival of 'naturally dying' animals. Our mice trembled and there was a drastic increase

of death due to hypoxia (11% FIO<sub>2</sub>). To our knowledge this has not been described before and it is certainly due to the old age of the mice. In particular we also studied young adult mice (8 with DHEA and 16 without, unpublished data) for 4 months in the same hypoxic chamber, with no trembling behavior nor death ( $p < 0.001$ ).

*This age-related frailty to chronic hypoxia* was not foreseen. In particular, there does not seem to be an age-related frailty with respect to severe acute hypoxia [38,39]. In other species, flies and nematodes live longer under moderate hypoxia, possibly because of reduced oxidative stress, and it could be expected that the same might apply to mammals [40]. Our degree of hypoxia (11% oxygen) was clearly too severe to allow mice to benefit from reduced oxygen stress but a less severe degree (16% oxygen, unpublished data) still slightly reduced lifespan. Starting hypoxia at a younger age still reduces lifespan: a recent study has shown that rats kept under hypoxia from a young adult age rapidly develop cardiopulmonary remodeling and die when they are around 18 months old [25]. These rats were Wistar rats, which have a similar lifespan to C57BL/6 mice. If we suppose that hypoxia has similar effects on survival in both strains, this suggests that hypoxia only threatens life after ~18 months of age, what-

**Table 1: Blood patterns**

	Treatment		before treatments	t = 5 weeks	t = 5 months	
Blood hemoglobin content (g/dl)	Normoxia	Water	10.6 ± 0.5 (6)	12.4 ± 0.3 (7)	11.3 ± 0.9 (7)	**
	Normoxia	DHEAS	11.3 ± 0.3 (9)	11.5 ± 0.4 (7)	12.0 ± 0.2 (5)	
	Hypoxia	Water	11.0 ± 0.4 (6)	14.3 ± 0.3 (6)	13.9 ± 0.6 (8)	
	Hypoxia	DHEAS	11.3 ± 0.7 (6)	14.3 ± 0.9 (6)	14.3 ± 1.2 (8)	
Hematocrit (%)	Normoxia	Water	7.86 ± 0.3 (6)	8.65 ± 0.3 (7)	7.97 ± 0.7 (7)	**
	Normoxia	DHEAS	8.03 ± 0.4 (9)	7.83 ± 0.3 (7)	8.61 ± 0.2 (5)	
	Hypoxia	Water	7.81 ± 0.3 (6)	9.70 ± 0.3 (6)	9.29 ± 0.5 (8)	
	Hypoxia	DHEAS	7.74 ± 0.6 (6)	9.68 ± 0.5 (6)	9.21 ± 0.7 (8)	
Red cell count (10 <sup>3</sup> /mm <sup>3</sup> )	Normoxia	Water	42.2 ± 0.3 (6)	43.1 ± 0.5 (7)	42.9 ± 0.6 (7)	**
	Normoxia	DHEAS	43.1 ± 0.5 (9)	43.2 ± 0.7 (7)	43.0 ± 0.6 (5)	
	Hypoxia	Water	43.3 ± 0.4 (6)	46.0 ± 0.8 (6)	47.5 ± 0.4 (8)	
	Hypoxia	DHEAS	42.9 ± 0.4 (6)	46.2 ± 1.8 (6)	47.8 ± 1.2 (9)	
Mean red blood cell volume (μm <sup>3</sup> )	Normoxia	Water	42.2 ± 0.3 (6)	43.1 ± 0.5 (7)	42.9 ± 0.6 (7)	*
	Normoxia	DHEAS	43.1 ± 0.5 (9)	43.2 ± 0.7 (7)	43.0 ± 0.6 (5)	
	Hypoxia	Water	43.3 ± 0.4 (6)	46.0 ± 0.8 (6)	47.5 ± 0.4 (8)	
	Hypoxia	DHEAS	42.9 ± 0.4 (6)	46.2 ± 1.8 (6)	47.8 ± 1.2 (9)	
Mean cell hemoglobin concentration (g/dl)	Normoxia	Water	32.1 ± 0.2 (6)	33.1 ± 0.2 (7)	32.8 ± 0.3 (7)	
	Normoxia	DHEAS	32.6 ± 1.4 (9)	34.1 ± 0.3 (7)	32.6 ± 0.3 (5)	
	Hypoxia	Water	32.8 ± 0.4 (6)	32.0 ± 0.5 (6)	31.5 ± 0.3 (8)	
	Hypoxia	DHEAS	34.6 ± 0.3 (6)	32.4 ± 0.2 (6)	32.4 ± 0.3 (8)	
Mean cell hemoglobin (pg)	Normoxia	Water	13.5 ± 0.1 (6)	14.3 ± 0.2 (7)	14.1 ± 0.3 (7)	
	Normoxia	DHEAS	14.1 ± 0.7 (9)	14.7 ± 0.1 (7)	14.0 ± 0.2 (5)	
	Hypoxia	Water	14.2 ± 0.2 (6)	14.7 ± 0.1 (7)	15 ± 0.1 (8)	
	Hypoxia	DHEAS	14.9 ± 0.2 (6)	14.9 ± 0.5 (6)	15.4 ± 0.3 (8)	

Red blood parameters for the different treatments at different times. Blood hemoglobin content, hematocrit and red cell count were elevated under hypoxia compared to normoxia, at t = 5 weeks ( $p < 0.01$ ) and similarly at t = 5 months ( $p < 0.01$ ) (\*\*). Mean red blood cell volume was slightly elevated under hypoxia compared to normoxia, at t = 5 weeks ( $p < 0.05$ ) and slightly more at t = 5 months ( $p < 0.05$ ) (\*). Mice treated with dehydroepiandrosterone (named DHEAS in the table) had the same blood patterns than matching mice that did not received dehydroepiandrosterone (named Water in the table), whether under normoxia or hypoxia.

ever the duration of hypoxia before that age. The combination of this rat study with our mouse study suggests that in mammals, although hypoxic PH develops within a few weeks at any age, hypoxic PH becomes dangerous for health at later ages rather than after some disorder duration.

*In humans too*, there could be an age-related frailty to PH. It happens that the incidence of hospitalization and mortality from the disorder increases exponentially with age [5]. Moreover, there seems to be an age, around 45 years, when pulmonary arterial hypertension becomes life-threatening [4]. In fact hypoxic PH severity could be more related to patient age than disease duration. This could perhaps explain why apparently minor PH may be determinant for (older) COPD patients [10], and why some old smokers suddenly suffer after many years of COPD. Of course, this age-related concept does not concern all types of PH (such as PH in the newborn and probably fenfluramine-induced PH, [3]).

*We propose that our model* – consisting of studying survival of old animals under hypoxia accompanied or not by some treatment – may be useful for studying the overall effects of PH treatments which are destined for aged persons. If we accept the difference that time goes 30 to 40 times faster in mice, there is a surprisingly good match between our survival curves of old hypoxic mice, treated or not by DHEA, and the survival curves of COPD patients, mostly over 65 years old, treated or not by oxygenotherapy [13]. This, may suggest that hypoxic mice survival could be a speeded-up model for human PH survival.

#### **DHEA was detrimental to long-term survival**

*An appropriate control* should not affect survival and should be transposable to humans. However DHEA induced an unexpected decrease of survival after the age of 24 months compared to the control mice ( $p = 0.0025$ ), and this may not at all apply to humans. In humans it was shown that DHEA may be safely administered to older



persons at the daily oral dose of 50 mg ( $\sim 1$  mg/kg/day) for one year [28]. In comparison, the doses used to treat PH in animals are larger ( $\sim 9$  mg/kg/day by Hampl V *et al.* [24],  $\sim 15$  mg/kg/day by Bonnet *et al.* [23] and  $\sim 25$  mg/kg/day in our study). In fact, whereas in humans DHEA(S) is a major steroid circulating in the blood, no detectable DHEA(S) was found in the blood of laboratory animals such as mice or rats [41]. Therefore, DHEA "supplementation" is pharmacological (i.e. non physiological) in mice and cannot be considered as a hormonal replacement therapy.

*The effect on lifespan of DHEA administration in mice* has been studied several times. High doses of free DHEA incorporated into the diet (on the order of 0.4%, which corresponds to  $\sim 12$  mg/day/mouse, that is 10 to 20 times more than in our study) have been shown to increase the lifespan of particular short-lived mice [42-44]. As C57BL/6 mice do not seem to like DHEA [29,30,45], we preferred to use lower doses and the sulfate form in drinking water (0.25 mg/ml dissolves well in water) to avoid survival bias by caloric restriction, and we found that it reduced the lifespan of 21-month-old male C57BL/6 mice.

*We are not the first to find that DHEAS does not extend the lifespan of mice.* A previous study found that 10 times less DHEAS (0.025 mg/ml in drinking water) did not affect the lifespan of 12-month-old male C57BL/6 mice [31]. The authors suggested that the lack of effect could come from an insufficient dosage. Another study found that the intermediate dose of 0.1 mg/ml in drinking water from weaning age insignificantly decreased the lifespan of genetically heterogeneous mice [46]. We multiplied the dose by 3 and the decrease of lifespan became very significant. Although multiple parameters make the comparisons complex, a global interpretation of these results would be that DHEAS in drinking water does not affect mouse lifespan at doses smaller than 0.1 mg/ml ( $\sim 9$  mg/kg/day) and decreases mouse lifespan at larger doses. In fact, positive effects of dehydroepiandrosterone may be present but masked by negative effects due to the dose and way of administration, such as long-term hepatic disturbances [47][48].

### **DHEA largely prevented hypoxic death**

#### *DHEA globally treated hypoxic old mice*

Although DHEAS administration appeared to be detrimental in the long term (as seen by late mortality under normoxia), and although hypoxic animals treated by DHEA still lost weight and trembled, DHEA largely (but not completely) prevented the hypoxic mortality over the whole experiment. This overall beneficial survival effect is the best possible answer to our questions: DHEA not only treats hypoxic PH but also hypoxic (old) mice.

### **A role for high hematocrit?**

The vasorelaxation of pulmonary arteries by DHEA could have led to overall negative effects since hypoxic vasoconstriction of pulmonary arteries is useful to improve blood oxygenation. The question arises of whether, with DHEA treatment, the body managed without the oxygen provided by vasoconstriction or another mechanism for providing an adequate oxygen supply came into play. The high blood hemoglobin content here may play a role. By preventing cardiopulmonary remodeling but permitting increased hematocrit under hypoxia, DHEA could be favorable to the animal's health by preventing heart failure (due to PH) while allowing high oxygenation.

*The prevention of hypoxic death by DHEA in mice recalls us the prospective PAQUID study in humans*, where a strong inverse correlation between natural DHEA(S) blood levels and the ten year mortality in old male smokers and former smokers has been reported [49]. There is an interesting analogy between  $\geq 65$ -year-old male human smokers and  $\geq 21$ -month-old male hypoxic mice, on the time scale of the mouse. This analogy is important as we designed our mice survival study with the results of the PAQUID study in mind. Nevertheless it must be remembered that mice, unlike humans, do not have detectable endogenous circulating DHEA(S) [41]. Therefore the above analogies compare pharmacological (mice) effects with physiological/pharmacological (human) effects. It remains that large doses of DHEA may be safely administered to humans and that PH complicating COPD is a morbid condition. Thus it seems that specific human clinical trials aimed at deriving statistics from humans taking DHEA supplementation, and including females who have not been taken into account in this (mouse) study, would be justified. In the meanwhile, care should be taken to avoid uncontrolled consequences of our findings.

### **Conclusion**

There seems to be a frailty to hypoxic PH that is particular to old age, in mice and possibly in humans. This suggests that survival studies with aged mice under hypoxia may be pertinent for evaluating therapies for aged patients having PH. In that framework, DHEA was found to remarkably improve survival under hypoxia. The comparison between mice and humans is not obvious, but our findings interestingly resemble human observations, that together suggest trials of DHEA treatment to PH and COPD in humans.

### **Abbreviations**

FIO<sub>2</sub>: Fraction of Inspired Oxygen

PH: Pulmonary Hypertension

COPD: Chronic Obstructive Pulmonary Disease

DHEA(S): DeHydroEpiAndrosterone (sulfate)

## Competing interests

This work was financed by the Association pour la Recherche sur les Nicotianés (Fleury-Les-Aubrais, France).

## Authors' contributions

EHD carried out the design of the study, performed the statistical analysis, carried out the environmental setting, participated in blood analysis, anatomopathological analysis and drafted the manuscript. JQ carried out the anatomopathological analysis and helped to design the study. EEB participated in design and coordination of the study and helped to draft the manuscript.

## Additional material

### Additional file 1

SurvivalPower

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1465-9921-7-144-S1.pdf>]

## Acknowledgements

This work was supported by a grant to EEB from the Agence Nationale de la Recherche (Paris, France). The nitrogen generator was generously provided by Air Liquide Santé Gaz Médicaux (Paris, France). Nathalie Ba technically contributed to the histological studies. We would like to mention the excellent technical contribution made by Rachid Mekri in helping setting the environment, in taking care of the mice and weighing them. We are grateful to Marie-Pierre Morin-Surun for stimulating discussions about respiratory adaptation to hypoxia. We thank Olivier Trassard for stimulating discussions about setting the environment and coordinating the study.

## References

- Liu C, Liu K, Ji Z, Liu G: **Treatments for pulmonary arterial hypertension.** *Respir Med* 2006, **100**:765-774.
- Levine DJ: **Diagnosis and management of pulmonary arterial hypertension: Implications for respiratory care.** *Respir Care* 2006, **51**:368-381.
- Kuhn KP, Byrne DW, Arbogast PG, Doyle TP, Loyd JE, Robbins IM: **Outcome in 91 consecutive patients with pulmonary arterial hypertension receiving epoprostenol.** *Am J Respir Crit Care Med* 2003, **167**:580-586.
- Hyduk A, Croft JB, Ayala C, Zheng K, Zheng ZJ, Mensah GA: **Pulmonary hypertension surveillance - United States, 1980-2002.** *MMWR Surveill Summ* 2005, **54**:1-28.
- Rubin LJ: **Primary pulmonary hypertension.** *N Engl J Med* 1997, **336**:111-117.
- Naeije R: **Pulmonary Hypertension and Right Heart Failure in Chronic Obstructive Pulmonary Disease.** *Proceedings of the ATS* 2005, **2**:20-22.
- Weitzenblum E, Chaouat A: **Obstructive sleep apnea syndrome and the pulmonary circulation.** *Ital Heart J* 2005, **6**:795-798.
- Chaouat A, Bugnet AS, Kadaoui N, Schott R, Enache I, Ducolone A, Ehrhart M, Kessler R, Weitzenblum E: **Severe pulmonary hypertension and chronic obstructive pulmonary disease.** *Am J Respir Crit Care Med* 2005, **172**:189-194.
- Chaouat A, Kraemer JP, Canuet M, Kadaoui N, Ducolone A, Kessler R, Weitzenblum E: **Pulmonary hypertension associated with disorders of the respiratory system.** *Presse Med* 2005, **34**:1465-1474.
- Oswald-Mammoser M, Weitzenblum E, Quoix E, Moser G, Chaouat A, Charpentier C, Kessler R: **Prognostic factors in COPD patients receiving long-term oxygen therapy. Importance of pulmonary artery pressure.** *Chest* 1995, **107**:1193-1198.
- Report of the Medical Research Council Working Party: **Long term domiciliary oxygen therapy in chronic hypoxic cor pulmonale complicating chronic bronchitis and emphysema.** *Lancet* 1981, **1**:681-686.
- Nocturnal Oxygen Therapy Trial Group: **Continuous or nocturnal oxygen therapy in hypoxemic chronic obstructive lung disease: a clinical trial.** *Ann Intern Med* 1980, **93**:391-398.
- Calverley PM, Walker P: **Chronic obstructive pulmonary disease.** *Lancet* 2003, **362**:1053-1061.
- Abe K, Shimokawa H, Morikawa K, Uwatoku T, Oi K, Matsumoto Y, Hattori T, Nakashima Y, Kaibuchi K, Sueishi K, Takeshita A: **Long-term treatment with a Rho-kinase inhibitor improves monocrotaline-induced fatal pulmonary hypertension in rats.** *Circ Res* 2004, **94**:385-393.
- Schermlay RT, Kreisselmeier KP, Ghofrani HA, Yilmaz H, Butrous G, Ermert L, Ermert M, Weissmann N, Rose F, Guenther A, Walrath D, Seeger VV, Grimminger F: **Chronic sildenafil treatment inhibits monocrotaline-induced pulmonary hypertension in rats.** *Am J Respir Crit Care Med* 2004, **169**:39-45.
- Kodama K, Adachi H: **Improvement of mortality by long-term E4010 treatment in monocrotaline-induced pulmonary hypertensive rats.** *J Pharmacol Exp Ther* 1999, **290**:748-752.
- Cowan KN, Heilbut A, Humpl T, Lam C, Ito S, Rabinovitch M: **Complete reversal of fatal pulmonary hypertension in rats by a serine elastase inhibitor.** *Nat Med* 2000, **6**:698-702.
- Petry TW, Sipes IG: **Modulation of monocrotaline-induced hepatic genotoxicity in rats.** *Carcinogenesis* 1987, **8**:415-419.
- Boyd MR: **Effects of inducers and inhibitors on drug-metabolizing enzymes and on drug toxicity in extrahepatic tissues.** *Giba Found Symp* 1980, **76**:43-66.
- Kim HY, Stermitz FR, Molyneux RJ, Wilson DW, Taylor D, Coulombe RA Jr: **Structural influences on pyrrolizidine alkaloid-induced cytopathology.** *Toxicol Appl Pharmacol* 1993, **122**:61-69.
- Zhao L, Mason NA, Morrell NW, Kojonazarov B, Sadykov A, Maripov A, Mirrakhimov MM, Aldashev A, Wilkins MR: **Sildenafil inhibits hypoxia-induced pulmonary hypertension.** *Circulation* 2001, **104**:424-428.
- Nakanishi K, Tajima F, Osada H, Nakamura A, Yagura S, Kawai T, Suzuki M, Torikata C: **Pulmonary, vascular responses in rats exposed to chronic hypobaric hypoxia at two different altitude levels.** *Pathol Res Pract* 1996, **192**:1057-1067.
- Bonnet S, Dumas-de-La-Roque E, Begueret H, Marthan R, Fayon M, Dos Santos P, Savineau JP, Baulieu EE: **Dehydroepiandrosterone (DHEA) prevents and reverses chronic hypoxic pulmonary hypertension.** *Proc Natl Acad Sci USA* 2003, **100**:9488-9493.
- Hampel V, Bibova J, Povysilova V, Herget J: **Dehydroepiandrosterone sulphate reduces chronic hypoxic pulmonary hypertension in rats.** *Eur Respir J* 2003, **21**:862-865.
- La Padula P, Costa LE: **Effect of sustained hypobaric hypoxia during maturation and aging on rat myocardium. I. Mechanical activity.** *J Appl Physiol* 2005, **98**:2363-2369.
- Farrukh IS, Peng W, Orlinska U, Hoidal JR: **Effect of dehydroepiandrosterone on hypoxic pulmonary vasoconstriction: a Ca<sup>2+</sup>-activated K<sup>+</sup>-channel opener.** *Am J Respir Cell Mol Biol* 1999, **20**:737-745.
- Gupte SA, Li K, Okada T, Sato K, Oka M: **Inhibitors of Pentose Phosphate Pathway Cause Vasodilation: Involvement of Voltage-Gated Potassium Channels.** *Am J Respir Cell Mol Biol* 1999, **20**:737-745.
- Baulieu EE, Thomas G, Legrain S, Lahlou N, Roger M, Debuire B, Faucounau V, Girard L, Hervy MP, Latour F, Leaud MC, Mokrane A, Pitti-Ferrandi H, Trivalle C, de Lacharriere O, Nouveau S, Rakoto-Arison B, Souberbielle JC, Raison J, Le Bouc Y, Raynaud A, Girerd X, Forette F: **Dehydroepiandrosterone (DHEA), DHEA sulfate, and aging: contribution of the DHEA Study to a sociobiomedical issue.** *Proc Natl Acad Sci USA* 2000, **97**:4279-4284.
- Bradlow HL: **Feeding DHEA to C57/B16 mice.** *Proc Soc Exp Biol Med* 2000, **224**:201-202.
- Wright BE, Abadie J, Svec F, Porter JR: **Does taste aversion play a role in the effect of dehydroepiandrosterone in Zucker rats?** *Physiol Behav* 1994, **55**:225-229.

31. Pugh TD, Oberley TD, Weindruch R: **Dietary Intervention at Middle Age: Caloric Restriction but not Dehydroepiandrosterone Sulfate Increases Lifespan and Lifetime Cancer Incidence in Mice.** *Cancer Res* 1999, **59**:1642-1648.
32. Forster MJ, Morris P, Sohal RS: **Genotype and age influence the effect of caloric intake on mortality in mice.** *FASEB* 2003, **17**:690-692.
33. Weissmann N, Manz D, Buchspies D, Keller S, Mehling T, Voswinckel R, Quanz K, Ghofrani HA, Schermuly RT, Fink L, Seeger W, Gassmann M, Grimminger F: **Congenital erythropoietin over-expression causes "anti-pulmonary hypertensive" structural and functional changes in mice, both in normoxia and hypoxia.** *Thromb Haemost* 2005, **94**:630-638.
34. Peng W, Hoidal JR, Farrukh IS: **Role of a novel KCa opener in regulating K<sup>+</sup> channels of hypoxic human pulmonary vascular cells.** *Am J Respir Cell Mol Biol* 1999, **20**:737-745.
35. Rüttiger L, Sausbier M, Zimmermann U, Winter H, Braig C, Engel J, Knirsch M, Arntz C, Langer P, Hirt B, Müller M, Kopschall I, Pfister M, Munkner S, Rohbock K, Pfaffl I, Rusch A, Ruth P, Knipper M: **Deletion of the Ca<sup>2+</sup>-activated potassium (BK) alpha-subunit but not the BKbeta1-subunit leads to progressive hearing loss.** *Proc Natl Acad Sci USA* 2004, **101**:12922-12927.
36. Sausbier M, Hu H, Arntz C, Feil S, Kamm S, Adelsberger H, Sausbier U, Sailer CA, Feil R, Hofmann F, Korth M, Shipston MJ, Knaus HG, Wolfer DP, Pedroarena CM, Storm JF, Ruth P: **Cerebellar ataxia and Purkinje cell dysfunction caused by Ca<sup>2+</sup>-activated K<sup>+</sup> channel deficiency.** *Proc Natl Acad Sci USA* 2004, **101**:9474-9478.
37. Brenner R, Perez GJ, Bonev AD, Eckman DM, Kosek JC, Wiler SW, Patterson AJ, Nelson MT, Aldrich RW: **Vasoregulation by the beta1 subunit of the calcium-activated potassium channel.** *Nature* 2000, **407**:870-876.
38. Stupfel M, Moutet JP, Magnier M: **An apparently paradoxical action of aging: decrease of acute hypoxic mortality in male aged rats.** *J Gerontol* 1975, **30**:154-156.
39. Sato Y, Kitani K, Kanai S, Nokubo M, Ohta M: **Differences in tolerance to hypoxia/anoxia in mice of different ages.** *Res Commun Chem Pathol Pharmacol* 1991, **73**:209-220.
40. Longo VD: **Oxygen? No thanks, I'm on a diet.** *Sci Aging Knowledge Environ* 2002:pe10.
41. Baulieu EE: **Dehydroepiandrosterone (DHEA): a fountain of youth?** *J Clin Endocrinol Metab* 1996, **88**:3147-3151.
42. Lucas JA, Ahmed SA, Casey ML, MacDonald PC: **Prevention of autoantibody formation and prolonged survival in New Zealand black/New Zealand white F1 mice fed dehydroisoandrosterone.** *J Clin Invest* 1985, **75**:2091-2093.
43. Perkins SN, Hursting SD, Haines DC, James SJ, Miller BJ, Phang JM: **Chemoprevention of spontaneous tumorigenesis in nullizygous p53-deficient mice by dehydroepiandrosterone and its analog 16alpha-fluoro-5-androsten-17-one.** *Carcinogenesis* 1997, **18**:989-994.
44. Schwartz AG: **Inhibition of spontaneous breast cancer formation in female C3H(Avy/a) mice by long-term treatment with dehydroepiandrosterone.** *Cancer Res* 1979, **39**:1129-1132.
45. Catalina F, Kumar V, Milewich L, Bennett M: **Food restriction-like effects of dehydroepiandrosterone: decreased lymphocyte numbers and functions with increased apoptosis.** *Proc Soc Exp Biol Med* 1999, **221**:326-335.
46. Miller RA, Chrisp C: **Lifelong treatment with oral DHEA sulfate does not preserve immune function, prevent disease, or improve survival in genetically heterogeneous mice.** *J Am Geriatr Soc* 1999, **47**:960-966.
47. Rao MS, Subbarao V, Yeldandi AV, Reddy JK: **Hepatocarcinogenicity of dehydroepiandrosterone in the rat.** *Cancer Res* 1992, **52**:2977-9.
48. Mayer D, Forstner K: **Impact of dehydroepiandrosterone on hepatocarcinogenesis in the rat (Review).** *Int J Oncol* 2004, **25**:1021-30.
49. Mazat L, Lafont S, Berr C, Debuire B, Tessier JF, Dartigues JF, Baulieu EE: **Prospective measurements of dehydroepiandrosterone sulfate in a cohort of elderly subjects: relationship to gender, subjective health, smoking habits, and 10-year mortality.** *Proc Natl Acad Sci USA* 2001, **98**:8145-8150.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

