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Oxidative stress induced Interleukin-32 mRNA expression in human bronchial epithelial cells

Megumi Kudo¹, Emiko Ogawa^{1,2,4*}, Daisuke Kinose¹, Akane Haruna¹, Tamaki Takahashi¹, Naoya Tanabe¹, Satoshi Marumo¹, Yuma Hoshino¹, Toyohiro Hirai¹, Hiroaki Sakai³, Shigeo Muro¹, Hiroshi Date³ and Michiaki Mishima¹

Abstract

Background: Chronic obstructive pulmonary disease (COPD) is characterized by airflow obstruction and persistent inflammation in the airways and lung parenchyma. Oxidative stress contributes to the pathogenesis of COPD. Interleukin (IL)-32 expression has been reported to increase in the lung tissue of patients with COPD. Here, we show that IFN γ upregulated IL-32 expression and that oxidative stress augmented IFN γ -induced-IL-32 expression in airway epithelial cells. We further investigated transcriptional regulation responsible for IFN γ induced IL-32 expression in human airway epithelial cells.

Methods: Human bronchial epithelial (HBE) cells were stimulated with H₂O₂ and IFN γ , and IL-32 expression was evaluated. The cell viability was confirmed by MTT assay. The intracellular signaling pathways regulating IL-32 expression were investigated by examining the regulatory effects of MAPK inhibitors and JAK inhibitor after treatment with H₂O₂ and IFN γ , and by using a ChIP assay to identify transcription factors (i.e. c-Jun, CREB) binding to the IL-32 promoter. Promoter activity assays were conducted after mutations were introduced into binding sites of c-Jun and CREB in the IL-32 promoter. IL-32 expression was also examined in HBE cells in which the expression of either c-Jun or CREB was knocked out by siRNA of indicated transcription factors.

Results: There were no significant differences of cell viability among groups. After stimulation with H₂O₂ or IFN γ for 48 hours, IL-32 expression in HBE cells was increased by IFN γ and synergistically upregulated by the addition of H₂O₂. The H₂O₂ augmented IFN γ induced IL-32 mRNA expression was suppressed by a JNK inhibitor, but not by MEK inhibitor, p38 inhibitor, and JAK inhibitor I. Significant binding of c-Jun and CREB to the IL-32 promoter was observed in the IFN γ + H₂O₂ stimulated HBE cells. Introducing mutations into the c-Jun/CREB binding sites in the IL-32 promoter prominently suppressed its transcriptional activity. Further, knocking down CREB expression by siRNA resulted in significant suppression of IL-32 induction by IFN γ and H₂O₂ in HBE cells.

Conclusion: IL-32 expression in airway epithelium may be augmented by inflammation and oxidative stress, which may occur in COPD acute exacerbation. c-Jun and CREB are key transcriptional factors in IFN γ and H₂O₂ induced IL-32 expression.

Keywords: COPD, acute exacerbation, IFN γ

Background

Chronic obstructive pulmonary disease (COPD) is characterized by non-fully reversible airflow obstruction and persistent inflammation in the airways and lung parenchyma [1-3]. Airway epithelial cells are one of the

most important sources of inflammatory mediators that play important roles in the pathogenesis of COPD. Several reports have indicated that various factors such as smoking, infection, and proteases activate airway epithelial cells in COPD patients [2,4-6] and this is followed by the secretion of chemokines (CCL2, CXCL5, and CXCL10), inflammatory cytokines (TNF α , IL (interleukin)-7 family members (TSLP), and IL-12), and growth

* Correspondence: eogawa@belle.shiga-med.ac.jp

¹Department of Respiratory Medicine, Kyoto University Graduate School of Medicine, Kyoto, Japan

Full list of author information is available at the end of the article

factors (GM-CSF and TGF- β) [1,7,8] by these activated cells.

Cigarette smoke contains oxidants and free radicals, many of which remain in the airway for long periods [9,10], is a major source of the oxidative stress that contributes to the pathogenesis of COPD. Also, activated inflammatory cells such as alveolar macrophages, neutrophils, eosinophils, monocytes, lymphocytes, and epithelial cells in the airways of COPD patients generate reactive oxidant species in response to inflammatory mediators [10].

IL-32, which was originally reported as natural killer cell transcript 4, is known to be expressed in mononuclear cells, T cells, epithelial cells, and endothelial cells in human tissues [11-14]. Cytokines such as IFN γ and TNF α promote IL-32 expression in these cells [15]. Lipopolysaccharide (LPS) also upregulates IL-32 expression in human mononuclear cells [16]. On the other hand, IL-32 induces monocytes and macrophages to produce inflammatory cytokines including TNF α , IL-1 β , IL-6, IL-4, MIP, IFN γ , and IL-8 and also regulates T cell apoptosis and monocyte differentiation to macrophages [16-18].

IL-32 has also been reported to be associated with the pathogenesis of inflammatory bowel disease and rheumatoid arthritis [12,14]. Recently, it was reported that the expression of IL-32 was increased in macrophages and airway epithelial cells in the lung tissues of COPD patients compared to that in the lungs of control smokers and non-smoking subjects. Furthermore, this IL-32 expression was reported to be negatively correlated with FEV1 and positively correlated with the expression levels of TNF α and CD8 [19]. However, the mechanisms of IL-32 gene regulation in the lungs of COPD patients are still unknown.

In this study, we evaluated whether oxidative stress affects IL-32 expression induced by IFN γ and determined a mechanism by which this expression is regulated using a human bronchial epithelial (HBE) cell culture system.

Methods

Reagents

IFN γ was obtained from Peprotech (Rocky Hill, NJ, USA). The JNK inhibitor SP600125 was from SABioscience (Frederick MD, USA), the MEK1 inhibitor PD98059 was purchased from Invitrogen (Carlsbad, CA, USA), and the p38 inhibitor SB203580 was from Enzo Life Sciences (Plymouth meeting, PA, USA). JAK inhibitor SC204021 was from Santa Cruz Biotechnology (Santa Cruz, CA, USA). The antibodies used were as follows: anti-human IL-32 monoclonal antibody, #KU32-52, was from Biogenex (San Diego, CA, USA); anti- β -actin polyclonal antibody was from Imgenex (San Diego,

CA, USA); anti-c-Jun antibody and anti-CREB antibody were from Cell Signaling Technology (Danvers, MA, USA); and anti-RNA polymerase II antibody was from Santa Cruz Biotechnology (Santa Cruz). CREB and c-Jun siRNA were from Cell Signaling Technology. Control siRNA was from Invitrogen.

Human bronchial epithelial (HBE) cells

HBE cells were provided from the Lung Registry of Kyoto University. The Kyoto University review board for human studies approved the protocols employed in this study, and written informed consent was obtained from all patients. Briefly, HBE cells were isolated from patients who underwent lung lobectomy for tumor resection at Kyoto University Hospital and cultured in LHC-9 medium, (Invitrogen, Carlsbad, CA, USA). The patients were all never smokers with normal pulmonary function and had no history of pulmonary diseases. The cells were isolated from the portions of bronchus that were not involved in tumor according to a modified version of a previous method [20].

Cell culture

All experiments were carried out using cells at passages 1 to 4. To get rid of corticosteroid from the media, HBE cells were cultured in corticosteroid-free Bronchial/Tracheal Epithelial Cell Basal Medium with growth factors (BEGM) (Lonza, Basel, Switzerland) instead of using LHC-9 for 24 hours prior to IFN γ stimulation. For H₂O₂ treatment, the indicated concentration of H₂O₂ was added to the medium 2 hours before stimulation with 10 ng/mL IFN γ . To determine the effect of modulating MAPK activity on H₂O₂ augmented IFN γ induced IL-32 mRNA expression, 10 μ M of JNK inhibitor, 20 μ M of MEK inhibitor, 10 μ M of p38 inhibitor, 5 μ M JAK inhibitor I, or DMSO as the vehicle were added to the cell culture media 2 hours before IFN γ stimulation. In all the treatments, the final concentration of DMSO was less than 0.075%.

MTT cell growth assay

After stimulation with 250 μ M H₂O₂, 500 μ M H₂O₂, 10 ng/ml IFN γ or 250 μ M H₂O₂ and 10 ng/ml IFN γ for 48 hours, the viability of the cells were examined using MTT cell growth assay kit (Millipore, Billerica, MA, USA) according to the manufacturer's instruction.

RNA isolation and quantitative real-time PCR

RNA was extracted from the HBE cells after H₂O₂ treatment plus 4, 8, and 24 hours of IFN γ stimulation using TRIzol reagent[®] (Invitrogen) according to the manufacturer's protocol. Quantitative real-time PCR was carried out to determine IL-32 gene expression and β -actin gene expression as an internal control using the ABI-

PRISM7300 Sequence-Detection-System (Applied Biosystems). The PCR primers used to detect all splice variants [21] of IL-32 mRNA and the β -actin mRNA were as follows: forward primer for IL-32, 5'-ATC CTC AAC ATC CGG GAC AG-3'; reverse primer for IL-32, 5'-ATG AGG AGC AGC ACC CAG A-3'; forward primer for β -actin, 5'-CCG ATC CAC ACG GAG TAC TTG-3; reverse primer for β -actin, 5'-CCG ATC CAC ACG GAG TAC TTG-3'.

Protein extraction and Western blot analysis

HBE cell lysate protein was extracted after H₂O₂ treatment plus 48 hours IFN γ stimulation. HBE cells were lysed in RIPA buffer (50 mM Tris HCl, 150 mM NaCl, 1% TritonX) together with protease inhibitor cocktail (SIGMA, St. Louis, MO, USA). Twenty μ g of proteins were electrophoresed on SDS/15% polyacrylamide gels and transferred to nitrocellulose membranes. IL-32 and β -actin were detected using specific antibodies.

Chromatin immunoprecipitation assays

Chromatin immunoprecipitation assays were performed using the Low Cell CHIP kit[®] (Nippongene, Tokyo, Japan) according to the manufacturer's protocol. Briefly, HBE cells were treated with or without H₂O₂ and/or IFN γ for 30 minutes, and then the cells were fixed with formaldehyde to cross-link proteins with DNA. The reaction was quenched by 5 minutes treatment with 1.25 M glycine. Then, the cells were lysed, and chromatin molecules were sonicated to a length of between 200 and 1000 bps. The sonicated chromatin was incubated overnight at 4°C with magnetic bead-bound antibodies against c-Jun, CREB, RNA polymerase II or normal rabbit IgG as a negative control. After being washed, the immunoprecipitated chromatins were reverse cross-linked, and the recovered DNA was purified for real-time PCR. The PCR primers used to amplify the c-Jun and CREB binding sites in the IL-32 promoter region, which are located between nucleotides -96 to 134, relative to the transcription start site at +1, were designed according to the NM_001012631 NCBI sequence. The PCR primers used were as follows: the forward primer, 5'-CAA GGA CAG GGT CCA AAT TC-3', and the reverse primer, 5'-GGT CCG TCC CTG GCT GGG C-3',

Mutagenesis of the IL-32 promoter sequence and luciferase reporter assays

We generated an IL-32 promoter construct with the sequence between nucleotides -120 and +530 relative to the transcription start site at +1 of the IL-32 gene containing the c-Jun/CREB consensus sequence by PCR using human genomic DNA as a template. The PCR primers used were as follows: the forward primer, 5'-TGA

TCC AGA AGT TTC TCT GGC CTC TGG A-3', and the reverse primer, 5'-GCA GCC TCT CAC TCA CCT TCG-3', and a 4 bp mutation described in a previous report [22] was introduced into the c-Jun/CREB consensus sequence. Briefly, the TGACGTCA sequence which contains the tgacgtca c-Jun and tggctgacgtcacctt CREB consensus binding sequences and which is located from nucleotides -30 to -23 in the IL-32 gene promoter was changed to TcaatTCA using PCR primers that included these changes. Promoter constructs with mutated or respective wild-type sequences were ligated into the cloning vector and amplified in *E.coli*. As the cloned promoter sequences (with or without the mutation) were located between XhoI and HindIII restriction sites, the restriction enzyme (XhoI and HindIII) -treated promoter sequences were ligated with firefly luciferase coding reporter vectors (pGL4.10[luc2], Promega, Madison WI, USA) and referred to as pWild-Luc (without the mutation) and pMutant-Luc (with the mutation), respectively. All constructs were verified by sequencing.

To determine the transcriptional activity of these IL-32 promoter constructs, HBE cells plated at 0.5×10^5 cells/well in 12-well plates were transiently transfected with 1 μ g of either pWild-Luc or pMutant-Luc per well using PrimeFect II (Takara, Otsu, Japan) according to the manufacturer's protocol. The cells were then co-transfected with 0.5 μ g/well of the Renilla luciferase reporter construct (pGL4.74[hRLuc/TK], Promega) to normalize for differences in transfection efficiency between wells. After 2 hours of transfection, the medium was replaced with corticosteroid-free BEGM. Sixteen hours after transfection, the cells were pretreated with 250 μ M H₂O₂ for 2 hours and then stimulated with 10 ng/mL of IFN γ . Six hours after the IFN γ stimulation, the cells were lysed with Passive Lysis Buffer (Promega). Luciferase assays were performed using the Dual luciferase reporter assay system[®] (Promega), and the expression of firefly and Renilla luciferase was measured with a luminometer (Lumat LB 9507[®], Bethold, Bad Wildbad, Germany). Firefly luciferase activity was measured by adding luciferase assay reagent II (LARII, Promega), and Renilla luciferase was measured in a separate tube by adding LARII and Stop & Glo reagent (Promega).

Knockdown of c-Jun and CREB by siRNA transfection into HBE cells

To confirm a role of c-Jun and/or CREB in H₂O₂ and IFN γ induced IL-32 gene regulation, c-Jun or CREB was knocked out in HBE cells using either c-Jun siRNA or CREB siRNA. HBE cells were incubated with the indicated siRNA and transfection reagent (Lipofectamine RNAiMAX, Invitrogen) for 24 hours. Then cells were stimulated with H₂O₂ for 2 hours and then with IFN γ

for 24 hours before extraction of mRNA for evaluation of IL-32 by real time RT-PCR.

Statistical analysis

Analysis of variance (ANOVA) with the post hoc Fisher's test was used for comparisons among groups. All analyses were performed using GraphPad Prism Ver.4 (Graphpad software, San Diego, CA, USA). Values of $p < 0.05$ were considered statistically significant.

Results

H₂O₂ augmented IFN γ -induced IL-32 expression in HBE cells

The influence of the H₂O₂ concentration on the cell death of HBE cells was determined. Although treatment with 500 μ M H₂O₂ resulted in cell death, treatment with 250 μ M H₂O₂ did not (data not shown). The cells were confirmed to be alive at least 48 hours after stimulation with 250 μ M H₂O₂ and/or 10 ng/ml IFN γ and the viability of the cells were confirmed by MTT assay (Figure 1A). After these preliminary experiments, we used H₂O₂ at a concentration of 250 μ M for 2 hours prior to IFN γ stimulation in the rest of our investigations.

As shown in Figure 1B, IL-32 mRNA expression was upregulated by IFN γ stimulation time dependently. After 4 and 8 hours stimulation with IFN γ , IL-32 mRNA expression was increased by 3.6 and 6.7 times, respectively, compared with control. Furthermore, IL-32 mRNA expression was increased by 4.9 and 11.0 times after 4 and 8 hours by IFN γ with H₂O₂. However, synergistic upregulation by H₂O₂ was not significant at both times. Twenty four hours stimulation with IFN γ alone significantly upregulated IL-32 mRNA expression in HBE cells. Pretreatment with 250 μ M H₂O₂ in addition to IFN γ synergistically upregulated IL-32 mRNA expression, although H₂O₂ alone did not influence its expression.

To determine the protein expression of IL-32 in HBE cells, Western blot analyses were performed using whole cell lysates from HBE cells treated with or without H₂O₂ and/or 48 hours of IFN γ stimulation. The antibody used for detecting IL-32 protein recognizes the 4 splice variants of IL-32, α , β , γ , and δ . However, the difference in size between IL-32 β and δ was so small, it was hard to distinguish between the two in the Western blotting analyses. As shown in Figure 1C, bands appeared at 22 and 26 kDa, which represented IL-32 β and/or δ , and IL-32 γ , respectively. IL-32 α could not be detected, suggesting that it is weakly expressed in HBE cells.

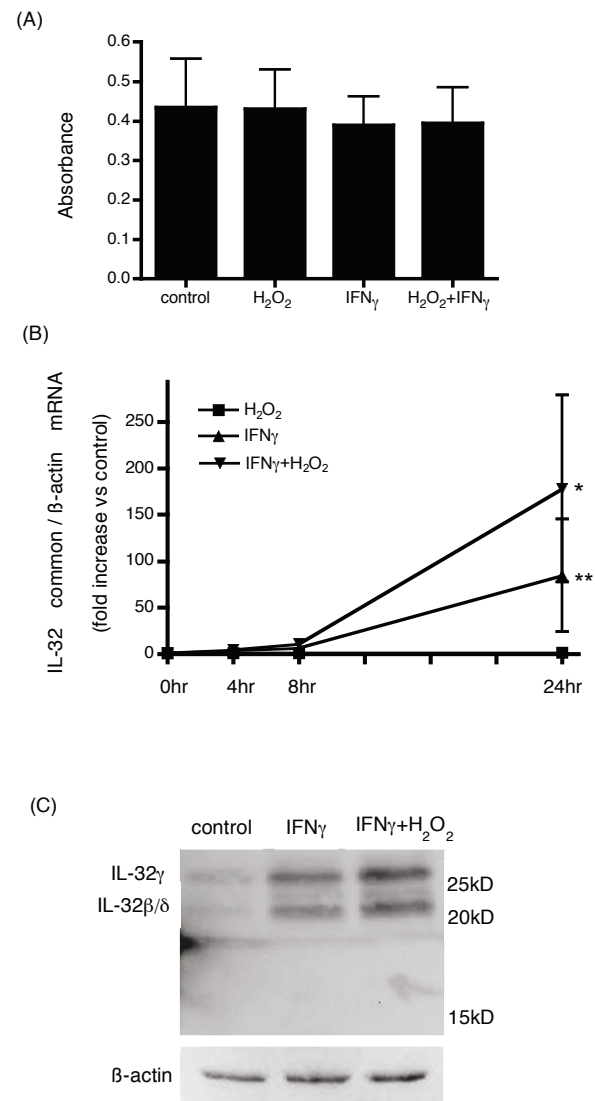


Figure 1 Cell viability and IL-32 expression in IFN γ and H₂O₂ + IFN γ stimulated HBE cells. HBE cells were incubated with 10 ng/ml IFN γ or 250 μ M H₂O₂ and 10 ng/ml IFN γ for 48 hours. The viability of the cells was examined using MTT assay (A). HBE cells were incubated with or without 2 hour-H₂O₂ pretreatment and then with or without IFN γ stimulation for 0, 4, 8, and 24 hours. The expression levels of IL-32 mRNA adjusted with expression levels in non-stimulated cells in each time point were determined by quantitative real-time PCR. HBE cells stimulated with H₂O₂, with IFN γ , and with IFN γ and H₂O₂, were presented in square plots, triangle plots, and inverted-triangle plots, respectively in graph (B). The bars show the means \pm SE from data performed on 3 different individuals. * $p < 0.01$ significantly different. (C) After 48 hours of IFN γ treatment 20 μ g of cell lysates were subjected to Western blotting. The 22 kDa band represents IL-32 β / δ protein, and the 26 kDa band represents IL-32 γ protein. The results shown are representative of 3 independent experiments.

IFN γ induced IL-32 expression was suppressed by inhibiting the JNK pathway

To investigate which signaling pathways are responsible for regulating IL-32 in HBE cells, we examined the effects of MAPK inhibitors selective for JNK, MEK1, and p38 on H₂O₂ and/or IFN γ induced IL-32 mRNA expression in HBE cells (Figures 2A, B, and 2C). JNK inhibitor exerted an inhibitory effect on both IFN γ alone (Figure 2B) and IFN γ combined with H₂O₂ induced IL-32 mRNA expression (Figures 2A and 2B). However, neither MEK1 inhibitor nor p38 inhibitor affected the mRNA expression of IL-32 (Figure 2A).

There was no effect of JAK inhibitor I on both IFN γ alone and IFN γ combined with H₂O₂ induced IL-32 mRNA expression (Figure 2C).

Stimulation of HBE cells by H₂O₂ followed by IFN γ promotes binding of transcription factors to the IL-32 promoter

As shown in Figure 3A, c-Jun binding to the IL-32 promoter was only increased by the combined treatment of H₂O₂ followed by IFN γ , but not with H₂O₂ or IFN γ alone.

CREB binding to the IL-32 promoter was significantly increased in the IFN γ alone as well as with H₂O₂ followed by IFN γ (Figure 3B). As expected, H₂O₂ alone did not increase CREB binding to the IL-32 promoter.

RNA polymerase II binding to the IL-32 promoter was increased in the IFN γ treated cells and was further increased in the H₂O₂+IFN γ stimulated HBE cells (Figure 3C). These results are consistent with our results regarding the transcriptional activity of IL-32.

Mutations in the c-Jun/CREB binding site of the IL-32 promoter largely suppressed its transcriptional activity

To confirm the roles of c-Jun and CREB in IL-32 gene transcription, which were indicated by ChIP assays, the transcriptional activity of IL-32 promoter with mutations in the c-Jun/CREB binding site was investigated. Compared to the wild-type promoter, the transcriptional activity of the mutant promoter was significantly reduced even in unstimulated HBE cells (control) and, while the wild-type promoter activity was increased by the combined stimuli of H₂O₂ followed by IFN γ no increase was found with promoter mutated at the c-Jun/CREB binding site, neither with H₂O₂, IFN γ alone, nor with the combination (Figure 4).

Knockdown of CREB resulted in significant suppression of IL-32 induction by oxidative stress and IFN γ

Knock down of c-Jun and CREB mRNA expressions by each siRNA transfection in HBE cells were confirmed by real time-PCR. Both were successfully suppressed up to 10% (Figures 5A and 5B, respectively). Although

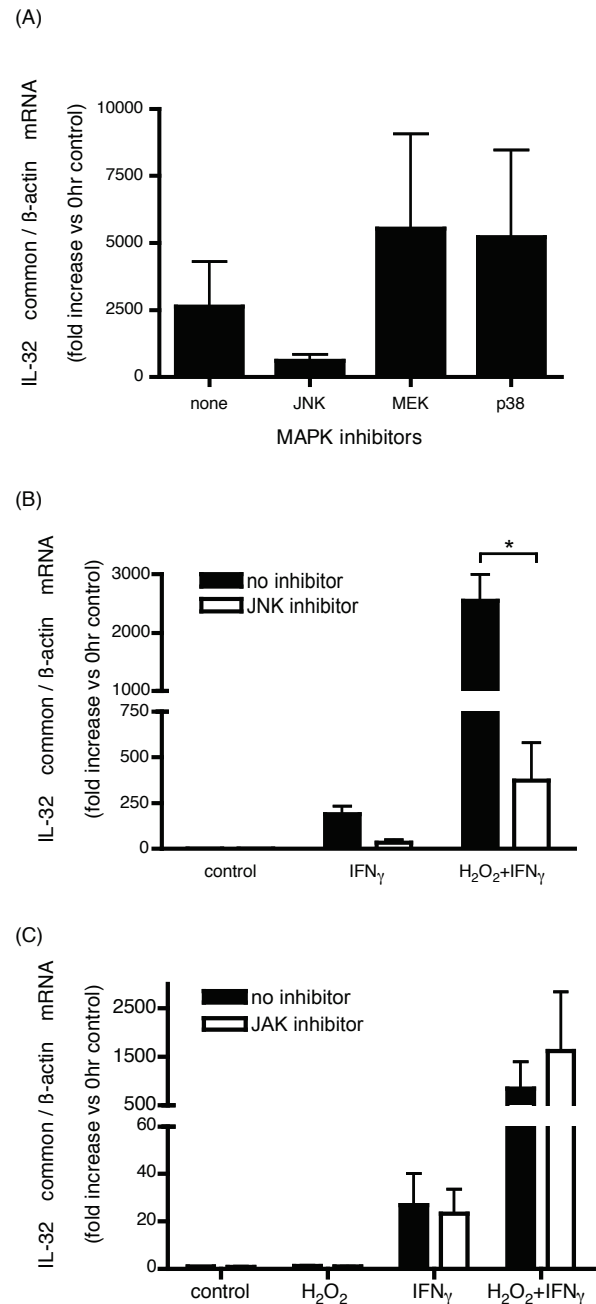


Figure 2 Influence of MAPK inhibitors on H₂O₂ + IFN γ induced IL-32 mRNA expression in HBE cells. After treatment with the JNK inhibitor, the MEK inhibitor, the p38 inhibitor for 24 hours, IL-32 mRNA expression in H₂O₂ and IFN γ stimulated HBE cells (A) and the effect of JNK inhibitor (B) or JAK inhibitor (C) upon IL-32 expression stimulated by IFN γ with or without H₂O₂ in HBE cells were examined by quantitative real-time PCR. All mRNA quantities were adjusted to the quantities at 0 hour control without stimulation. In graph (B) and (C), the closed bars represent the results of vehicle control and the open bars represent the results of JNK inhibitor (B) and JAK inhibitor (C). The bars show the means \pm SE from 3 different individuals. *p < 0.05 significantly different.

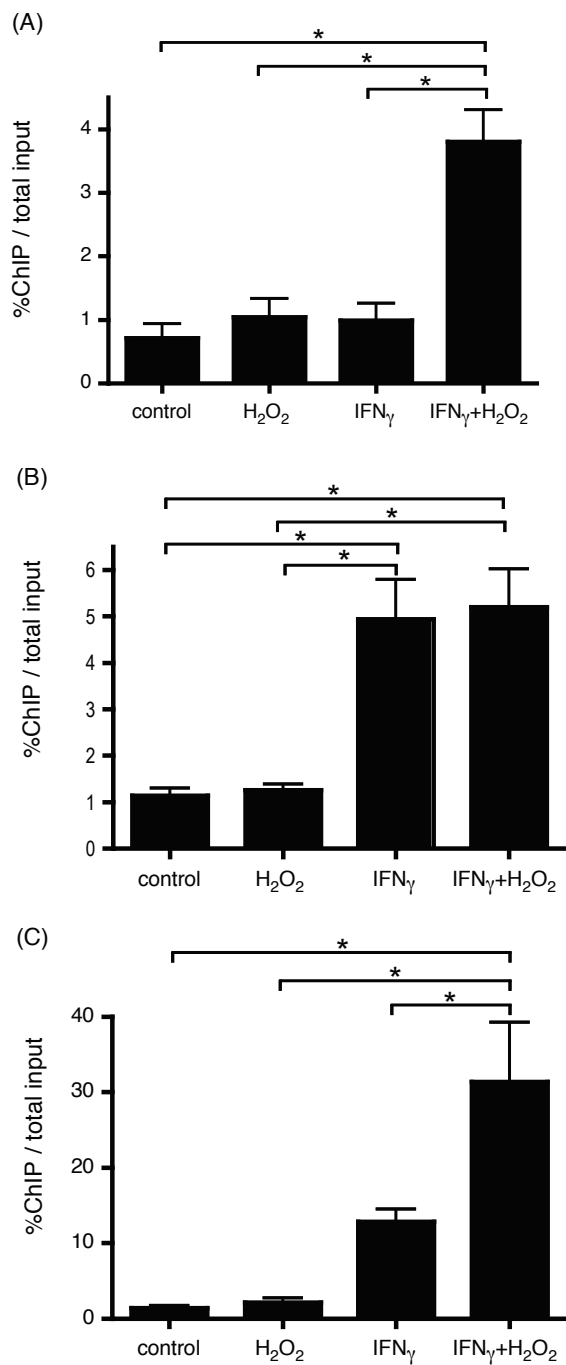


Figure 3 Transcription factors that bind to the IL-32 gene promoter. HBE cells were stimulated with H₂O₂, IFN_γ, H₂O₂ + IFN_γ, or vehicle for 30 minutes. The ChIP assay was performed to identify which transcription factors bind to the IL-32 gene promoter. Binding activity was compared using quantitative real-time PCR of the IL-32 promoter in DNA from chromatin complexes immunoprecipitated by antibodies to c-Jun (A), CREB (B), and RNA polymerase II (C). The bars show the means ± SE from 3 different individuals. *p < 0.01 significantly different.

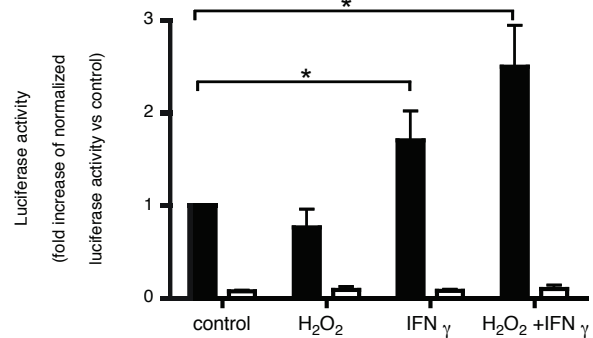
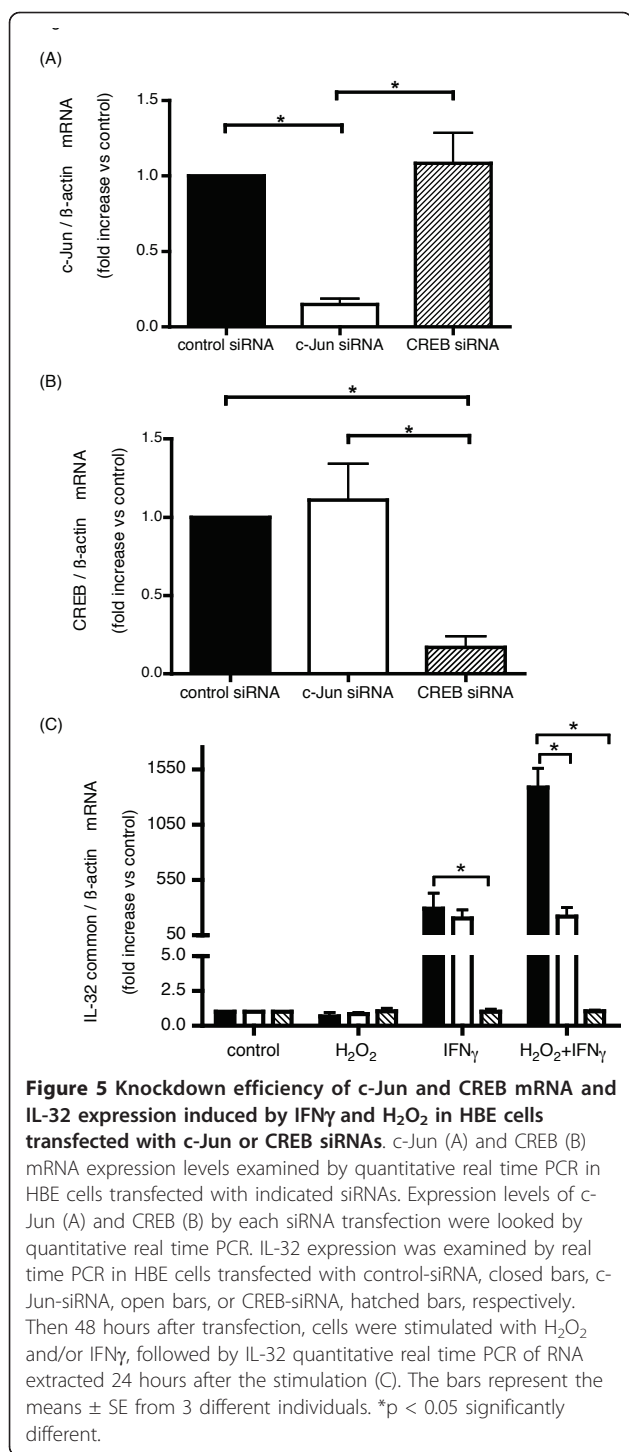


Figure 4 Transcriptional activity of promoter of IL-32 gene after IFN_γ or H₂O₂ + IFN_γ stimulation. HBE cells were transfected with a luciferase promoter vector without mutations (pWild-Luc), closed bars, or the same vector with mutations in the c-Jun/CREB binding site (pMutant-Luc), open bars. Cells were stimulated with H₂O₂, IFN_γ, H₂O₂ + IFN, or vehicle. Six hours after indicated stimulation, the cells were lysed and luciferase activity was measured. Luciferase activity in the cells was normalized to Renilla luciferase activity. The bars represent the means ± SE from 3 different individuals. *p < 0.05 significantly different.

knockdown of c-Jun did not influence IL-32 induction by IFN_γ alone, as shown in Figure 5C, it significantly suppressed H₂O₂ + IFN_γ induced IL-32 expression. On the other hand, knocking down of CREB resulted in significant suppression of IL-32 expression after stimulation with IFN_γ alone and also with H₂O₂ + IFN_γ compared with control.

Discussion

In this study, we revealed that IFN_γ upregulated IL-32 mRNA expression in HBE cells. Incubation with H₂O₂ alone did not upregulate its expression; however, pretreatment with H₂O₂ augmented IFN_γ induced IL-32 mRNA and expression in HBE cells. And IL-32 induction was suppressed by JNK inhibition but not by MEK inhibition or p38 inhibition. IFN_γ + H₂O₂ stimulated HBE cells, indicating expression induction of IL-32 by IFN_γ is regulated by signal pathway involving JNK and independent of p38 or MEK. JNK inhibitor also inhibited IFN_γ induced IL-32. JAK inhibitor I did not affect both on IFN_γ alone and IFN_γ + H₂O₂ induced IL-32. H₂O₂ alone did not increase c-Jun binding to the c-Jun binding site in the IL-32 promoter region. Binding of c-Jun was increased when HBE cells were stimulated with both H₂O₂ and IFN_γ. On the other hand, CREB was able to bind to the IL-32 promoter after IFN_γ stimulation with or without H₂O₂. Mutations in the c-Jun and CREB binding sites in the IL-32 promoter region inhibited the promoter activity induced by IFN_γ with or without H₂O₂. Further, knocking down of c-Jun resulted in



suppressed induction of IL-32 mRNA expression by H₂O₂ + IFN γ and knocking out of CREB resulted in suppressed induction by IFN γ alone and by H₂O₂ + IFN γ . This indicates that c-Jun and CREB binding to the promoter are the key mechanisms of IL-32 induction by H₂O₂ and IFN γ .

Several previous reports have indicated that IL-32 expression was upregulated by *Mycobacterium tuberculosis* infection or LPS in peripheral blood mononuclear cells and was also upregulated by influenza and HIV virus infection in the A549 and HEK293T human embryonic kidney cell lines, respectively. In addition, LPS and phorbol myristate acetate induced IL-32 expression in a leukemia cell line and in endothelial cells [23-26]. Recently, Li et al. reported the regulatory effect of influenza A virus upon IL-32 expression, indicating that CREB and NF- κ B are the key molecules in the induction of IL-32 expression [27]. No previous reports have determined the regulatory effect of oxidative stress on IL-32 expression. In this study, oxidative stress; i.e., H₂O₂ treatment, did not affect IL-32 expression alone, but it did augment IFN γ -induced IL-32 expression in HBE cells.

Increased oxidative stress, an important aggravating factor of the disease, is persistent not only in the lungs of currently smoking COPD patients but also in patients who have achieved smoking cessation for years [10,28]. On the other hand, IFN γ is known to be a representative cytokine of CD8+ T cells and is also associated with viral infection. Even in patients with stable COPD, IFN γ levels were reported to be increased. Furthermore, viral infection is a major cause of COPD exacerbation [29,30]. Exacerbations appear to accelerate the decreasing of lung function in COPD [31]. Taken together with our findings that H₂O₂ did not affect IL-32 expression alone but did augment IFN γ -induced IL-32 expression, IL-32 could be induced higher in airways of COPD patients who have increased oxidative stress under the exacerbation caused by viral infection and/or an inflammatory condition in which CD8+ T cells are activated. This suggests a possible mechanism for the increased expression of IL-32 in severe COPD patients compared to mild COPD patients and non-COPD smokers, as Calabrese F et al. reported [19].

The mechanisms regulating IL-32 expression have been examined in several reports using vascular endothelial cells, synovial fibroblasts, and pancreatic cancer cell lines [13,32,33]. We investigated the mechanism regulating IL-32 expression in airway epithelial cells to clarify whether the same pathways are involved or whether characteristic features are seen according to cell type and to examine whether the blockade of certain signal pathways results in reduced expression of IL-32.

We have searched transcription factor binding sites existing on IL-32 promoter using sequence retrieval software TFSEARCH^(TM) [34], and two adjacent binding sites that could be responsible for the downstream of JNK signaling pathway are those of c-Jun and CREB, which are located between nucleotides -30 to -23 and

-34 to -19, respectively, as a transcription start site at +1 (Figure 6). As shown in Figure 6, there are consensus binding sites of ATF and NF κ B other than c-Jun and CREB on IL-32 promoter region. The cellular signaling pathway induced by IFN γ has been investigated in bronchial epithelial cell line (BEAS-2B) [35], which revealed that the signal was dependent on IKKB1/2 but not the NF κ B pathway. In addition, IL-32 expression was not suppressed by an inhibitor of p38 which is supposed to be one of the kinases upstream of NF κ B. For these reasons, NF κ B was thought to be not associated with IL-32 expression induced by IFN γ (Figure 2A).

Further, a previous report showed that deletions of the IL-32 promoter sequence, including one of the consensus ATF binding sites, resulted in no decline in transcription activity [13]. Taking these results into consideration, we focused instead in our study on c-Jun and CREB as key molecules regulating IL-32 transcription induced by IFN γ and oxidative stress.

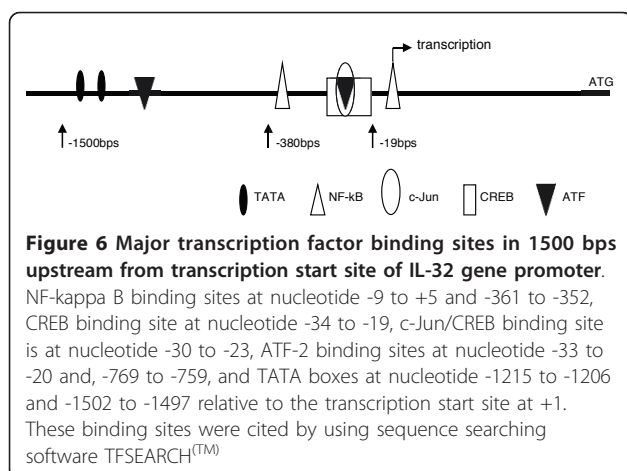
In HBE cells, c-Jun does not bind to the IL-32 promoter in the absence of IFN γ . Although the phosphorylation of c-Jun after H₂O₂ stimulation has been fully investigated in previous reports [36-38], it is unclear whether H₂O₂ induces or suppresses c-Jun binding to a specific gene promoter. Kumar et al. revealed that H₂O₂ directly suppressed AP-1 binding to the eNOS promoter [39]. Our results suggested c-Jun could be responsible for the transcription activity of IL-32 when HBE cells were stimulated by H₂O₂ + IFN γ .

On the contrary, CREB bound to IL-32 promoter after IFN γ stimulation with or without H₂O₂. CREB activation by IFN γ has been reported previously in murine macrophages [40,41], but a larger number of reports have indicated that IFN γ has a suppressive effect on CREB activation [42,43]. There are no reports about H₂O₂ activating CREB binding to gene promoters. In our study, CREB was bound to the IL-32 promoter after

IFN γ treatment but not after treatment with H₂O₂ alone. The activation and binding of CREB to the IL-32 promoter by IFN γ is one possible mechanism of IL-32 gene regulation in HBE cells and another is that IFN γ indirectly affects CREB activation through coactivators (e.g., CBP/p300) that are induced and activated by IFN γ [44] or other transcription factors including STAT-1 [45,46], which is the main downstream effector of IFN γ stimulation. IFN γ is known to exert its effect through not only JAK/STAT signal pathway but also through MAPK signaling pathway [47,48]. Some reports indicated presence of MAPK signal transduction by IFN γ which is not suppressed by inhibiting JAK/STAT [49], and Kim HA et al found IFN γ signal exerted by activation and upregulation of CREB but is not influenced by knocking out of STAT [40]. These research findings support our results that IL-32 expression regulation by IFN γ can be dependent on JNK (and its downstream c-Jun and CREB) but be independent of JAK/STAT (Figure 2C). Although no binding sites for STAT or interferon regulatory factor (IRF) were detected, but CREB binding site was detected at least within 1500 bps from transcription start site of IL-32 promoter, further investigation with the full length promoter will be needed to confirm whether the JAK/STAT pathways are involved or not.

As shown in Figure 4, mutations in the c-Jun/CREB binding sites resulted in markedly reduced transcription of IL-32, even after H₂O₂ and/or IFN γ stimulation. This is consistent with a previous report investigating the regulatory mechanism of IL-32 transcription in endothelial cells by Kobayashi et al. [13]. They demonstrated that the deletion of the IL-32 promoter between nucleotides 26 and 100 upstream of the transcription initiation site, identical to the region containing the CREB and c-Jun binding sequences, led to significantly suppressed transcription activity. Further, knocking down of CREB by siRNA transfection resulted in a significant suppression of the IL-32 expression that was induced by both IFN γ alone and IFN γ + H₂O₂ down to the baseline control levels. CREB is a key transcription factor for IL-32 transcription in HBE cells. This finding is also compatible with the report by Li et al. showing that mutations in the binding site for CREB or CREB knockdown resulted in the significant suppression of influenza A virus-induced IL-32 transcription [27]. In addition, decreased expression of c-Jun by c-Jun siRNA resulted the significant suppression of IL-32 induction by IFN γ + H₂O₂, though IL-32 induction by H₂O₂ or IFN γ alone was not affected (Figure 5C). These findings suggested synergistic effect of c-Jun inducing IL-32 expression by IFN γ under oxidative stress.

Although p38 is suggested to be one of MAPKs located upstream of AP-1 (including c-Jun), we found



no effect of p38 inhibitor on IL-32 induction by IFN γ in HBE cells. This could be explained by number of reports showing that signal transduction induced by cytokines such as IFN γ or oxidizing substance that involve AP-1 was dependent of JNK but was independent of p38 in HBE cells and macrophages [50,51]

Putting the results of our experiments together, we suggest that IFN γ induced CREB binding to the IL-32 promoter, which was followed by an increase in the transcription of IL-32, and an additive effect of c-Jun binding to the IL-32 promoter by H₂O₂ resulted in a further acceleration of IL-32 transcription. How IFN γ affects c-Jun binding to the promoter has not been clarified. One possibility is that it involves a coactivator, e.g. CBP/p300, which is known to bind to AP-1 similar to c-Jun and promotes transcriptional activity [52]. CBP/p300 could be activated by IFN γ [49], and CREB could form a heterodimer with c-Jun [53], which may explain the necessity of IFN γ for the binding of c-Jun to the IL-32 promoter during H₂O₂ stimulation. In addition, both activated c-Jun and CREB binding to the IL-32 promoter are necessary to induce significant transcription of IL-32 in IFN γ and H₂O₂ stimulated HBE cells.

The role of increased IL-32 expression in airway epithelial cells in the pathogenesis of COPD has not been clarified, and furthermore, it is still unclear whether suppressing its expression is beneficial with regards to preventing disease progression or improving the symptoms of the disease. IL-32 has several roles including inducing the expression of inflammatory cytokines and adhesion molecules in T-lymphocytes, monocytes, macrophages, and epithelial cells and promoting monocyte differentiation into macrophages, which alters the responses of inflammatory cells against infection [54,55]. Also, proteinase-3, which is activated by neutrophilic inflammation, cleaves IL-32 into its highly activated form [17,56]. Thus, increased IL-32 expression may modify airway inflammation in COPD. Further investigations to identify the roles of IL-32 in COPD are necessary.

As IL-32 is expressed in several types of cells, the mechanism of IL-32 gene regulation suggested in this study may not only be applicable to the pathogenesis of COPD but also those of other inflammatory diseases associated with oxidative stress. Further studies are needed.

Abbreviations

IL-32: interleukin-32; COPD: chronic obstructive pulmonary disease; HBE: human bronchial epithelial cell; ChIP: chromatin immunoprecipitation; CCL: CC chemokine ligand; CXCL: CXC chemokine ligand; TSLP: thymic stromal lymphopoietin; GM-CSF: granulocyte macrophage colony stimulating factor; LPS: lipopolysaccharide; FEV₁: forced expiratory volume in one second; DMSO: dimethyl sulfoxide; PCR: polymerase chain reaction; RT-PCR: Reverse

Transcription -polymerase chain reaction; eNOS: endothelial nitric oxide synthase.

Acknowledgements

We thank Ms. Aya Inazumi who helped us to maintain cell cultures and to prepare samples for Western blot analysis.

Author details

¹Department of Respiratory Medicine, Kyoto University Graduate School of Medicine, Kyoto, Japan. ²Health Administration Center and Division of Respiratory Medicine, Department of Medicine, Shiga University of Medical Science, Otsu, Shiga, Japan. ³Department of thoracic surgery, Graduate School of Medicine, Kyoto University, Kyoto, Japan. ⁴Health Administration Center, Shiga University of Medical Science, Seta-Tsukinowa-cho, Otsu, Shiga 520-2192, Japan.

Authors' contributions

MK and EO performed the whole experiments. DK, AH, TT, NT and SM contributed in collecting clinical data and samples. HS and HD contributed in providing surgery samples. EO designed the study. DK, AH, TT, NT, YH, TH, SM, and MM advised the design of the study and participated in the analysis of the data. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 20 December 2011 Accepted: 14 March 2012

Published: 14 March 2012

References

1. Barnes PJ: The cytokine network in chronic obstructive pulmonary disease. *Am J Respir Cell Mol Biol* 2009, **41**(6):631-638.
2. Cosio MG, Saetta M, Agusti A: Immunologic aspects of chronic obstructive pulmonary disease. *N Engl J Med* 2009, **360**(23):2445-2454.
3. Lopez AD, Shibuya K, Rao C, Mathers CD, Hansell AL, Held LS, Schmid V, Buist S: Chronic obstructive pulmonary disease: current burden and future projections. *Eur Respir J* 2006, **27**(2):397-412.
4. Kouzaki H, O'Grady SM, Lawrence CB, Kita H: Proteases induce production of thymic stromal lymphopoietin by airway epithelial cells through protease-activated receptor-2. *J Immunol* 2009, **183**(2):1427-1434.
5. Proud D, Chow CW: Role of viral infections in asthma and chronic obstructive pulmonary disease. *Am J Respir Cell Mol Biol* 2006, **35**(5):513-518.
6. Richter A, O'Donnell RA, Powell RM, Sanders MW, Holgate ST, Djukanovic R, Davies DE: Autocrine ligands for the epidermal growth factor receptor mediate interleukin-8 release from bronchial epithelial cells in response to cigarette smoke. *Am J Respir Cell Mol Biol* 2002, **27**(1):85-90.
7. Takizawa H, Tanaka M, Takami K, Ohtoshi T, Ito K, Satoh M, Okada Y, Yamasawa F, Nakahara K, Umeda A: Increased expression of transforming growth factor-beta1 in small airway epithelium from tobacco smokers and patients with chronic obstructive pulmonary disease (COPD). *Am J Respir Crit Care Med* 2001, **163**(6):1476-1483.
8. Ying S, O'Connor B, Ratoff J, Meng Q, Fang C, Cousins D, Zhang G, Gu S, Gao Z, Shamji B, et al: Expression and cellular provenance of thymic stromal lymphopoietin and chemokines in patients with severe asthma and chronic obstructive pulmonary disease. *J Immunol* 2008, **181**(4):2790-2798.
9. Nakayama T, Church DF, Pryor WA: Quantitative analysis of the hydrogen peroxide formed in aqueous cigarette tar extracts. *Free Radic Biol Med* 1989, **7**(1):9-15.
10. Rahman I: Oxidative stress in pathogenesis of chronic obstructive pulmonary disease: cellular and molecular mechanisms. *Cell Biochem Biophys* 2005, **43**(1):167-188.
11. Dahl CA, Schall RP, He HL, Cairns JS: Identification of a novel gene expressed in activated natural killer cells and T cells. *J Immunol* 1992, **148**(2):597-603.
12. Joosten LA, Netea MG, Kim SH, Yoon DY, Oppers-Walgreen B, Radstake TR, Barrera P, van de Loo FA, Dinarello CA, van den Berg WB: IL-32, a proinflammatory cytokine in rheumatoid arthritis. *Proc Natl Acad Sci USA* 2006, **103**(9):3298-3303.

13. Kobayashi H, Lin PC: **Molecular characterization of IL-32 in human endothelial cells.** *Cytokine* 2009, **46**(3):351-358.
14. Shioya M, Nishida A, Yagi Y, Ogawa A, Tsujikawa T, Kim-Mitsuyama S, Takayanagi A, Shimizu N, Fujiyama Y, Andoh A: **Epithelial overexpression of interleukin-32alpha in inflammatory bowel disease.** *Clin Exp Immunol* 2007, **149**(3):480-486.
15. Goda C, Kanaji T, Kanaji S, Tanaka G, Arima K, Ohno S, Izuhara K: **Involvement of IL32 in activation-induced cell death in T cells.** *Int Immunol* 2006, **18**(2):233-240.
16. Netea MG, Lewis EC, Azam T, Joosten LA, Jaekel J, Bae SY, Dinarello CA, Kim SH: **Interleukin-32 induces the differentiation of monocytes into macrophage-like cells.** *Proc Natl Acad Sci USA* 2008, **105**(9):3515-3520.
17. Novick D, Rubinstein M, Azam T, Rabinkov A, Dinarello CA, Kim SH: **Proteinase 3 is an IL-32 binding protein.** *Proc Natl Acad Sci USA* 2006, **103**(9):3316-3321.
18. Shoda H, Fujio K, Yamaguchi Y, Okamoto A, Sawada T, Kochi Y, Yamamoto K: **Interactions between IL-32 and tumor necrosis factor alpha contribute to the exacerbation of immune-inflammatory diseases.** *Arthritis Res Ther* 2006, **8**(6):R166.
19. Calabrese F, Baraldo S, Bazzan E, Lunardi F, Rea F, Maestrelli P, Turato G, Lokar-Oliani K, Papi A, Zuin R, et al: **IL-32, a novel proinflammatory cytokine in chronic obstructive pulmonary disease.** *Am J Respir Crit Care Med* 2008, **178**(9):894-901.
20. Higashimoto Y, Elliott WM, Behzad AR, Sedgwick EG, Takei T, Hogg JC, Hayashi S: **Inflammatory mediator mRNA expression by adenovirus E1A-transfected bronchial epithelial cells.** *Am J Respir Crit Care Med* 2002, **166**(2):200-207.
21. Kim SH, Han SY, Azam T, Yoon DY, Dinarello CA: **Interleukin-32: a cytokine and inducer of TNFalpha.** *Immunity* 2005, **22**(1):131-142.
22. Cevik MO, Katsuyama M, Kanda S, Kaneko T, Iwata K, Ibi M, Matsuno K, Kakehi T, Cui W, Sasaki M, et al: **The AP-1 site is essential for the promoter activity of NOX1/NADPH oxidase, a vascular superoxide-producing enzyme: Possible involvement of the ERK1/2-JunB pathway.** *Biochem Biophys Res Commun* 2008, **374**(2):351-355.
23. Kang JW, Choi SC, Cho MC, Kim HJ, Kim JH, Lim JS, Kim SH, Han JY, Yoon DY: **A proinflammatory cytokine interleukin-32beta promotes the production of an anti-inflammatory cytokine interleukin-10.** *Immunology* 2009, **128**(1 Suppl):e532-540.
24. Li W, Liu Y, Mukhtar MM, Gong R, Pan Y, Rasool ST, Gao Y, Kang L, Hao Q, Peng G, et al: **Activation of interleukin-32 pro-inflammatory pathway in response to influenza A virus infection.** *PLoS One* 2008, **3**(4):e1985.
25. Netea MG, Azam T, Lewis EC, Joosten LA, Wang M, Langenberg D, Meng X, Chan ED, Yoon DY, Ottenhoff T, et al: **Mycobacterium tuberculosis induces interleukin-32 production through a caspase-1/IL-18/interferon-gamma-dependent mechanism.** *PLoS Med* 2006, **3**(8):e277.
26. Rasool ST, Tang H, Wu J, Li W, Mukhtar MM, Zhang J, Mu Y, Xing HX, Wu J, Zhu Y: **Increased level of IL-32 during human immunodeficiency virus infection suppresses HIV replication.** *Immunol Lett* 2008, **117**(2):161-167.
27. Li W, Sun W, Liu L, Yang F, Li Y, Chen Y, Fang J, Zhang W, Wu J, Zhu Y: **IL-32: a host proinflammatory factor against influenza viral replication is upregulated by aberrant epigenetic modifications during influenza A virus infection.** *J Immunol* 2010, **185**(9):5056-5065.
28. Louhelainen N, Rytala P, Haahela T, Kinnula VL, Djukanovic R: **Persistence of oxidant and protease burden in the airways after smoking cessation.** *BMC Pulm Med* 2009, **9**:25.
29. Anzueto A, Sethi S, Martinez FJ: **Exacerbations of chronic obstructive pulmonary disease.** *Proc Am Thorac Soc* 2007, **4**(7):554-564.
30. Moermans C, Heinen V, Nguyen M, Henket M, Sele J, Manise M, Corhay JL, Louis R: **Local and systemic cellular inflammation and cytokine release in chronic obstructive pulmonary disease.** *Cytokine* 2011, **56**(2):298-304.
31. Kanner RE, Anthonisen NR, Connett JE: **Lower respiratory illnesses promote FEV(1) decline in current smokers but not ex-smokers with mild chronic obstructive pulmonary disease: results from the lung health study.** *Am J Respir Crit Care Med* 2001, **164**(3):358-364.
32. Mun SH, Kim JW, Nah SS, Ko NY, Lee JH, Kim JD, Kim do K, Kim HS, Choi JD, Kim SH: **Tumor necrosis factor alpha-induced interleukin-32 is positively regulated via the Syk/protein kinase Cdelta/JNK pathway in rheumatoid synovial fibroblasts.** *Arthritis Rheum* 2009, **60**(3):678-685, 3.
33. Nishida A, Andoh A, Inatomi O, Fujiyama Y: **Interleukin-32 expression in the pancreas.** *J Biol Chem* 2009, **284**(26):17868-17876.
34. Heinemeyer T, Wingender E, Reuter I, Hermjakob H, Kel AE, Kel OV, Ignatieva EV, Ananko EA, Podkolodnaya OA, Kolpakov FA, et al: **Databases on transcriptional regulation: TRANSFAC, TRRD and COMPEL.** *Nucleic Acids Res* 1998, **26**(1):362-367.
35. Tudhope SJ, Catley MC, Fenwick PS, Russell RE, Rumsey WL, Newton R, Barnes PJ, Donnelly LE: **The role of I kappa B kinase 2, but not activation of NF-kappa B, in the release of CXCR3 ligands from IFN-gamma-stimulated human bronchial epithelial cells.** *J Immunol* 2007, **179**(9):6237-6245.
36. Aggeli IK, Gaitanaki C, Beis I: **Involvement of JNKs and p38-MAPK/MSK1 pathways in H₂O₂-induced upregulation of heme oxygenase-1 mRNA in H9c2 cells.** *Cell Signal* 2006, **18**(10):1801-1812.
37. Teng CH, Huang WN, Meng TC: **Several dual specificity phosphatases coordinate to control the magnitude and duration of JNK activation in signaling response to oxidative stress.** *J Biol Chem* 2007, **282**(39):28395-28407.
38. Zhang S, Lin Y, Kim YS, Hande MP, Liu ZG, Shen HM: **c-Jun N-terminal kinase mediates hydrogen peroxide-induced cell death via sustained poly (ADP-ribose) polymerase-1 activation.** *Cell Death Differ* 2007, **14**(5):1001-1010.
39. Kumar S, Sun X, Wedgwood S, Black SM: **Hydrogen peroxide decreases endothelial nitric oxide synthase promoter activity through the inhibition of AP-1 activity.** *Am J Physiol Lung Cell Mol Physiol* 2008, **295**(2):L370-377.
40. Kim HA, Jeon SH, Seo GY, Park JB, Kim PH: **TGF-beta1 and IFN-gamma stimulate mouse macrophages to express BAFF via different signaling pathways.** *J Leukoc Biol* 2008, **83**(6):1431-1439.
41. Liu L, Wang Y, Fan Y, Li CL, Chang ZL: **IFN-gamma activates cAMP/PKA/CREB signaling pathway in murine peritoneal macrophages.** *J Interferon Cytokine Res* 2004, **24**(6):334-342.
42. Hu X, Chen J, Wang L, Ivashkiv LB: **Crosstalk among Jak-STAT, Toll-like receptor, and ITAM-dependent pathways in macrophage activation.** *J Leukoc Biol* 2007, **82**(2):237-243.
43. Hu X, Paik PK, Chen J, Yarinina A, Kockeritz L, Lu TT, Woodgett JR, Ivashkiv LB: **IFN-gamma suppresses IL-10 production and synergizes with TLR2 by regulating GSK3 and CREB/AP-1 proteins.** *Immunity* 2006, **24**(5):563-574.
44. Ahmad R, Qureshi HY, El Mabrouk M, Sylvester J, Ahmad M, Zafarullah M: **Inhibition of interleukin 1-induced matrix metalloproteinase 13 expression in human chondrocytes by interferon gamma.** *Ann Rheum Dis* 2007, **66**(6):782-789.
45. Strassheim D, Riddle SR, Burke DL, Geraci MW, Stenmark KR: **Prostacyclin inhibits IFN-gamma-stimulated cytokine expression by reduced recruitment of CBP/p300 to STAT1 in a SOCS-1-independent manner.** *J Immunol* 2009, **183**(11):6981-6988.
46. Zhang JJ, Vinkemeier U, Gu W, Chakravarti D, Horvath CM, Darnell JE Jr: **Two contact regions between Stat1 and CBP/p300 in interferon gamma signaling.** *Proc Natl Acad Sci USA* 1996, **93**(26):15092-15096.
47. Du Z, Wei L, Murti A, Pfeffer SR, Fan M, Yang CH, Pfeffer LM: **Non-conventional signal transduction by type 1 interferons: the NF-kappa B pathway.** *J Cell Biochem* 2007, **102**(5):1087-1094.
48. Pokrovskaja K, Panaretakis T, Grander D: **Alternative signaling pathways regulating type I interferon-induced apoptosis.** *J Interferon Cytokine Res* 2005, **25**(12):799-810.
49. Li H, Gade P, Xiao W, Kalvakolanu DV: **The interferon signaling network and transcription factor C/EBP-beta.** *Cell Mol Immunol* 2007, **4**(6):407-418.
50. Blanchette J, Abu-Dayyeh I, Hassani K, Whitcombe L, Olivier M: **Regulation of macrophage nitric oxide production by the protein tyrosine phosphatase Src homology 2 domain phosphotyrosine phosphatase 1 (SHP-1).** *Immunology* 2009, **127**(1):123-133.
51. Zhang H, Forman HJ: **Signaling pathways involved in phase II gene induction by alpha, beta-unsaturated aldehydes.** *Toxicol Ind Health* 2009, **25**(4-5):269-278.
52. Kamei Y, Xu L, Heinzl T, Torchia J, Kurokawa R, Gloss B, Lin SC, Heyman RA, Rose DW, Glass CK, et al: **A CBP integrator complex mediates transcriptional activation and AP-1 inhibition by nuclear receptors.** *Cell* 1996, **85**(3):403-414.
53. Benbrook DM, Jones NC: **Heterodimer formation between CREB and JUN proteins.** *Oncogene* 1990, **5**(3):295-302.
54. Bai X, Kim SH, Azam T, McGibney MT, Huang H, Dinarello CA, Chan ED: **IL-32 is a host protective cytokine against Mycobacterium tuberculosis in**

differentiated THP-1 human macrophages. *J Immunol* 2010, **184**(7):3830-3840.

55. Netea MG, Azam T, Ferwerda G, Girardin SE, Walsh M, Park JS, Abraham E, Kim JM, Yoon DY, Dinarello CA, *et al*: IL-32 synergizes with nucleotide oligomerization domain (NOD) 1 and NOD2 ligands for IL-1beta and IL-6 production through a caspase 1 dependent mechanism. *Proc Natl Acad Sci USA* 2005, **102**(45):16309-16314.
56. Choi JD, Bae SY, Hong JW, Azam T, Dinarello CA, Her E, Choi WS, Kim BK, Lee CK, Yoon DY, *et al*: Identification of the most active interleukin-32 isoform. *Immunology* 2009, **126**(4):535-542.

doi:10.1186/1465-9921-13-19

Cite this article as: Kudo *et al.*: Oxidative stress induced Interleukin-32 mRNA expression in human bronchial epithelial cells. *Respiratory Research* 2012 **13**:19.

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