RESEARCH



Open Access

Diacylglycerol regulates acute hypoxic pulmonary vasoconstriction via TRPC6

Beate Fuchs¹, Markus Rupp¹, Hossein A Ghofrani¹, Ralph T Schermuly^{1,2}, Werner Seeger^{1,2}, Friedrich Grimminger³, Thomas Gudermann⁴, Alexander Dietrich⁴, Norbert Weissmann^{1*}

Abstract

Background: Hypoxic pulmonary vasoconstriction (HPV) is an essential mechanism of the lung that matches blood perfusion to alveolar ventilation to optimize gas exchange. Recently we have demonstrated that acute but not sustained HPV is critically dependent on the classical transient receptor potential 6 (TRPC6) channel. However, the mechanism of TRPC6 activation during acute HPV remains elusive. We hypothesize that a diacylglycerol (DAG)-dependent activation of TRPC6 regulates acute HPV.

Methods: We investigated the effect of the DAG analog 1-oleoyl-2-acetyl-sn-glycerol (OAG) on normoxic vascular tone in isolated perfused and ventilated mouse lungs from TRPC6-deficient and wild-type mice. Moreover, the effects of OAG, the DAG kinase inhibitor R59949 and the phospholipase C inhibitor U73122 on the strength of HPV were investigated compared to those on non-hypoxia-induced vasoconstriction elicited by the thromboxane mimeticum U46619.

Results: OAG increased normoxic vascular tone in lungs from wild-type mice, but not in lungs from TRPC6deficient mice. Under conditions of repetitive hypoxic ventilation, OAG as well as R59949 dose-dependently attenuated the strength of acute HPV whereas U46619-induced vasoconstrictions were not reduced. Like OAG, R59949 mimicked HPV, since it induced a dose-dependent vasoconstriction during normoxic ventilation. In contrast, U73122, a blocker of DAG synthesis, inhibited acute HPV whereas U73343, the inactive form of U73122, had no effect on HPV.

Conclusion: These findings support the conclusion that the TRPC6-dependency of acute HPV is induced via DAG.

Introduction

Hypoxic pulmonary vasoconstriction (HPV) is an essential mechanism in the lung matching blood perfusion to alveolar ventilation, thus optimising gas exchange [1]. Despite decades of research, the signaling pathway underlying HPV has still not been fully resolved. An increase in intracellular calcium concentration ($[Ca^{2+}]_i$) is an essential component in this process, leading to the contraction of precapillary pulmonary arteries [2-4]. However, how $[Ca^{2+}]_i$ is regulated in HPV is still a matter of debate [2,3,5,6]. In addition to L-type voltageoperated calcium channels (VOCC), non-selective transient receptor potential (TRP) channels have been suggested as important regulators of vascular tone in hypoxia [7-9]. In mammals, the family of TRP channels comprises 6 subfamilies, based on their sequence homology [10]. Among these, classical TRPC proteins are expressed in pulmonary arterial smooth muscle [4,5,9], specifically, in smooth muscle cells of distal pulmonary arteries [11], which are suggested to be O_2 sensor and effector cells of acute HPV [12]. Focusing on these aspects, the transient receptor potential channel (TRPC) 6 has recently been identified to be essential for acute but not sustained HPV in mice [9]. In this regard it is important to mention that HPV has repeatedly been shown to consist of two phases. An acute phase occurring within several minutes and a sustained phase developing within more than 30 min of hypoxic ventilation [12-17]. TRPC6 belongs to the TRPC3/6/7 subfamily of TRP channels which can be activated by diacylglycerol (DAG) [18], independently of protein



© 2011 Fuchs et al; licensee BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

^{*} Correspondence: norbert.weissmann@innere.med.uni-giessen.de

¹Excellence Cluster Cardio-Pulmonary System, University of Giessen Lung Center, Dept. of Internal Medicine II, Justus-Liebig-University Giessen, Giessen, Germany

Full list of author information is available at the end of the article

kinase C [19,20]. Hypoxia induces an accumulation of DAG in isolated pulmonary artery smooth muscle cells (PASMC) [9].

We therefore hypothesize that DAG contributes also to the regulation of acute HPV and that the DAG signaling pathway involves TRPC6 in intact lungs.

DAG synthesis results from activation of G-protein coupled receptors or receptor tyrosine kinases and subsequent activation of phospholipase C isoforms (PLC β or PLC γ) leading to hydrolysis of phosphatidylinositol 4,5bisphosphate (PIP₂) [21]. The degradation of DAG is catalyzed by DAG kinases to phosphatidic acid (PA) [22].

Materials and methods

Animals

All animal experiments were approved by the local authorities. Adult C57/BL6 mice were obtained from Charles River Laboratories (Sulzfeld, Germany). TRPC6 deficient (TRPC6^{-/-}) mice were generated as described previously [7]. Respective wild-type (WT) littermates from this colony were used as controls.

Isolated lung perfusion and ventilation

The model of isolated, perfused mouse lungs has been described previously [23]. Briefly, lungs were excised under deep anesthesia, perfused with Krebs-Henseleit buffer (pH 7.37-7.40) in a recirculating system, and ventilated with a mixture of 21% O_2 , 5.3% CO_2 and the balance N_2 (normoxic ventilation). The pressure in the pulmonary artery and in the left atrium was measured by small diameter catheters.

Induction of acute vasoconstriction

Repetitive hypoxic maneuvers of 10-minute duration interrupted by 15-min periods of normoxia were performed. The effects of the various pharmacological agents on pressure responses provoked by alveolar hypoxia (1% O_2 , 5.3% CO_2 and the balance N_2) were determined within such a sequence of repetitive hypoxic maneuvers. 1% O_2 was chosen for hypoxic ventilation as this degree of hypoxia resulted in the most prominent HPV as described before also for other species [24]. Normoxic pulmonary arterial pressure was quantified directly before each hypoxic ventilation maneuver.

To evaluate the specific role of the agents applied in HPV intrinsic pathways, the effect on hypoxia-independent vasoconstriction was determined, using the thromboxane mimetic U46619 [25]. In these experiments, the ventilation remained normoxic, and the lungs were challenged with 4.5 nM U46619 (Paesel and Lorei, Duisburg, Germany; 5.7 mM stock solution in DMSO), applied as a bolus into the pulmonary arterial line. Such bolus applications were repeated every 25 min. The experiments with TRPC6^{-/-} mice as well as their WT controls were performed during continuous normoxic ventilation.

Application of the agents

After the second hypoxic maneuver, the DAG analog 1-oleoyl-2-acetyl-sn-glycerol (OAG), the DAG kinase inhibitor R59949, the PLC inhibitor U73122 or its inactive form (U73343) were applied into the recirculating perfusion medium 10 min prior to the next hypoxic or U46619 challenge with a stepwise increasing dose. For application stock solutions of OAG (50 mM; Sigma-Aldrich, Steinheim, Germany) and R59949 (100 mM; Calbiochem, Bad Soden, Germany) were prepared in DMSO (Merck, Darmstadt, Germany). U73122 and U73343 were dissolved in 96% Ethanol (Fischer, Saarbrücken, Germany) with a stock concentration of 1 mM.

Control experiments were performed with the application of the respective solvents alone.

The effect of the DAG kinase inhibitor R59949 on sustained HPV

To investigate the effect of the DAG kinase inhibitor R59949 on sustained HPV lungs were ventilated for 120 min with a hypoxic gas mixture (1% O_2 , 5.3% CO_2 and the balance N_2) and R59949 was added to the perfusate 10 min prior to the onset of hypoxia. R59949 was applied in a concentration (10 μ M) that inhibited acute HPV by approximately 50%. To characterize a possible sustained effect of R59949 on normoxic vascular tone this agent was applied in an analog schedule but during normoxic ventilation. Control experiments were carried out with the application of the solvent only.

Analysis

The strength of acute HPV is given as the maximum increase in pulmonary arterial pressure (Δ PAP), referenced to the second hypoxic challenge (set at 100%). Changes of the normoxic pulmonary arterial pressure (Δ PAP) in these experiments were referenced to the normoxic PAP directly before the second hypoxic maneuver. The U46619 and normoxic experiments are displayed accordingly. For sustained HPV the increase in pulmonary arterial pressure is given. All values are expressed as means ± SEM (standard error of the mean). Statistical analysis was performed using ANOVA with the Student-Newman-Keuls *post hoc* test or Student's *t*-test with Welsh's correction as appropriate. A p-value < 0.05 was considered significant.

Results

OAG-induced activation of TRPC6 under normoxic conditions To decipher a possible role for DAG in TRPC6 activation, the effect of the membrane-permeable analog of DAG, OAG, on pulmonary artery pressure was investigated in TRPC6^{-/-} and WT mice. As illustrated in Figure 1, OAG increased PAP dose-dependently only in WT mice but not in TRPC6^{-/-} mice. PAP measured after the initial steady state period was 9.9 \pm 0.1 mmHg (n = 9) and was not different between WT and TRPC6^{-/-} mice.

To address the role played by DAG in acute HPV, the effect of the DAG analog OAG as well as of inhibitors of both PLC and DAG kinase on HPV and normoxic vascular tone was assessed.

The effect of OAG on acute hypoxia-induced vasoconstriction

In experiments with repetitive hypoxic ventilation maneuvers, normoxic PAP, assessed prior to each repetitive hypoxic challenge, was dose-dependently increased by OAG (Figure 2A). In parallel with this increase, the strength of HPV was diminished (Figure 2B). This effect was specific for HPV, since vasoconstriction induced by the thromboxane mimetic U46619 was not suppressed, but was rather increased by OAG (Figure 2C).

PAP measured after the initial steady state period, prior to the first vasoconstrictor provocation was $9.9 \pm 0.1 \text{ mmHg}$ (n = 20), and was not different between experiments with OAG application and solvent alone. Absolute Δ PAP values for the strength of HPV and U46619-induced vasoconstrictions were calculated at 1.6 \pm 0.2 mmHg (n = 10) and 2.0 \pm 0.2 mmHg (n = 10).

The effect of the DAG kinase inhibitor R59949 on acute hypoxia-induced vasoconstriction

As DAG is degraded by DAG kinases [22,26], the effect of the DAG kinase inhibitor R59949 on normoxic PAP,



the strength of HPV, as well as U46619-induced vasoconstrictions was investigated. As seen for OAG, the DAG kinase inhibitor dose-dependently increased normoxic PAP (Figure 3A). In parallel, the strength of HPV was reduced by R59949 and was completely abolished at 25 μ M (Figure 3B). The inhibitory effect was specific for HPV, since R59949 caused no significant inhibition of U46619-induced vasoconstrictions, but amplification at higher concentrations (Figure 3C).

The effect of the PLC inhibitor U73122 on acute hypoxiainduced vasoconstriction

As PLC catalyzes the production of DAG [27], the impact of PLC on HPV was investigated by application of the PLC inhibitor U73122. As expected, U73122 did not significantly alter normoxic PAP. The same was true for control experiments with the inactive form of the PLC inhibitor (U73343) or the solvent of the PLC inhibitor (Figure 4A). In contrast, U73122 dose-dependently inhibited the strength of HPV (Figure 4B). However, U46619-induced vasoconstrictions were also diminished (Figure 4C). These effects were absent in experiments with the inactive compound, or the solvent of the PLC inhibitor applied alone.

The effect of the DAG kinase inhibitor R59949 on sustained $\ensuremath{\mathsf{HPV}}$

A single application of 10 μ M R59949, a concentration that inhibited acute HPV by approximately 50%, resulted in a transient vasoconstriction under normoxic ventilation. The maximum increase in PAP occurred after 18 ± 2 min (Figure 5a). Although this dosage still inhibited acute HPV when hypoxic ventilation was performed 120 min after a single application of R59949 (data not given), no sustained elevation of PAP was noted during continuous normoxic ventilation: significant differences in PAP in comparison to the solvent control could be detected for time points \leq 70 min only (Figure 5a). If R59949 was applied prior to a 120 min period of hypoxic ventilation, only the acute but not the sustained phase of HPV was inhibited (Figure 5b).

Discussion

The major finding of this study is that the initiation of acute HPV occurs via a DAG-mediated activation of TRPC6 in mice.

Regarding the vascular effects of alveolar hypoxia, it has previously been shown that these can be divided into three phases, one occurring within seconds, a second developing upon hypoxic ventilation of more than 20 min, and a third which includes a vascular remodeling process, permanently decreasing the area of the vascular lumen [12]. Although some heterogeneity as to the kinetics of the first two phases has been described



Figure 2 1-oleoyl-2-acetyl-sn-glycerol (OAG) diminished HPV specifically. The effect of OAG as well as its solvent on the normoxic vascular tone, the strength of HPV, and the strength of U46619-induced vasoconstriction was investigated in isolated wild-type mouse lungs. (A) Increase in normoxic pulmonary arterial pressure (Δ PAP), assessed directly before each hypoxic ventilation maneuver. (B) Strength of HPV referenced to the effect of the second hypoxic ventilation maneuver (= 100%). (C) Strength of the U46619-induced vasconstriction, referenced to the effect of the second U46619 application (= 100%). Data are derived from n = 5 isolated lung preparations each. * Significant differences compared to control experiments with application of the solvent only (p < 0.05).







(U73122) and inactive (U73343) form of phospholipase C inhibitor as well as their solvent on the normoxic vascular tone, the strength of HPV, and the strength of U46619-induced vasoconstriction was investigated in isolated wild-type mouse lungs. (A) Increase in normoxic pulmonary arterial pressure (Δ PAP), assessed directly before each hypoxic ventilation maneuver. (B) Strength of HPV referenced to the effect of the second hypoxic ventilation maneuver (= 100%). (C) Strength of U46619-induced vasoconstrictions, referenced to the effect of the second U46619 application (= 100%). Data are derived from n = 5 isolated lung preparations each. * Significant differences compared to control experiments with application of the solvent only (p < 0.05).



[26,28-31], it was recently demonstrated that TRPC6 is essential for the acute phase, but not the sustained or chronic vascular effects in mice [9]. However, it has not yet been clarified which pathway controls TRPC6 in acute HPV. TRPC6 is a member of the DAG-sensitive TRPC3/6/7 subfamily, which has been shown to increase $[Ca^{2+}]_i$ in a membrane-delimited fashion, independently of protein kinase C [32]. TRPC6 is insensitive to activation by inositol 1,4,5-trisphosphate at the cellular level [33]. As to the DAG-sensitive nature of TRPC6, it was hypothesized that an increase in DAG induces HPV via this channel. This hypothesis was derived from previous findings in isolated PASMC that 1) hypoxia causes an accumulation of DAG at the cell membrane and that 2) the DAG kinase inhibitor causes an increase in $[Ca^{2+}]_i$ in these cells. In this study, it is documented for the first time in intact lungs that the membrane-permeable analog of DAG, OAG, induces a vasoconstriction in WT mice which is mediated via TRPC6. This can be concluded from the observation that the

DAG-induced vasoconstriction was absent in TRPC6^{-/-} mice, although it has been previously demonstrated that TRPC6^{-/-} mice do not lack non-hypoxia-induced vaso- constrictor responses [9]. The latter finding excludes a general lack of vasocontractility in TRPC6-/- mice.

The central role of DAG in acute HPV was further supported by the effects of the DAG analog OAG, as well as the DAG kinase inhibitor and the phospholipase C inhibitor. Along these lines, it has been shown that DAG accumulation can be caused by inhibition of DAG kinases [34] but also by activation of PLC [35].

OAG did not only induce a vasoconstrictor response under normoxic conditions but also inhibited the subsequent acute HPV during hypoxia. Consistent with this idea, an increase in cellular DAG levels by the application of DAG kinase inhibitor [34] increased vasoconstriction, but decreased the subsequent acute HPV. Thus, both OAG and R59949 interfere with the same signal transduction cascade inducing acute HPV and therefore mimic this physiological response under normoxic conditions. The fact that low doses of OAG slightly increased the strength of HPV maybe is induced by the supplementary effect of the exogenous OAG in addition to the endogenous DAG produced by acute hypoxia. The inhibition of HPV caused by OAG or R59949 was specific for HPV as non-hypoxia-induced vasoconstriction induced by U46619 was not inhibited by either agent.

The notion that OAG-induced vasoconstrictions under normoxia were somewhat higher than the strength of HPV indicates, as expected, that DAG not only specifically mediates hypoxia-induced vasoconstrictions but also contributes to non-hypoxia-induced vasoconstrictions as previously shown for G protein coupled receptors [36].

In contrast to those agents which mimic acute HPV or increase DAG levels, the PLC inhibitor U73122 should decrease intracellular DAG production [37] and thus should suppress but not mimic HPV. As expected, while not mimicking acute HPV, U73122 suppressed HPV. It is unlikely that this effect is due to an unspecific inhibition of other signaling processes, because the inactive but structurally analogous - compound U73343 had no effect on acute HPV. The fact, that U46619-induced vasoconstrictions were also suppressed by the PLC but not the DAG kinase inhibitor, supports the notion that the thromboxane mimetic-induced vasoconstriction is dependent on PLC without triggering the DAG-TRPC6axis. In this regard it was shown that activation of thromboxane receptors can contribute to contraction of bovine pulmonary arteries by depletion of intracellular calcium stores and calcium entry via store-operated calcium channels [38]. Besides a role of store-operated calcium channels, U46619-induced contraction was also shown to be dependent at least in part on calcium entry through VOCC [35]. In this process, ROS-induced PKC zeta activation inhibited voltage-gated potassium (Kv) channel activity leading to membrane depolarization and activation of VOCC [36,37]. Moreover, PKC can generally also be activated by DAG [38]. Triggering different pathways, cell-compartmentalization, as well as some synergism of the above pathways may explain the increase of U46619-induced vasoconstrictions after application of the DAG analog OAG. Thus, the enhancement of U46619-induced vasoconstrictions, in contrast to the inhibition of HPV, further supports the specific role of DAG in acute HPV signaling. A compartmentalized effect of DAG regulation in HPV is further supported by the fact that the DAG kinase inhibitor R59449 did not amplify U46619-induced but selectively inhibited HPV.

The fact that the DAG kinase inhibitor R59449 inhibited only acute HPV but not sustained HPV and mimicked only acute but not sustained HPV during normoxic ventilation is well in line with our previous finding that DAG-regulated TRPC6 channels are essential for acute but not sustained HPV.

With regard to previous findings that acute HPV may be regulated by reactive oxygen species [39-43], we speculate that DAG levels activating TRPC6 can be increased by a redox-dependent mechanism. This can hypothetically be caused by a redox-dependent modulation of e.g. DAG kinase activity. In addition to TRPC6, it was shown that Kv channels are essential for acute HPV [44,45]. Although our current study did not investigate a possible effect of the DAG-TRPC6-axis on the closure of Kv channels in HPV, these two systems may be linked via modulation of the cellular sodium concentration as previously suggested [46]. This concept would be in line with the lack of acute HPV recently described in malonyl-CoA decarboxylase deficient mice, which lack hypoxic mitochondrial ROS signaling and Kv channel inhibition [40].

The specific role of DAG in mediating acute HPV shown in our isolated lung experiments (where the endothelium is present) and in our previous study [9] in isolated PASMC (where the increase in $[Ca^{2+}]_i$ was used as a readout for HPV) indicates that the oxygen sensing process underlying HPV resides in the PASMC. This is well in line with numerous previous findings [47-50] and the notion expressed by others that acute HPV (in contrast to sustained) is independent from the endothelium [13,15,17,51]. This, however, does not exclude an important role of the endothelium in modulating HPV, though not contributing to oxygen sensing. This interpretation is well in line with the finding by us and others that isolated PASMC need to be "primed" by e.g. endothelin-1 as a prerequisite for functional oxygen

sensing [9,52-54] and allows the suggestion that a basic stimulation of DAG production by G-protein coupled receptors is a prerequisite for oxygen sensing and signal transduction of acute HPV, occurring cell compartment-specific by DAG kinase inhibition.

Conclusions

To summarize, DAG was identified as an important mediator in the signaling pathway underlying acute HPV. Moreover, these data indicate that DAG activates TRPC6 as an essential step in the mechanism of acute HPV.

Acknowledgements

This work was supported by the Deutsche Forschungsgemeinschaft (DFG DI 514/8-1, DFG WE 1978/4-1), the Excellence Cluster Pulmonary System (ECCPS), and the von-Behring-Röntgen-Stiftung. The authors thank Dr. Rory Morty, Giessen, for linguistic editing of the manuscript.

Author details

¹Excellence Cluster Cardio-Pulmonary System, University of Giessen Lung Center, Dept. of Internal Medicine II, Justus-Liebig-University Giessen, Giessen, Germany. ²Max-Planck-Institute for Heart and Lung Research, Bad Nauheim, Germany. ³Excellence Cluster Cardio-Pulmonary System, University of Giessen Lung Center, Dept. of Internal Medicine IV/V, Justus-Liebig-University Giessen, Giessen, Germany. ⁴Walter-Straub-Institute for Pharmacology and Toxicology, Ludwig-Maximilians-University Munich, Munich, Germany.

Authors' contributions

BF participated in the design, analysis and interpretation of experiments, performed statistical analysis, and designed and drafted the manuscript together with NW and AD. MR performed most of the experiments and has made substantial contributions to analysis and interpretation and statistical analysis of the data. HAG, RTS, WS, and FG participated in the design of the experiments and in the interpretation of the results. TG and AD supplied and/or generated the TRPC6^{-/-} mice and developed the hypothesis together with NW and BF. Their contribution was critical for intellectual content and designed portions of the experiments. NW designed and supervised the study, interpreted the data and drafted the manuscript together with BF and AD. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 13 October 2010 Accepted: 4 February 2011 Published: 4 February 2011

References

- Weissmann N, Sommer N, Schermuly RT, Ghofrani HA, Seeger W, Grimminger F: Oxygen sensors in hypoxic pulmonary vasoconstriction. *Cardiovasc Res* 2006, 71:620-629.
- Weigand L, Foxson J, Wang J, Shimoda LA, Sylvester JT: Inhibition of hypoxic pulmonary vasoconstriction by antagonists of store-operated Ca2+ and nonselective cation channels. *Am J Physiol Lung Cell Mol Physiol* 2005, 289:L5-L13.
- Wang J, Shimoda LA, Weigand L, Wang W, Sun D, Sylvester JT: Acute hypoxia increases intracellular [Ca2+] in pulmonary arterial smooth muscle by enhancing capacitative Ca2+ entry. Am J Physiol Lung Cell Mol Physiol 2005, 288:L1059-L1069.
- McDaniel SS, Platoshyn O, Wang J, Yu Y, Sweeney M, Krick S, Rubin LJ, Yuan JX: Capacitative Ca(2+) entry in agonist-induced pulmonary vasoconstriction. Am J Physiol Lung Cell Mol Physiol 2001, 280:L870-L880.
- Ward JP, Robertson TP, Aaronson PI: Capacitative calcium entry: a central role in hypoxic pulmonary vasoconstriction? Am J Physiol Lung Cell Mol Physiol 2005, 289:L2-L4.

- Aaronson PI, Robertson TP, Knock GA, Becker S, Lewis TH, Snetkov V, Ward JP: Hypoxic pulmonary vasoconstriction: mechanisms and controversies. J Physiol 2006, 570:53-58.
- Dietrich A, Mederos YS, Gollasch M, Gross V, Storch U, Dubrovska G, Obst M, Yildirim E, Salanova B, Kalwa H, Essin K, Pinkenburg O, Luft FC, Gudermann T, Birnbaumer L: Increased vascular smooth muscle contractility in TRPC6-/- mice. Mol Cell Biol 2005, 25:6980-6989.
- Owsianik G, Talavera K, Voets T, Nilius B: Permeation and selectivity of TRP channels. Annu Rev Physiol 2006, 68:685-717.
- Weissmann N, Dietrich A, Fuchs B, Kalwa H, Ay M, Dumitrascu R, Olschewski A, Storch U, Schnitzler M, Ghofrani HA, Schermuly RT, Pinkenburg O, Seeger W, Grimminger F, Gudermann T: Classical transient receptor potential channel 6 (TRPC6) is essential for hypoxic pulmonary vasoconstriction and alveolar gas exchange. *Proc Natl Acad Sci USA* 2006, 103:19093-19098.
- Dietrich A, Chubanov V, Kalwa H, Rost BR, Gudermann T: Cation channels of the transient receptor potential superfamily: their role in physiological and pathophysiological processes of smooth muscle cells. *Pharmacol Ther* 2006, 112:744-760.
- Wang J, Shimoda LA, Sylvester JT: Capacitative calcium entry and TRPC channel proteins are expressed in rat distal pulmonary arterial smooth muscle. Am J Physiol Lung Cell Mol Physiol 2004, 286:L848-L858.
- Weissmann N, Grimminger F, Olschewski A, Seeger W: Hypoxic pulmonary vasoconstriction: a multifactorial response? Am J Physiol Lung Cell Mol Physiol 2001, 281:L314-L317.
- Ward JP, Aaronson PI: Mechanisms of hypoxic pulmonary vasoconstriction: can anyone be right? *Respir Physiol* 1999, 115:261-271.
 Weissmann N, Zeller S, Schafer RU, Turowski C, Ay M, Quanz K, Ghofrani HA, Schermuly RT, Fink L, Seeger W, Grimminger F: Impact of mitochondria
- and NADPH oxidases on acute and sustained hypoxic pulmonary vasoconstriction. *Am J Respir Cell Mol Biol* 2006, 34:505-513.
 Robertson TP, Aaronson PI, Ward JP: Ca2+ sensitization during sustained hypoxic pulmonary vasoconstriction is endothelium dependent. *Am J*
- Physiol Lung Cell Mol Physiol 2003, 284:L1121-L1126.
 Dipp M, Nye PC, Evans AM: Hypoxic release of calcium from the sarcoplasmic reticulum of pulmonary artery smooth muscle. Am J Physiol
- Lung Cell Mol Physiol 2001, 281:L318-L325.
 Aaronson PI, Robertson TP, Ward JP: Endothelium-derived mediators and hypoxic pulmonary vasoconstriction. *Respir Physiol Neurobiol* 2002,
- 132:107-120.
 Dietrich A, Kalwa H, Rost BR, Gudermann T: The diacylgylcerol-sensitive TRPC3/6/7 subfamily of cation channels: functional characterization and
- physiological relevance. *Pflugers Arch* 2005, 451:72-80.
 Helliwell RM, Large WA: Alpha 1-adrenoceptor activation of a non-selective cation current in rabbit portal vein by 1,2-diacyl-sn-glycerol. *J*
- Physiol 1997, 499(Pt 2):417-428.
 Inoue R, Okada T, Onoue H, Hara Y, Shimizu S, Naitoh S, Ito Y, Mori Y: The transient receptor potential protein homologue TRP6 is the essential component of vascular alpha(1)-adrenoceptor-activated Ca(2 +)-permeable cation channel. *Circ Res* 2001, 88:325-332.
- Freichel M, Vennekens R, Olausson J, Hoffmann M, Muller C, Stolz S, Scheunemann J, Weissgerber P, Flockerzi V: Functional role of TRPC proteins in vivo: lessons from TRPC-deficient mouse models. *Biochem Biophys Res Commun* 2004, 322:1352-1358.
- Merida I, vila-Flores A, Merino E: Diacylglycerol kinases: at the hub of cell signalling. Biochem J 2008, 409:1-18.
- Weissmann N, Akkayagil E, Quanz K, Schermuly RT, Ghofrani HA, Fink L, Hanze J, Rose F, Seeger W, Grimminger F: Basic features of hypoxic pulmonary vasoconstriction in mice. *Respir Physiol Neurobiol* 2004, 139:191-202.
- Weissmann N, Grimminger F, Walmrath D, Seeger W: Hypoxic vasoconstriction in buffer-perfused rabbit lungs. *Respir Physiol* 1995, 100:159-169.
- Bolla M, You D, Loufrani L, Levy BI, Levy-Toledano S, Habib A, Henrion D: Cyclooxygenase involvement in thromboxane-dependent contraction in rat mesenteric resistance arteries. *Hypertension* 2004, 43:1264-1269.
- Bennie RE, Packer CS, Powell DR, Jin N, Rhoades RA: Biphasic contractile response of pulmonary artery to hypoxia. Am J Physiol 1991, 261:L156-L163.
- Rebecchi MJ, Pentyala SN: Structure, function, and control of phosphoinositide-specific phospholipase C. *Physiol Rev* 2000, 80:1291-1335.

- Domino KB, Chen L, Alexander CM, Williams JJ, Marshall C, Marshall BE: Time course and responses of sustained hypoxic pulmonary vasoconstriction in the dog. *Anesthesiology* 1984, 60:562-566.
- Welling KL, Sanchez R, Ravn JB, Larsen B, Amtorp O: Effect of prolonged alveolar hypoxia on pulmonary arterial pressure and segmental vascular resistance. J Appl Physiol 1993, 75:1194-1200.
- Woodmansey PA, Zhang F, Channer KS, Morice AH: Effect of the calcium antagonist amlodipine on the two phases of hypoxic pulmonary vasoconstriction in rat large and small isolated pulmonary arteries. J Cardiovasc Pharmacol 1995, 25:324-329.
- Ozaki M, Marshall C, Amaki Y, Marshall BE: Role of wall tension in hypoxic responses of isolated rat pulmonary arteries. Am J Physiol 1998, 275: L1069-L1077.
- Hofmann T, Obukhov AG, Schaefer M, Harteneck C, Gudermann T, Schultz G: Direct activation of human TRPC6 and TRPC3 channels by diacylglycerol. *Nature* 1999, 397:259-263.
- Trebak M, Vazquez G, Bird GS, Putney JW Jr: The TRPC3/6/7 subfamily of cation channels. *Cell Calcium* 2003, 33:451-461.
- Friedlander G, Le GC, Sraer J, Amiel C: 12-HETE modulates Na-coupled uptakes in proximal tubular cells: role of diacylglycerol kinase inhibition. *Am J Physiol* 1990, 259:F816-F822.
- Patterson RL, van Rossum DB, Nikolaidis N, Gill DL, Snyder SH: Phospholipase C-gamma: diverse roles in receptor-mediated calcium signaling. Trends Biochem Sci 2005, 30:688-697.
- Wynne BM, Chiao CW, Webb RC: Vascular Smooth Muscle Cell Signaling Mechanisms for Contraction to Angiotensin II and Endothelin-1. J Am Soc Hypertens 2009, 3:84-95.
- Thompson AK, Mostafapour SP, Denlinger LC, Bleasdale JE, Fisher SK: The aminosteroid U-73122 inhibits muscarinic receptor sequestration and phosphoinositide hydrolysis in SK-N-SH neuroblastoma cells. A role for Gp in receptor compartmentation. J Biol Chem 1991, 266:23856-23862.
- Alapati VR, McKenzie C, Blair A, Kenny D, MacDonald A, Shaw AM: Mechanisms of U46619- and 5-HT-induced contraction of bovine pulmonary arteries: role of chloride ions. *Br J Pharmacol* 2007, 151:1224-1234.
- Archer SL, Reeve HL, Michelakis E, Puttagunta L, Waite R, Nelson DP, Dinauer MC, Weir EK: O2 sensing is preserved in mice lacking the gp91 phox subunit of NADPH oxidase. Proc Natl Acad Sci USA 1999, 96:7944-7949.
- Sutendra G, Bonnet S, Rochefort G, Haromy A, Folmes KD, Lopaschuk GD, Dyck JR, Michelakis ED: Fatty acid oxidation and malonyl-CoA decarboxylase in the vascular remodeling of pulmonary hypertension. *Sci Transl Med* 2010, 2:44ra58.
- Fuchs B, Sommer N, Dietrich A, Schermuly RT, Ghofrani HA, Grimminger F, Seeger W, Gudermann T, Weissmann N: Redox signaling and reactive oxygen species in hypoxic pulmonary vasoconstriction. *Respir Physiol Neurobiol* 2010, 174:282-291.
- Sommer N, Dietrich A, Schermuly RT, Ghofrani HA, Gudermann T, Schulz R, Seeger W, Grimminger F, Weissmann N: Regulation of hypoxic pulmonary vasoconstriction: basic mechanisms. *Eur Respir J* 2008, 32:1639-1651.
- 43. Waypa GB, Schumacker PT: Hypoxic pulmonary vasoconstriction: redox events in oxygen sensing. J Appl Physiol 2005, 98:404-414.
- Archer SL, London B, Hampl V, Wu X, Nsair A, Puttagunta L, Hashimoto K, Waite RE, Michelakis ED: Impairment of hypoxic pulmonary vasoconstriction in mice lacking the voltage-gated potassium channel Kv1.5. FASEB J 2001, 15:1801-1803.
- Archer SL, Souil E, nh-Xuan AT, Schremmer B, Mercier JC, El YA, Nguyen-Huu L, Reeve HL, Hampl V: Molecular identification of the role of voltagegated K+ channels, Kv1.5 and Kv2.1, in hypoxic pulmonary vasoconstriction and control of resting membrane potential in rat pulmonary artery myocytes. J Clin Invest 1998, 101:2319-2330.
- Dietrich A, Kalwa H, Fuchs B, Grimminger F, Weissmann N, Gudermann T: In vivo TRPC functions in the cardiopulmonary vasculature. *Cell Calcium* 2007, 42:233-244.
- Cornfield DN, Stevens T, McMurtry IF, Abman SH, Rodman DM: Acute hypoxia increases cytosolic calcium in fetal pulmonary artery smooth muscle cells. Am J Physiol 1993, 265:L53-L56.
- Salvaterra CG, Goldman WF: Acute hypoxia increases cytosolic calcium in cultured pulmonary arterial myocytes. Am J Physiol 1993, 264:L323-L328.

- Vadula MS, Kleinman JG, Madden JA: Effect of hypoxia and norepinephrine on cytoplasmic free Ca2+ in pulmonary and cerebral arterial myocytes. *Am J Physiol* 1993, 265:L591-L597.
- Zhang F, Carson RC, Zhang H, Gibson G, Thomas HM III: Pulmonary artery smooth muscle cell [Ca2+]i and contraction: responses to diphenyleneiodonium and hypoxia. *Am J Physiol* 1997, 273:L603-L611.
- Ward JP, Robertson TP: The role of the endothelium in hypoxic pulmonary vasoconstriction. *Exp Physiol* 1995, 80:793-801.
- Waypa GB, Chandel NS, Schumacker PT: Model for hypoxic pulmonary vasoconstriction involving mitochondrial oxygen sensing. *Circ Res* 2001, 88:1259-1266.
- Sham JS, Crenshaw BR, Deng LH, Shimoda LA, Sylvester JT: Effects of hypoxia in porcine pulmonary arterial myocytes: roles of K(V) channel and endothelin-1. Am J Physiol Lung Cell Mol Physiol 2000, 279:L262-L272.
- Leach RM, Hill HM, Snetkov VA, Robertson TP, Ward JP: Divergent roles of glycolysis and the mitochondrial electron transport chain in hypoxic pulmonary vasoconstriction of the rat: identity of the hypoxic sensor. J Physiol 2001, 536:211-224.

doi:10.1186/1465-9921-12-20

Cite this article as: Fuchs et al.: Diacylglycerol regulates acute hypoxic pulmonary vasoconstriction via TRPC6. Respiratory Research 2011 12:20.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

) BioMed Central

Submit your manuscript at www.biomedcentral.com/submit