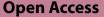
RESEARCH



Principal component analysis of flow-volume curves in COPDGene to link spirometry with phenotypes of COPD



Kenneth Verstraete^{1,2}, Nilakash Das¹, Iwein Gyselinck¹, Marko Topalovic³, Thierry Troosters⁴, James D. Crapo⁵, Edwin K. Silverman⁶, Barry J. Make⁵, Elizabeth A. Regan⁵, Robert Jensen⁷, Maarten De Vos^{2,8} and Wim Janssens^{1*}

Abstract

Background Parameters from maximal expiratory flow-volume curves (MEFVC) have been linked to CT-based parameters of COPD. However, the association between MEFVC shape and phenotypes like emphysema, small airways disease (SAD) and bronchial wall thickening (BWT) has not been investigated.

Research question We analyzed if the shape of MEFVC can be linked to CT-determined emphysema, SAD and BWT in a large cohort of COPDGene participants.

Study design and methods In the COPDGene cohort, we used principal component analysis (PCA) to extract patterns from MEFVC shape and performed multiple linear regression to assess the association of these patterns with CT parameters over the COPD spectrum, in mild and moderate-severe COPD.

Results Over the entire spectrum, in mild and moderate-severe COPD, principal components of MEFVC were important predictors for the continuous CT parameters. Their contribution to the prediction of emphysema diminished when classical pulmonary function test parameters were added. For SAD, the components remained very strong predictors. The adjusted R² was higher in moderate-severe COPD, while in mild COPD, the adjusted R² for all CT outcomes was low; 0.28 for emphysema, 0.21 for SAD and 0.19 for BWT.

Interpretation The shape of the maximal expiratory flow-volume curve as analyzed with PCA is not an appropriate screening tool for early disease phenotypes identified by CT scan. However, it contributes to assessing emphysema and SAD in moderate-severe COPD.

Keywords COPD, Computed tomography, Maximal expiratory flow-volume curve, Principal component analysis

*Correspondence:

Wim Janssens

wim.janssens@uzleuven.be

¹ Laboratory of Respiratory Diseases and Thoracic Surgery (BREATHE), Department of Chronic Diseases and Metabolism, KU Leuven, Herestraat 49, O&N 1Bis, Box 706, 3000 Leuven, Belgium

² STADIUS Center for Dynamical Systems, Signal Processing and Data Analytics, Department of Electrical Engineering (ESAT), KU Leuven, Leuven, Belgium

³ ArtiQ NV, Leuven, Belgium

⁵ National Jewish Medical and Research Center, Denver, CO, USA

⁶ Channing Division of Network Medicine, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, MA, USA

⁷ University of Utah, Salt Lake City, Utah, USA

⁸ Department of Development and Regeneration, KU Leuven, Leuven, Belgium



© The Author(s) 2023. Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativeco mmons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

⁴ Department of Rehabilitation Sciences, KU Leuven, Leuven, Belgium

Introduction

Chronic obstructive pulmonary disease (COPD) is often diagnosed after significant loss of lung function, as symptoms can remain mild to absent, and are often neglected by patients in the early disease stages. COPD is presumed to start as a smoldering disease, with small airways and parenchymal damage accumulating for many years without being noticed by patients or physicians [1, 2]. The ability to identify COPD in the early stage is key in the appropriate management of the disease aimed at improving patient outcomes, as well as reducing overall costs [3]. Spirometry is currently put forward as the most appropriate diagnostic tool, as it is non-invasive, easy to perform, and implementable at low cost. The spirometry diagnosis of COPD is based on a post-bronchodilator forced expiratory volume in one second/forced vital capacity (FEV1/FVC) ratio below the lower limit of the reference population in a clinical context of exposure to noxious particles [4].

A reduced forced expiratory flow between 25 and 75% of FVC (FEF25-75) has been proposed as a sign of small airways disease, in smokers only at risk of developing COPD [5, 6]. Moreover, recent large population studies in smoking individuals demonstrate that early pathological changes visualized on CT may also occur in subjects with 'normal' spirometry [7]. Normal, if not only defined by the FEV1/FVC ratio, is outlined by spirometry parameters varying within the range of a healthy non-smoking reference group [8, 9]. Even within the range of normality, the shape or contour of the maximal expiratory flowvolume curve (MEFVC) has been of continuous interest [10]. The concavity of the curve, often referred to as the kink, has been associated with emphysema and attributed to airway collapse and loss of elastic recoil [11]. Topalovic et al. proposed the angle of collapse of MEFVC to quantify airway collapse and detecting CT-defined emphysema in heavy smokers [12]. Dominelli et al. quantified the shape of MEFVC with the slope ratio index [13]. Bhatt et al. later proposed the parameter D which describes lung volume as an exponential function of time and the peak index, modeling the number of peaks adjusted for lung size [14, 15]. An overview of all indices can be found in the comprehensive review by Hoesterey et al. [16]. In this review, it has been postulated that further analysis on the shape of MEFVC yields the potential to discover parameters that can help detect early airway obstruction [16].

In a large subgroup of the Genetic Epidemiology of COPD study (COPDGene), we used principal component analysis (PCA) to comprehensively characterize the shape of MEFVC and linked the PCA components to CTbased parameters in subjects with mild and moderatesevere airflow obstruction.

Study design and methods Study subjects

We used subjects enrolled in the COPDGene study, which is a large US-based multicenter study including current and former smokers aged 45–80 years (n = 10,198) with at least ten pack-years. Details of the study design have been reported previously [17]. The study was approved by local Institutional Review boards at each of the 21 clinical centers and all subjects provided written documentation of informed consent. The available data included raw spirometry and CT imaging data. For this analysis, we split the subjects on stages of the Global Initiative for Chronic Obstructive Lung Diseases (GOLD) guidelines according to FEV1, FVC and FEV1/ FVC. GOLD I subjects belonged to the mild stage group while GOLD II-III-IV subjects belonged to the moderatesevere stage group.

Spirometry and CT imaging data

Using a standardized protocol [18] and spirometer (NDD EasyOne Spirometer), 9841 participants performed spirometry. Expiratory flow-volume curves and volumetime curves were available. CT scans were obtained at total lung capacity (TLC) and at the end of normal expiration (functional residual capacity, FRC) using multidetector CT scanners. CT densitometry was used to define the presence of emphysema and Small Airways Disease (SAD). Both %emphysema and %gas-trapping were computed using parametric response mapping (PRM) to identify the extent of emphysema (PRM^{emph}) and functional small airways disease (PRM^{fSAD}) based on CT scans at TLC and FRC simultaneously [19, 20]. Bronchial Wall Thickening (BWT) was assessed by airway wall thickness at an internal perimeter of 10 mm (Pi10). Pi10 was calculated by fitting a linear regression model on all airways of different internal perimeters with the square root of the wall area as dependent variable and perimeter as independent variable. Quantitative parameters of these scans were extracted using Thirona software.

Shape analysis

To focus purely on the shape of MEFVC, we scaled each curve in both axes by 1/FVC for each subject to normalize on FVC and to preserve the shape of the curves. To perform a shape analysis, we applied PCA on the curves (flow over volume datapoints) to extract the most dominant patterns ordered according to the proportion in shape variance they explain. Each MEFVC could then be accurately approximated as a linear combination of these principal components (PC) or patterns with the coefficients describing how much each pattern contributed to the shape of the MEFVC. We computed these coefficients for all subjects in the dataset and linked these to the continuous CT parameters. We denoted the PCs following their order, e.g., the first PC was denoted as PC1. A more extensive description of the PCA computation can be found in the online supplement.

CT-based phenotypes

With the quantitative CT (QCT) values, we defined the presence of emphysema, SAD and BWT using the upper limit of normal (95th percentile, ULN) cut-offs based on never-smoked normal control subjects in the COPDGene dataset, 107 of such control subjects were enrolled in Phase 1. Based on these cut-offs, we defined eight CT-based phenotypes according to the presence of emphysema and/or SAD and/or BWT. For notation of the phenotypes, emphysema, SAD and BWT were denoted as E, S and B, respectively. A dash was used in the absence of a disease. An overview of all notations can be found in Table 1. We compared the PRM cut-offs with the ULN cut-offs when the %voxels < - 950 Hounsfield Units (Hu) and %voxels < - 856 Hu definitions for emphysema and SAD, respectively, were used on the same never-smoked normal subjects.

Data and statistical analysis

We performed descriptive statistics on demographic, spirometric and CT variables per GOLD stage and per CT-based phenotype. The data is presented as no. (%) or median [Q1-Q3 interquartile range]. Multiple linear regression was used to assess the independent effect of each component in predicting PRM^{emph}, PRM^{fSAD} and Pi10 with adjustment for age, sex, height, weight and pack-years. Standard spirometric parameters (FEV1, FVC, FEV1/FVC, PEF, and FEF25-75) were then added and the standardized β coefficients of the model were used to assess the importance of each predictor. We used the adjusted R², the coefficient of determination,

 Table 1
 Table with the used abbreviations of CT-defined phenotypes

	Emphysema	Small airways disease	Bronchial wall thickening
	_	_	_
E	✓	-	-
-S-	-	✓	-
B	-	-	1
ES-	✓	1	-
-SB	-	1	1
E-B	✓	-	1
ESB	1	1	\checkmark

B, bronchial wall thickening; E, emphysema; S, small airways disease

to evaluate the goodness-of-fit of the models. Regression analysis was done over the entire spectrum, in mild COPD (GOLD I) and moderate-severe COPD (GOLD II-III-IV). We compared the principal components with existing MEFVC parameters: angle of collapse [21], area under the forced expiratory flow-volume loop [22], obstructive index [23] and peak index [15]. Statistical analysis was conducted using Python 3 (Python Software Foundation) with the scientific and statistical packages SciPy and StatsModels (open source, scipy.org and statsmodels.org), significance level was set at 0.05.

Results

Population characteristics

Of the 9841 patients that performed spirometry, 9207 (93.6%) had acceptable flow-volume loops according to the American Thoracic Society (ATS)/European Respiratory Society (ERS) guidelines [18]. Subjects with Preserved Ratio Impaired Spirometry (PRISm, FEV1/ FVC > = 0.7 but FEV1 < 80%, n = 1055) were not considered, since other disease factors such as thoracic wall restriction or cardiac disease, being more prevalent in this subgroup, would influence our findings [24, 25]. Ultimately, 6302 subjects were used for the analysis. The flow of the eligible subjects for this analysis is described in Additional file 1: Figure S1. The characteristics of the remaining participants per GOLD stage are reported in Table 1. Sixty-seven of 107 never-smoked control subjects had both spirometry and CT data available. In the 6302 subjects used for the analysis, 67 were non-smokers and 6235 were ever smokers. Of those 6235 ever smokers, 3214 were former smokers and 3021 were current smokers.

Principal components

The mean standardized flow-volume curves per GOLD stage are visualized in Fig. 1. The curves were sampled at 200 equidistant points resulting in 200 principal components (full decomposition) with the first ten explaining 78% of the variance in MEFVC shape (Fig. 2A). With the first 100 components, 98.4% of the variance could be explained. To visualize the influence of the components on MEFVC, we depicted a -45 to +45 percent change (5th to 95th percentile) of the four most dominant components as compared to the overall mean MEFVC of the population (Fig. 2B). We visually assessed the influence of each of these four components on the MEFVC: PC1 influences PEF and the descending limb without altering the angle of collapse or concavity. PC2 pivots the descending limb around a fixed point, thereby also influencing PEF. PC3 and PC4 mainly model concavity in MEFVC. The remainder of the analyses were done with the first four principal components since more

7	67	2842	567	1483	892	451
Age (years)	59.5 [53.9–66.8]	55.6 [50.2-63.0]	61.9 [55.0–68.9]	62.6 [55.9–69.4]	65.2 [58.7–71.1]	64.4 [59.2–69.8]
Sex (%Males)	23 (34%)	1434 (50%)	323 (57%)	782 (53%)	522 (59%)	269 (60%)
Weight (kg)	81.0 [66.0–88.3]	82.7 [71.1–95.2]	77.6 [67.0–87.0]	82.0 [69.2–94.8]	80.0 [65.4–94.0]	72.2 [60.2-84.6]
Height (cm)	166.4 [159.8–173.4]	170.0 [163.0-177.0]	170.0 [162.6–177.8]	170.0 [162.9–176.6]	169.9 [162.6–177.0]	170.2 [163.0–176.6]
BMI (kg/m2)	27.5 [24.5–31.7]	28.3 [25.0-32.5]	26.5 [23.7–29.8]	27.9 [24.5–32.1]	27.1 [23.6–31.7]	24.8 [21.6–28.8]
Pack-years	0.0 [0.0-0.0]	34.2 [22.6–46.3]	40.8 [30.0-54.5]	45.5 [33.6-64.5]	49.5 [38.0-69.0]	49.5 [38.0-74.5]
FEV1 (L)	2.8 [2.3–3.4]	2.8 [2.4–3.3]	2.6 [2.2–3.1]	1.8 [1.5–2.2]	1.1 [0.9–1.3]	0.7 [0.5-0.8]
FEV1 (%Pred)	103.4 [93.8-115.6]	95.9 [88.5–104.4]	88.3 [84.2–95.2]	65.6 [57.7–72.0]	40.5 [35.4-45.0]	23.6 [19.0–26.7]
FVC (L)	3.6 [2.9–4.3]	3.6 [3.0–4.3]	4.1 [3.4–4.8]	3.1 [2.6–3.8]	2.6 [2.1–3.2]	2.1 [1.7–2.6]
FVC (%Pred)	98.5 [90.5–109.3]	95.5 [88.4–104.1]	105.3 [98.5–112.7]	85.2 [77.5–93.2]	70.7 [62.3–79.0]	55.5 [46.9–65.7]
FEV1/FVC	0.8 [0.8-0.8]	0.8 [0.7-0.8]	0.7 [0.6-0.7]	0.6 [0.5–0.7]	0.4 [0.4–0.5]	0.3 [0.3-0.4]
PEF (L/s)	7.8 [6.5–9.2]	7.4 [6.2–9.0]	6.9 [5.6–8.7]	5.2 [4.2-6.5]	3.4 [2.7–4.3]	2.3 [1.8–2.9]
FEF25-75 (L/s)	3.1 [2.3–3.6]	2.6 [2.0–3.3]	1.3 [0.9–1.6]	0.7 [0.5–1.0]	0.4 [0.3–0.4]	0.2 [0.2-0.3]
%Emphysema	0.2 [0.0–0.5]	0.2 [0.1–0.8]	1.5 [0.4–4.3]	2.7 [0.7–8.2]	13.6 [4.8–24.8]	27.0 [16.6–39.2]
%Gas Trapping	6.5 [4.3–10.0]	6.6 [3.1–11.2]	13.9 [8.0–21.5]	19.6 [11.8–28.5]	33.0 [25.0–39.5]	36.9 [31.7–43.5]
Pi10 (mm)	1.7 [1.5–1.9]	1.9 [1.7–2.2]	2.0 [1.8–2.4]	2.5 [2.2–2.9]	2.8 [2.5–3.2]	2.9 [2.5–3.2]

 Table 2
 Characteristics per GOLD stage

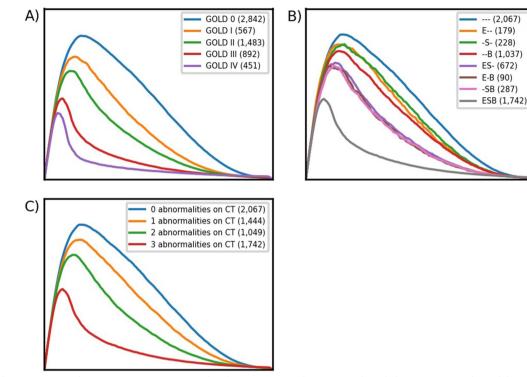


Fig. 1 The mean MEFVC shapes per GOLD stage in panel **A**, the mean MEFVC shapes per CT-based phenotype in panel **B** and the mean MEFVC shapes per number of abnormalities on CT in panel **C**. B, bronchial wall thickening; E, emphysema; GOLD, Global Initiative for Chronic Obstructive Lung Disease; MEFVC, maximal expiratory flow-volume curve; S, small airways disease

components did not improve the model fits (adjusted R²) in the following analyses.

Multivariate analysis on PRM^{emph}

When only considering principal components and adjusting for age, sex, height, weight and pack-years, PC1 ($\beta = -4.9$, P < 0.001), PC2 ($\beta = -4.3$, P < 0.001) and PC3 ($\beta = -1.4$, P < 0.001) were significant predictors in the entire COPDGene population. Adjusted R² was 0.50. In mild COPD, PC1 ($\beta = -1.6$, P < 0.001), PC2 ($\beta = -1.4$, P < 0.001) and PC3 ($\beta = 0.7$, P = 0.001) were significant predictors and adjusted R² was 0.17. In moderate-severe COPD, PC1 ($\beta = -2.5$, P < 0.001), PC2 ($\beta = -7.3$, P < 0.001), PC3 ($\beta = 2.3$, P < 0.001) and PC4 ($\beta = 0.9$, P = 0.001) were significant predictors with an adjusted R² of 0.50. Full results can be found in Table 3.

When adding standard spirometric parameters tests to the model, the principal components, PC1 (β =0.7, P<0.01), PC2 (β =0.7, P<0.01) and PC3 (β =-0.4, P<0.01) were still significantly associated in the entire COPDGene population. The adjusted R² was 0.67. In mild COPD, adjusted R² was 0.28 with FEV1 the dominant predictor (β =-10.7, P<0.001). PC4 was a significant predictor (β =-0.5, P=0.04). In moderate-severe COPD, adjusted R² was 0.59 with FEV1/FVC the dominant predictor ($\beta = -11.3$, *P*<0.001), none of the components were significantly associated. Full results can be found in Additional file 1: Table S1.

Multivariate analysis on PRM^{fSAD}

Adjusting for age, sex, height, weight and pack-years and only considering principal components, PC1 (β =-7.1, *P*<0.001), PC2 (β =-5.1, *P*<0.001), PC3 (β =1.1, *P*<0.001) and PC4 (β =-0.7, *P*<0.001) were significant predictors. Adjusted R² was 0.60. In mild COPD, PC1 (β =-2.3, *P*<0.001), PC2 (β =-2.6, *P*<0.001) and PC3 (β =0.9, *P*=0.03) were significant predictors while adjusted R² was 0.2. In moderate-severe COPD, PC1 (β =-2.5, *P*<0.001), PC2 (β =-6.8, *P*<0.001), PC3 (β =2.4, *P*<0.001) and PC4 (β =0.7, *P*=0.01) were significant predictors with an adjusted R² of 0.48. Full results can be found in Table 4.

When adding standard spirometric parameters to the model, the principal components PC1 (β =-2.2, P<0.001), PC2 (β =-1.5, P<0.001) and PC3 (β =0.7, P<0.001) were still significant predictors. Adjusted R² over the entire population was 0.65. In mild COPD, PC1, PC2 and PC4 were significant predictors (β =-1.4, -2.4 and -1.1, P=0.03, P=0.004 and P=0.02). In moderate-severe COPD, PC1, PC2 and PC3 were significant

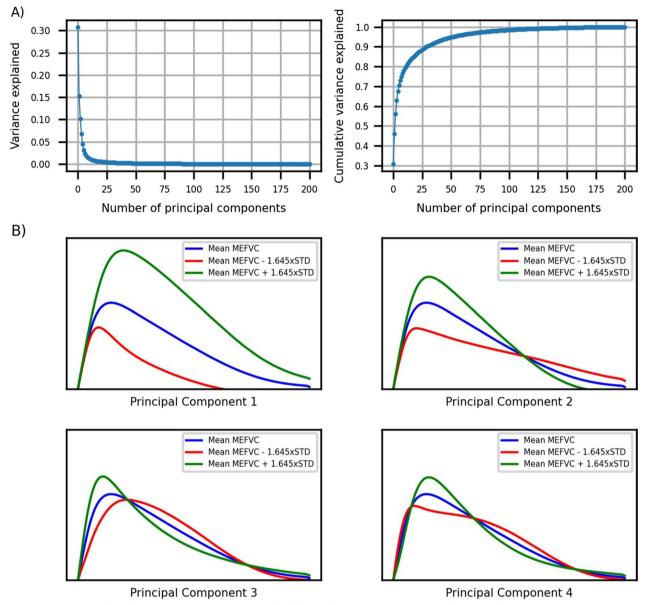


Fig. 2 A Variance explained by the principal components on the left, cumulative variance explained by the principal components on the right. **B** influence of the first four principal components visualized. The blue curve is the overall mean MEFVC shape. The green curve illustrates the influence of each principal component when the coefficient is increased to the 95th percentile, the red curve when the coefficient is decreased to the 5th percentile. Component 1 influences PEF and the descending limb without altering the angle of collapse or concavity. Component 2 pivots the descending limb around a fixed point, thereby also influencing PEF. Component 3 and 4 mainly model concavity in MEFVC

predictors ($\beta = -1.4, -5.9$ and 2.5, *P*<0.001, *P*<0.001 and *P*<0.001) with an adjusted R² of 0.49. Full results can be found in Additional file 1: Table S2.

Multivariate analysis on BWT

Only considering principal components and after adjusting for age, sex, height, weight and pack-years, all four PCs were significant predictors over the entire spectrum ($\beta = -0.35$, $\beta = -0.19$, $\beta = 0.045$ and

 β =0.02, *P*<0.001, respectively) and adjusted R² was 0.39. In mild COPD, PC1 and PC2 were significant predictors (β =-0.06 and β =-0.06, *P*=0.04 and *P*=0.1, respectively) with adjusted R² 0.14 while in moderate-severe COPD, PC2, PC3 and PC4 were significant predictors (β =-0.13, β =-0.03 and β =-0.05, *P*<0.001, *P*=0.03 and *P*=0.001, respectively) with an adjusted R² of 0.16. Full results can be found in Additional file 1: Table S3.

	Entire spectrum (n= 6302)		Mild COPD (n $=$ 567)	(n = 567)	Moderate–severe COPD (n=2826)	(n=2826)
	Adjusted $R^2 = 0.50$		Adjusted $R^2 = 0.17$	=0.17	Adjusted $R^2 = 0.50$	
	β	<i>P</i> -value	β	<i>P</i> -value	β	<i>P</i> -value
Age	1.32	< 0.001	1.08	< 0.001	1.40	< 0.001
Sex	- 0.58	< 0.001	- 0.70	0.01	- 0.06	0.80
Height	0.63	< 0.001	0.38	0.16	1.71	< 0.001
Weight	- 1.80	< 0.001	- 0.91	< 0.001	- 3.18	< 0.001
Pack-years	0.17	0.09	0.35	0.05	- 0.12	0.51
PC1	— 4.87	< 0.001	- 1.55	< 0.001	— 2.49	< 0.001
PC2	- 4.34	< 0.001	- 1.42	< 0.001	- 7.28	< 0.001
PC3	— 0.05	0.61	0.70	0.001	2.25	< 0.001
PC4	- 1.37	< 0.001	0.01	0.97	06.0	0.001
COPD, Chronic	COPD, Chronic Obstructive Pulmonary Disease; PC, principal component	PC, principal component				

PRM ^{emph}
for
analysis ⁻
Multivariate a
ŝ
Table

	Entire spectrum (n = 6302)		Mild COPD (n = 567)		Moderate—severe COPD (n = 2826)	
	Adjusted R ² =	= 0.60	Adjusted R ² =	= 0.20	Adjusted R ² =0.48	
	β	P-value	β	<i>P</i> -value	β	P-value
Age	2.73	< 0.001	3.33	< 0.001	2.13	< 0.001
Sex	- 1.24	< 0.001	- 1.12	0.03	- 0.96	< 0.001
Height	0.63	< 0.001	1.21	0.03	0.70	0.01
Weight	- 1.53	< 0.001	- 1.68	< 0.001	- 1.93	< 0.001
Pack-years	0.61	< 0.001	0.30	0.40	0.56	0.002
PC1	- 7.14	< 0.001	- 2.35	< 0.001	- 2.54	< 0.001
PC2	- 5.10	< 0.001	- 2.63	< 0.001	- 6.79	< 0.001
PC3	1.11	< 0.001	0.88	0.03	2.68	< 0.001
PC4	- 0.66	< 0.001	- 0.52	0.17	0.69	0.01

Table 4	Multivariate	analysis	for PRM ^{fSAD}
---------	--------------	----------	-------------------------

COPD, Chronic Obstructive Pulmonary Disease; PC, principal component

Again, the principal components were of little benefit on top of classical pulmonary function variables with an adjusted R² over the entire population of 0.48 and with PC1, PC3 and P4 as significant predictors (β =-0.04, β =0.04 and β =0.02, *P*=0.02, *P*<0.001 and *P*=0.004, respectively). In mild COPD, adjusted R² was 0.19 and no component was a significant predictor. In moderatesevere COPD, adjusted R² was 0.23 with PC2 and PC3 significant predictors (β =-0.11 and β =0.05, *P*=0.001 and *P*=0.02, respectively). Full results can be found in Additional file 1: Table S4.

CT-Phenotypes

We determined 1.7% for PRM^{emph}, 14.7% for PRM^{fSAD} and 2.2 mm for Pi10 (Fig. 3) as the upper limit of normal

(ULN) in a cohort of never-smoked normal controls (n=67) and considered them as cut-offs for the presence of CT-based abnormalities. Figure 1B shows the mean MEFVC per CT-based phenotype and Fig. 1C the mean MEFVC per number of abnormalities as seen on CT. The characteristics of the subjects per CT-based phenotype are reported in Additional file 1: Table S5. The cut-offs for emphysema and SAD were 5.8 and 18.6% when the classic %voxels < -950 and %voxels < -856 definitions for emphysema and SAD were used.

Comparison with other MEFVC-derived parameters

Adjusted R^2 for each parameter per CT outcome and subgroup can be found in Additional file 1: Table S6. Compared to other MEFVC-derived parameters, the

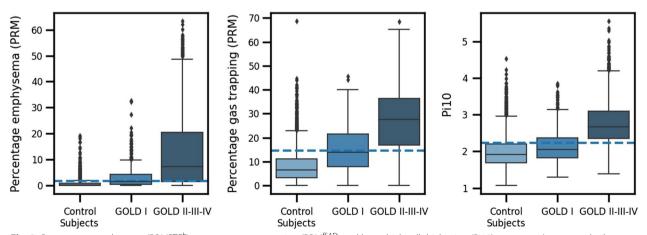


Fig. 3 Percentage emphysema (PRM^{emph}), percentage gas trapping (PRM^{fSAD}) and bronchial wall thickening (Pi10) per group (never-smoked normal control subjects; mild COPD (GOLD I); moderate-severe COPD (GOLD II-III-IV)). The cut-offs (dashed lines) are determined by using the 95th percentile (upper limit of normal) on the control subjects

principal components provided a better fit for PRM^{emph} and PRM^{SAD}. For Pi10, area under the forced expiratory flow-volume loop was the superior parameter. Overall, the classical pulmonary function parameters were superior.

Discussion

In 6302 subjects in the COPDGene study, we used principal component analysis (PCA) to extract dominant patterns from the shapes of the MEFVC and explored their association with continuous CT-based parameters and eight CT-defined phenotypes based on cut-offs for emphysema, small airways disease and bronchial wall thickening. The advantage of this PCA analysis is that no hand-engineered features were required to analyze the MEFVC and the large collection of curves was fully exploited in extracting potential patterns. When compared with existing hand-engineered features, the principal components were superior for emphysema and small airways disease and closely matched the area under the MEFVC for bronchial wall thickening.

We found that a small number of components were sufficient to model a large proportion of the variance in shape of MEFVC. Multivariate analysis for the first four principal components showed that 49, 60 and 39 percent of the variance could be explained for emphysema (PRMemph), small airways disease (PRMfSAD) and bronchial wall thickening (Pi10), respectively. However, when adding classical pulmonary function tests (FEV1, FVC, FEV1/FVC, PEF and FEF25-75) to the models, independent contributions of the principal components were strongly reduced because of high intra-correlations (Additional file 1: Table S7). For emphysema (PRM^{emph}), shape-derived components PC1, PC2 and PC3 were still independent contributors. For small airways disease (PRM^{fSAD}) in mild COPD, PC2 was the third important predictor, whilst it became the most important predictor in moderate-severe COPD for which FEV1, FVC and FEV1/FVC were no longer significant. These findings highlight the impact of small airways disease on PEF, particularly in more advanced disease stages. For bronchial wall thickening, the fit of the regression model was generally very low, indicating that presence of abnormal thickening of the larger airway bronchial walls, did not profoundly affect the shape of the MEFVC.

Interestingly, in multinomial logistic regression modelling the number of CT disease abnormalities present on CT rather the type of CT abnormalities (E, SAD, BWT), the pseudo R^2 was negative and not significant for mild COPD, indicating that in the mild disease stage, any CT abnormalities are unlikely to be detected by the shape of MEFVC or even the standard lung function parameters. Hence, the current data suggest that our initial hypothesis should be rejected, and that early disease processes as identified on CT cannot be predicted by parameters isolated from the relative form of the maximal expiratory flow-volume curve. In particular, FEF25-75, as surrogate marker of small airways disease on spirometry was not predictive in the mild COPD subgroup. It raises the question if in patients with normal spirometry, risk behavior and chronic respiratory symptoms may point to the need of a CT scan, as suggested by Celli et al. [26].

By using the upper limit of normal on the 67 neversmoked normal control subjects in this cohort, we obtained cut-offs for abnormal values of the CT outcomes. With these cut-off values, most of the patients in the mild COPD subgroup had PRM^{emph} and PRM^{fSAD} values within the normal range (Fig. 3). It also demonstrates that mild airflow limitation as diagnosed by an FEV1/FVC below 0.7, can present with CT scans within normal limits. In these individuals, early airway pathology in terminal or respiratory bronchioles may still be present as this is beyond the resolution of conventional CT [27]. An alternative explanation may come from the initial lung function values determined by lung growth, which may result in a lower FEV1/FVC ratio and FEV1 without true pathology on CT.

We normalized the MEFVC curve for FVC to adjust for lung volume and hence also anthropometry and age, and to maximally visualize the changes in shape across the different phenotypes. Next, we calculated the mean MEFVC shape per GOLD stage and per CT-based phenotype. The area under the curve decreases and the concavity or so-called kink in the curve increases as lung function deteriorates. For the CT-based phenotypes, the mean shapes are similar for the phenotypes with only one abnormality [--B, -S-, E--], while for those with two abnormalities [-SB, E-B, ES-], the concavity is larger and the area under the curve smaller, which is to be expected as these are the subjects in the higher GOLD stages. Subjects showing evidence of three abnormalities on CT have the largest concavity and the smallest area under the curve on average. Overall, our findings indicate that concavity of the flow-volume loop is linked to more advanced COPD in which emphysema, but also other radiological phenotypes co-occur.

Interpretation

Our analysis demonstrates that the shape of the maximal expiratory flow-volume curve is not an appropriate screening tool for early disease phenotypes identified by CT scan since neither the principal components and classical pulmonary function parameters were linked with emphysema, small airways disease or bronchial wall thickening as seen on CT. In moderate-severe airflow obstruction (GOLD II-III-IV) the concavity of the curve is mainly related to the presence of emphysema in a combined phenotype with small airways disease, with MEFVC shape parameters having a limited but statistically significant association with CT defined pathologies.

Abbreviations

ATS	American Thoracic Society
BWT	Bronchial wall thickening
COPD	Chronic obstructive pulmonary disease
CT	Computed tomography
emph	Emphysema
ERS	European Respiratory Society
FEF	Forced expiratory flow
FEV1	Forced expiratory volume in 1 second
FRC	Functional residual capacity
fSAD	Functional Small Airways Disease
FVC	Forced vital capacity
GOLD	Global Initiative for Chronic Obstructive Lung Disease
Hu	Hounsfield unit
MEFVC	Maximal Expiratory Flow-volume Curve
PC	Principal component
PCA	Principal component analysis
PEF	Peak expiratory flow
Pi10	Airway wall thickness at an internal perimeter of 10 mm
PRISm	Preserved ratio impaired spirometry
PRM	Parametric response mapping
SAD	Small Airways Disease
TLC	Total lung capacity
ULN	Upper limit of normal

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s12931-023-02318-4.

Additional file 1: Table S1. Multivariate analysis for PRM^{emph} with pulmonary function test parameters. Table S2. Multivariate analysis for PRM^{ISAD} with pulmonary function test parameters. Table S3. Multivariate analysis for Pi10. Table S4. Multivariate analysis for Pi10 with pulmonary function test parameters. Table S5. Characteristics per CT-based phenotype. Table S6. Linear regression adjusted R²s for different parameters derived from maximal expiratory flow-volume curves for emphysema, small airways disease and bronchial wall thickening (PRM^{emph}, PRM^{ISAD}, Pi10 on CT) per subgroup. Table S7. Pearson correlation coefficients between the first four principal components and the classical pulmonary function parameters FEV1, FVC, FEV1/FVC, FEF25-75, PEF. Figure S1. Flow of eligible subjects for this study. CT, computed tomography; COPDGene, Genetic Epidemiology of COPD; PRISm, preserved ratio impaired spirometry.

Acknowledgements

Financial/nonfinancial disclosures: KV has nothing to disclose. ND has nothing to disclose. IG receives personal funding from Research Foundation Flanders (FWO). MT is CEO and co-founder of ArtiQ but received no payments related to the manuscript. TT has nothing to disclose. JDC received funding from NIH Grant Support R01-089897 and is chairman of the COPD Foundation Board of Directors. EKS received funding from NIH Grant Support and institutional grant support from GlaxoSmithKline and Bayer. BJM received funding from NHLBI, grants from AstraZeneca, GlaxoSmithKline and Pearl Research and fees from AstraZeneca, GlaxoSmithKline, Sunovion, Verona, Boehringer Ingelheim, Takeda, Third Pole, Phillips and Circasia. He is member of the NHLBI and Spiration Data Safety Monitoring Boards and chairman of Mt. Sinai SOM Data Safety Monitoring Board. RJ received consulting fees from National Jewish Health for an LLC he owns. MDV received funding from the AI in Flanders project, WJ received grants from AstraZeneca and Chiesi and obtained fees from AstraZeneca, Chiesi and GlaxoSmithKline. He is chairman of Board of Flemish Society for TBC prevention and board member of ArtiQ.

Role of the sponsors: None of the sponsors had input or contributions related to the study and manuscript.

Other contributions

Grant support and disclaimer The project described was supported by Award Number U01 HL089897 and Award Number U01 HL089856 from the National Heart, Lung, and Blood Institute. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Heart, Lung, and Blood Institute or the National Institutes of Health. COPD Foundation Funding

COPDGene is also supported by the COPD Foundation through contributions made to an Industry Advisory Board that has included AstraZeneca, Bayer Pharmaceuticals, Boehringer-Ingelheim, Genentech, GlaxoSmithKline, Novartis, Pfizer, and Sunovion.

COPDGene[®] Investigators – Core Units

Administrative Center: James D. Crapo, MD (PI); Edwin K. Silverman, MD, PhD (PI); Barry J. Make, MD; Elizabeth A. Regan, MD, PhD.

Genetic Analysis Center: Terri H. Beaty, PhD; Peter J. Castaldi, MD, MSc; Michael H. Cho, MD, MPH; Dawn L. DeMeo, MD, MPH; Adel El Boueiz, MD, MMSc; Marilyn G. Foreman, MD, MS; Auyon Ghosh, MD; Lystra P. Hayden, MD, MMSc; Craig P. Hersh, MD, MPH; Jacqueline Hetmanski, MS; Brian D. Hobbs, MD, MMSc; John E. Hokanson, MPH, PhD; Wonji Kim, PhD; Nan Laird, PhD; Christoph Lange, PhD; Sharon M. Lutz, PhD; Merry-Lynn McDonald, PhD; Dmitry Prokopenko, PhD; Matthew Moll, MD, MPH; Jarrett Morrow, PhD; Dandi Qiao, PhD; Elizabeth A. Regan, MD, PhD; Aabida Saferali, PhD; Phuwanat Sakornsakolpat, MD; Edwin K. Silverman, MD, PhD; Emily S. Wan, MD; Jeong Yun, MD, MPH. Imaging Center: Juan Pablo Centeno; Jean-Paul Charbonnier, PhD; Harvey O. Coxson, PhD; Craig J. Galban, PhD; MeiLan K. Han, MD, MS; Eric A. Hoffman, Stephen Humphries, PhD; Francine L. Jacobson, MD, MPH; Philip F. Judy, PhD; Ella A. Kazerooni, MD; Alex Kluiber; David A. Lynch, MB; Pietro Nardelli, PhD; John D. Newell, Jr., MD; Aleena Notary; Andrea Oh, MD; Elizabeth A. Regan, MD, PhD; James C. Ross, PhD; Raul San Jose Estepar, PhD; Joyce Schroeder, MD; Jered Sieren; Berend C. Stoel, PhD; Juerg Tschirren, PhD; Edwin Van Beek, MD, PhD; Bram van Ginneken, PhD; Eva van Rikxoort, PhD; Gonzalo Vegas Sanchez-Ferrero, PhD; Lucas Veitel; George R. Washko, MD; Carla G. Wilson, MS; PFT QA Center, Salt Lake City, UT: Robert Jensen, PhD.

Data Coordinating Center and Biostatistics, National Jewish Health, Denver, CO: Douglas Everett, PhD; Jim Crooks, PhD; Katherine Pratte, PhD; Matt Strand, PhD; Carla G. Wilson, MS.

Epidemiology Core, University of Colorado Anschutz Medical Campus, Aurora, CO: John E. Hokanson, MPH, PhD; Erin Austin, PhD; Gregory Kinney, MPH, PhD; Sharon M. Lutz, PhD; Kendra A. Young, PhD.

Mortality Adjudication Core: Surya P. Bhatt, MD; Jessica Bon, MD; Alejandro A. Diaz, MD, MPH; MeiLan K. Han, MD, MS; Barry Make, MD; Susan Murray, ScD; Elizabeth Regan, MD; Xavier Soler, MD; Carla G. Wilson, MS.

Biomarker Core: Russell P. Bowler, MD, PhD; Katerina Kechris, PhD; Farnoush Banaei- Kashani, PhD.

COPDGene[®] Investigators – Clinical Centers

Ann Arbor VA: Jeffrey L. Curtis, MD; Perry G. Pernicano, MD. Baylor College of Medicine, Houston, TX: Nicola Hanania, MD, MS; Mustafa Atik, MD; Aladin Boriek, PhD; Kalpatha Guntupalli, MD; Elizabeth Guy, MD; Amit Parulekar, MD;

Brigham and Women's Hospital, Boston, MA: Dawn L. DeMeo, MD, MPH; Craig Hersh, MD, MPH; Francine L. Jacobson, MD, MPH; George Washko, MD. Columbia University, New York, NY: R. Graham Barr, MD, DrPH; John Austin, MD; Belinda D'Souza, MD; Byron Thomashow, MD.

Duke University Medical Center, Durham, NC: Neil MacIntyre, Jr., MD; H. Page McAdams, MD; Lacey Washington, MD.

HealthPartners Research Institute, Minneapolis, MN: Charlene McEvoy, MD, MPH; Joseph Tashjian, MD.

Johns Hopkins University, Baltimore, MD: Robert Wise, MD; Robert Brown, MD; Nadia N. Hansel, MD, MPH; Karen Horton, MD; Allison Lambert, MD, MHS; Nirupama Putcha, MD, MHS.

Lundquist Institute for Biomedical Innovation at Harbor UCLA Medical Center, Torrance, CA: Richard Casaburi, PhD, MD; Alessandra Adami, PhD; Matthew Budoff, MD; Hans Fischer, MD; Janos Porszasz, MD, PhD; Harry Rossiter, PhD; William Stringer, MD.

Michael E. DeBakey VAMC, Houston, TX: Amir Sharafkhaneh, MD, PhD; Charlie Lan, DO.

Minneapolis VA: Christine Wendt, MD; Brian Bell, MD; Ken M. Kunisaki, MD, MS.

Morehouse School of Medicine, Atlanta, GA: Eric L. Flenaugh, MD; Hirut Gebrekristos, PhD; Mario Ponce, MD; Silanath Terpenning, MD; Gloria Westney, MD, MS.

National Jewish Health, Denver, CO: Russell Bowler, MD, PhD; David A. Lynch, MB.

Reliant Medical Group, Worcester, MA: Richard Rosiello, MD; David Pace, MD. Temple University, Philadelphia, PA: Gerard Criner, MD; David Ciccolella, MD; Francis Cordova, MD; Chandra Dass, MD; Gilbert D'Alonzo, DO; Parag Desai, MD; Michael Jacobs, PharmD; Steven Kelsen, MD, PhD; Victor Kim, MD; A. James Mamary, MD; Nathaniel Marchetti, DO; Aditi Satti, MD; Kartik Shenoy, MD; Robert M. Steiner, MD; Alex Swift, MD; Irene Swift, MD; Maria Elena Vega-Sanchez, MD.

University of Alabama, Birmingham, AL: Mark Dransfield, MD; William Bailey, MD; Surya P. Bhatt, MD; Anand Iyer, MD; Hrudaya Nath, MD; J. Michael Wells, MD. University of California, San Diego, CA: Douglas Conrad, MD; Xavier Soler, MD, PhD; Andrew Yen, MD.

University of Iowa, Iowa City, IA: Alejandro P. Comellas, MD; Karin F. Hoth, PhD; John Newell, Jr., MD; Brad Thompson, MD.

University of Michigan, Ann Arbor, MI: MeiLan K. Han, MD MS; Ella Kazerooni, MD MS; Wassim Labaki, MD MS; Craig Galban, PhD; Dharshan Vummidi, MD. University of Minnesota, Minneapolis, MN: Joanne Billings, MD; Abbie Begnaud, MD; Tadashi Allen, MD.

University of Pittsburgh, Pittsburgh, PA: Frank Sciurba, MD; Jessica Bon, MD; Divay Chandra, MD, MSc; Joel Weissfeld, MD, MPH.

University of Texas Health, San Antonio, San Antonio, TX: Antonio Anzueto, MD; Sandra Adams, MD; Diego Maselli-Caceres, MD; Mario E. Ruiz, MD; Harjinder Singh.

Take-home message

The shape of the maximal expiratory flow-volume curve is not an appropriate screening tool for early disease phenotypes identified by CT scan. However, it contributes to assessing emphysema and SAD in moderate-severe COPD.

Author contributions

KV, ND, IG, MT, MDV, and WJ contributed substantially to the study design, data analysis and interpretation, and writing of the manuscript. KV, ND, IG, MT, TT, JDC, EKS, BJM, EAR, RJ, MDV and WJ drafted the work or revised it critically for important intellectual content. KV, ND, IG, MT, TT, JDC, EKS, BJM, EAR, RJ, MDV and WJ provided final approval of the manuscript. KV, ND, IG, MT, TT, JDC, EKS, BJM, EAR, RJ, MDV and WJ agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All authors read and approved the final manuscript.

Funding

IG received personal funding (11N3922N) from Research Foundation Flanders (FWO). JDC received funding from NIH Grant Support R01-089897, EKS received funding from NIH Grant Support and institutional grant support from GlaxoSmithKline and Bayer. BJM received funding from NHLBI, grants from AstraZeneca, GlaxoSmithKline and Pearl Research and fees from AstraZeneca, GlaxoSmithKline, Sunovion, Verona, Boehringer Ingelheim, Takeda, Third Pole, Phillips and Circasia. ER received a COPDGene grant from NHBLI. RJ received consulting fees from National Jewish Health for an LLC he owns. MDV received funding from the AI in Flanders project. WJ received grants from AstraZeneca and Chiesi and obtained fees from AstraZeneca, Chiesi and GlaxoSmithKline.

Availability of data and materials

The dataset supporting the conclusions of this article is available by request through the COPDGene investigators (https://copdgene.org/).

Declarations

Ethics approval and consent to participate

The institutional review boards of all participating centres approved the COP-DGene study, and written informed consent was obtained from all subjects. The participating centres and its corresponding COPDGene investigators were:

Ann Arbor VA: Jeffrey L. Curtis, MD; Perry G. Pernicano, MD.

Baylor College of Medicine, Houston, TX: Nicola Hanania, MD, MS; Mustafa Atik, MD; Aladin Boriek, PhD; Kalpatha Guntupalli, MD; Elizabeth Guy, MD; Amit Parulekar, MD;

Brigham and Women's Hospital, Boston, MA: Dawn L. DeMeo, MD, MPH; Craig Hersh, MD, MPH; Francine L. Jacobson, MD, MPH; George Washko, MD. Columbia University, New York, NY: R. Graham Barr, MD, DrPH; John Austin,

MD; Belinda D'Souza, MD; Byron Thomashow, MD.

Duke University Medical Center, Durham, NC: Neil MacIntyre, Jr., MD; H. Page McAdams, MD; Lacey Washington, MD.

HealthPartners Research Institute, Minneapolis, MN: Charlene McEvoy, MD, MPH; Joseph Tashjian, MD.

Johns Hopkins University, Baltimore, MD: Robert Wise, MD; Robert Brown, MD; Nadia N. Hansel, MD, MPH; Karen Horton, MD; Allison Lambert, MD, MHS; Nirupama Putcha, MD, MHS.

Lundquist Institute for Biomedical Innovation at Harbor UCLA Medical Center, Torrance, CA: Richard Casaburi, PhD, MD; Alessandra Adami, PhD; Matthew Budoff, MD; Hans Fischer, MD; Janos Porszasz, MD, PhD; Harry Rossiter, PhD; William Stringer, MD.

Michael E. DeBakey VAMC, Houston, TX: Amir Sharafkhaneh, MD, PhD; Charlie Lan, DO.

Minneapolis VA: Christine Wendt, MD; Brian Bell, MD; Ken M. Kunisaki, MD, MS. *Morehouse School of Medicine, Atlanta, GA*: Eric L. Flenaugh, MD; Hirut Gebrekristos, PhD; Mario Ponce, MD; Silanath Terpenning, MD; Gloria Westney,

MD, MS. National Jewish Health, Denver, CO: Russell Bowler, MD, PhD; David A. Lynch, MB.

Reliant Medical Group, Worcester, MA: Richard Rosiello, MD; David Pace, MD. Temple University, Philadelphia, PA: Gerard Criner, MD; David Ciccolella, MD; Francis Cordova, MD; Chandra Dass, MD; Gilbert D'Alonzo, DO; Parag Desai, MD; Michael Jacobs, PharmD; Steven Kelsen, MD, PhD; Victor Kim, MD; A. James Mamary, MD; Nathaniel Marchetti, DO; Aditi Satti, MD; Kartik Shenoy, MD; Robert M. Steiner, MD; Alex Swift, MD; Irene Swift, MD; Maria Elena Vega-Sanchez, MD.

University of Alabama, Birmingham, AL: Mark Dransfield, MD; William Bailey, MD; Surya P. Bhatt, MD; Anand Iyer, MD; Hrudaya Nath, MD; J. Michael Wells, MD.

University of California, San Diego, CA: Douglas Conrad, MD; Xavier Soler, MD, PhD; Andrew Yen, MD.

University of Iowa, Iowa City, IA: Alejandro P. Comellas, MD; Karin F. Hoth, PhD; John Newell, Jr., MD; Brad Thompson, MD.

University of Michigan, Ann Arbor, MI: MeiLan K. Han, MD MS; Ella Kazerooni, MD MS; Wassim Labaki, MD MS; Craig Galban, PhD; Dharshan Vummidi, MD. University of Minnesota, Minneapolis, MN: Joanne Billings, MD; Abbie Begnaud, MD; Tadashi Allen, MD.

University of Pittsburgh, Pittsburgh, PA: Frank Sciurba, MD; Jessica Bon, MD; Divay Chandra, MD, MSc; Joel Weissfeld, MD, MPH.

University of Texas Health, San Antonio, San Antonio, TX: Antonio Anzueto, MD; Sandra Adams, MD; Diego Maselli-Caceres, MD; Mario E. Ruiz, MD; Harjinder Singh.

Consent for publication

Not applicable.

Competing interests

KV has nothing to disclose. ND has nothing to disclose. IG receives personal funding from Research Foundation Flanders (FWO). MT is CEO and co-founder of ArtiQ but received no payments related to the manuscript. TT has nothing to disclose. JDC received funding from NIH Grant Support R01-089897 and is chairman of the COPD Foundation Board of Directors. EKS received funding from NIH Grant Support and institutional grant support from GlaxoSmithKline and Bayer. BJM received funding from NHLBI, grants from AstraZeneca, Glaxo-SmithKline and Pearl Research and fees from AstraZeneca, GlaxoSmithKline, Sunovion, Verona, Boehringer Ingelheim, Takeda, Third Pole, Phillips and Circasia. He is member of the NHLBI and Spiration Data Safety Monitoring Boards and chairman of Mt. Sinai SOM Data Safety Monitoring Board. ER received a COPDGene grant from NHBLI. RJ received consulting fees from National Jewish Health for an LLC he owns. MDV received funding from the AI in Flanders project. WJ received grants from AstraZeneca and Chiesi and obtained fees from AstraZeneca, Chiesi and GlaxoSmithKline. He is chairman of Board of Flemish Society for TBC prevention and board member of ArtiQ.

Received: 16 June 2022 Accepted: 6 January 2023 Published online: 19 January 2023

References

- Mead J. The Lung's Quiet Zone. N Engl J Med. 1970;282(23):1318–9. https://doi.org/10.1056/nejm197006042822311.
- Hogg JC, McDonough JE, Suzuki M. Small airway obstruction in COPD: New insights based on micro-CT imaging and MRI imaging. Chest. 2013;143(5):1436–43. https://doi.org/10.1378/chest.12-1766.
- Vestbo J, Anderson JA, Brook RD, et al. Fluticasone furoate and vilanterol and survival in chronic obstructive pulmonary disease with heightened cardiovascular risk (SUMMIT): A double-blind randomised controlled trial. Lancet. 2016;387(10030):1817–26. https://doi.org/10.1016/S0140-6736(16)30069-1.
- Pantel J, Schröder J, Essig M, et al. Hirnvolumetrische befunde bei der spatdepression. Eine untersuchung mittels quantitativer magnetresonanztomographie. Nervenarzt. 1998;69(11):968–74. https://doi.org/10.1007/ s001150050371.
- McFadden ER, Linden DA. A reduction in maximum mid-expiratory flow rate. A spirographic manifestation of small airway disease. Am J Med. 1972;52(6):725–37. https://doi.org/10.1016/0002-9343(72)90078-2.
- Cosio M, Ghezzo H, Hogg JC, et al. The relations between structural changes in small airways and pulmonary-function tests. N Engl J Med. 1978;298(23):1277–81. https://doi.org/10.1056/nejm197806082982303.
- Regan EA, Lynch DA, Curran-Everett D, et al. Clinical and radiologic disease in smokers with normal spirometry. JAMA Intern Med. 2015;175(9):1539–49. https://doi.org/10.1001/jamainternmed.2015.2735.
- Hankinson JL, Odencrantz JR, Fedan KB. Spirometric reference values from a sample of the general US Population. Am J Respir Crit Care Med. 1999;159(1):179–87. https://doi.org/10.1164/ajrccm.159.1.9712108.
- Quanjer PH, Stanojevic S, Cole TJ, et al. Multi-ethnic reference values for spirometry for the 3–95-yr age range: The global lung function 2012 equations. Eur Respir J. 2012;40(6):1324–43. https://doi.org/10.1183/ 09031936.00080312.
- Dayman H. Mechanics of airflow in health and in emphysema. J Clin Invest. 1951;30(11):1175–90. https://doi.org/10.1172/JCl102537.
- Saltzman HP, Ciulla EM, Kuperman AS. The spirographic "kink". A sign of emphysema. Chest. 1976;69(1):51–5. https://doi.org/10.1378/chest.69.1. 51.
- Topalovic M, Exadaktylos V, Peeters A, et al. Computer quantification of airway collapse on forced expiration to predict the presence of emphysema. Respir Res. 2013. https://doi.org/10.1186/1465-9921-14-131.
- Dominelli PB, Foster GE, Guenette JA, et al. Quantifying the shape of the maximal expiratory flow-volume curve in mild COPD. Respir Physiol Neurobiol. 2015;219:30–5. https://doi.org/10.1016/j.resp.2015.08.002.
- Bhatt SP, Bhakta NR, Wilson CG, et al. New spirometry indices for detecting mild airflow obstruction. Sci Rep. 2018. https://doi.org/10.1038/ s41598-018-35930-2.
- Bhatt SP, Bodduluri S, Raghav V, et al. The peak index: Spirometry metric for airflow obstruction severity and heterogeneity. Ann Am Thorac Soc. 2019;16(8):982–9. https://doi.org/10.1513/AnnalsATS.201811-812OC.
- Hoesterey D, Das N, Janssens W, et al. Spirometric indices of early airflow impairment in individuals at risk of developing COPD: Spirometry beyond FEV1/FVC. Respir Med. 2019;156:58–68. https://doi.org/10.1016/j.rmed. 2019.08.004.
- Regan EA, Hokanson JE, Murphy JR, et al. Genetic epidemiology of COPD (COPDGene) study design. COPD J Chronic Obstr Pulm Dis. 2010;7(1):32– 43. https://doi.org/10.3109/15412550903499522.
- Miller MR, Hankinson J, Brusasco V, et al. Standardisation of spirometry. Eur Respir J. 2005;26(2):319–38. https://doi.org/10.1183/09031936.05. 00034805.
- Galbán CJ, Han MK, Boes JL, et al. Computed tomography-based biomarker provides unique signature for diagnosis of COPD phenotypes and disease progression. Nat Med. 2012;18(11):1711–5. https://doi.org/10. 1038/nm.2971.
- Pompe E, Galbán CJ, Ross BD, et al. Parametric response mapping on chest computed tomography associates with clinical and functional parameters in chronic obstructive pulmonary disease. Respir Med. 2017;123:48–55. https://doi.org/10.1016/j.rmed.2016.11.021.

- Topalovic M, Janssens W. Quantifying the shape of the maximal expiratory flow-volume curve to address flow limitation. Respir Physiol Neurobiol. 2016;227:69. https://doi.org/10.1016/j.resp.2015.12.002.
- Das N, Topalovic M, Aerts JM, Janssens W. Area under the forced expiratory flow-volume loop in spirometry indicates severe hyperinflation in COPD patients. Int J COPD. 2019;14:409–18. https://doi.org/10.2147/ COPD.\$185931.
- Mochizuki F, lijima H, Watanabe A, et al. The concavity of the maximal expiratory flow-volume curve reflects the extent of emphysema in obstructive lung diseases. Sci Rep. 2019. https://doi.org/10.1038/s41598-019-49591-2.
- Jankowich M, Elston B, Liu Q, et al. Restrictive spirometry pattern, cardiac structure and function, and incident heart failure in African Americans. The Jackson heart study. Ann Am Thorac Soc. 2018;15(10):1186–96. https://doi.org/10.1513/AnnalsATS.201803-184OC.
- Stringer WW, Porszasz J, Bhatt SP, McCormack MC, Make BJ, Casaburi R. Physiologic insights from the COPD genetic epidemiology study. Chronic Obstr Pulm Dis. 2019;6(3):256–66. https://doi.org/10.15326/jcopdf.6.3. 2019.0128.
- Celli BR, Wedzicha JA. Update on clinical aspects of chronic obstructive pulmonary disease. N Engl J Med. 2019;381(13):1257–66. https://doi.org/ 10.1056/nejmra1900500.
- Koo HK, Vasilescu DM, Booth S, et al. Small airways disease in mild and moderate chronic obstructive pulmonary disease: a cross-sectional study. Lancet Respir Med. 2018;6(8):591–602. https://doi.org/10.1016/S2213-2600(18)30196-6.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

